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Attorney Docket No. 18547-043210US

"Express Mail" Label No. EL623997634US

Date of Deposit: September 1, 2000

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

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BOX PATENT APPLICATION
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Sir:

Transmitted herewith for filing under 37 CFR 1.53(b) is the

- ☐ patent application of
☒ continuation patent application of
☐ divisional patent application of
☐ continuation-in-part patent application of

Inventor(s)/Applicant Identifier: Michael C. Pirrung, Lubert Stryer, Stephen P.A. Fodor and J. Leighton Read

For: **PHOTOLITHOGRAPHIC AND OTHER MEANS FOR MANUFACTURING ARRAYS (AS AMENDED)**

- [X] This application claims priority from each of the following Application Nos./filing dates:
09/557,875, filed April 24, 2000; 09/056,927, filed April 8, 1998; 08/670,118, filed June 25, 1996; 08/168,904, filed December 15, 1993; 07/624,114, filed December 6, 1990; 07/362,901, filed June 7, 1989; 08/348,471, filed November 30, 1994; 07/805,727, filed December 6, 1991 and 07/492,462, filed March 7, 1990
the disclosure(s) of which is (are) incorporated by reference.

- [X] Please amend this application by deleting the first paragraph and substitute therefor: --The present application is a continuation of 09/557,875, filed April 24, 2000, which is a continuation of 09/056,927 filed April 8, 1998, which is a continuation of 08/670,118 filed June 25, 1996, now U.S. Patent No. 5,800,992, which is a divisional of 08/168,904 filed December 15, 1993, which is a continuation of 07/624,114, filed December 6, 1990. The present application is also a continuation-in-part of USSN 07/362,901, filed June 7, 1989. This application is also a continuation-in-part of USSN 08/348,471 filed November 30, 1994, which is a continuation of USSN 07/805,727 filed December 6, 1991 (now U.S. Patent No. 5,424,186), which is a continuation-in-part of USSN 07/492,462, filed March 7, 1990 (now U.S. Patent No. 5,143,854). Additional commonly assigned applications Barrett et al., U.S.S.N. 07/435,316 (caged biotin parent), filed November 13, 1989; and Barrett et al., U.S.S.N. 07/612,671 (caged biotin CIP) filed November 13, 1990 are also incorporated herein by reference. Additional applications Pirrung et al., U.S.S.N. 07/624,120 (now abandoned) a divisional of which has issued as US 5,744,101 and Dower et al., U.S.S.N. 07/626,730 (now US 5,547,839), which are also commonly assigned are also hereby incorporated herein by reference.--

Enclosed are:

- [X] 203 page(s) of specification
[X] 28 page(s) of claims
[X] 1 page of Abstract
[X] 27 sheet(s) of ☐ formal ☒ informal drawing(s).
[X] A ☒ signed ☐ unsigned Declaration and Power of Attorney.
[X] Title Page
[X] Preliminary Amendment
[X] Return postcard

	(Col. 1)	(Col. 2)
FOR:	NO. FILED	NO. EXTRA
BASIC FEE		
TOTAL CLAIMS	38 - 20	= *18
INDEP. CLAIMS	3 - 3	= *0
[] MULTIPLE DEPENDENT CLAIM PRESENTED		

* If the difference in Col. 1 is less than 0, enter "0" in Col. 2.

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+ \$130.00 =	
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OTHER THAN SMALL ENTITY

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x \$18.00 =	\$324.00
x \$78.00 =	\$0.00
+ \$260.00 =	
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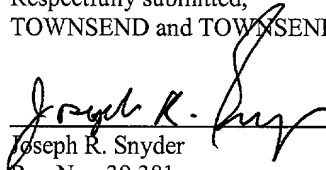
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Respectfully submitted,
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PATENT

Attorney Docket No.: 018547-043210US

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

By: _____

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

Michael C. Pirrung *et al.*

Application No.: Not yet Assigned

Filed: Herewith

For: PHOTOLITHOGRAPHIC AND
OTHER MEANS FOR
MANUFACTURING ARRAYS (AS
AMENDED)

Examiner: Not Yet Assigned

Art Unit: Not Yet Assigned

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to examination of the above-referenced application and calculation of the filing fee, please enter the following amendments and remarks.

IN THE TITLE:

Please delete the current title and substitute therefor:

-- PHOTOLITHOGRAPHIC AND OTHER MEANS FOR
MANUFACTURING ARRAYS --

IN THE CLAIMS:

Please cancel claims 1-171 without prejudice.

Please add the following new claims 172-209:

- 1 **172.** (New) A method of synthesizing a polypeptide array, wherein said
- 2 array comprises at least two different polypeptides immobilized on a substrate, and wherein
- 3 said polypeptides are synthesized on a surface of said substrate, said method comprising:

(a) contacting said surface with a first protected amino acid wherein said first protected amino acid is selectively coupled to a functional group in a first selectively activated region of said surface;

(b) contacting said surface with a second protected amino acid wherein said second protected amino acid is selectively coupled to a functional group in a second selectively activated region of said surface; and,

(c) repeating the above steps until at least two different polypeptides are formed at known locations on said substrate surface.

173. (New) The method as recited in claim **172**, wherein contacting said surface with said second protected amino acid is accomplished without physical segregation of said surface.

174. (New) The method as recited in claim **172**, wherein said at least two different polypeptides each occupy an area on said substrate of less than about 1 cm^2 to about $1 \times 10^{-5} \text{ cm}^2$.

175. (New) The method as recited in claim **174**, wherein said at least two different polypeptides each occupy an area on said substrate of less than about $1 \times 10^{-1} \text{ cm}^2$ to about $1 \times 10^{-4} \text{ cm}^2$.

176. (New) The method as recited in claim **175**, wherein said at least two different polypeptides each occupy an area on said substrate of less than about $1 \times 10^{-2} \text{ cm}^2$ to about $1 \times 10^{-3} \text{ cm}^2$.

177. (New) The method as recited in claim **172**, repeating said steps above until said at least two different polypeptides exceed a density of about 400 different polypeptides / cm^2 .

178. (New) The method as recited in claim **172**, repeating said steps above until said at least two different polypeptides exceed a density of about 1000 different polypeptides / cm^2 .

1 **179.** (New) The method as recited in claim **172**, wherein said method
2 produces a substrate that contains more than 100 different polypeptides per cm².

1 **180.** (New) The method as recited in claim **172**, wherein said method
2 produces a substrate that contains more than 1,000 different polypeptides per cm².

1 **181.** (New) The method as recited in claim **172**, wherein said method
2 produces a substrate that contains more than 10,000 different polypeptides per cm².

1 **182.** (New) The method as recited in claim **172**, wherein said method
2 produces a substrate that contains more than 100,000 different polypeptides per cm².

1 **183.** (New) A method for synthesizing polypeptides on a substrate, said
2 method comprising:

3 a) providing a substrate wherein said substrate comprises immobilized
4 polypeptide molecules, said polypeptide molecules coupled to a removable protecting
5 groups;

6 b) removing said protecting group from said polypeptide molecules in a
7 first predefined region of said substrate without removing said protecting groups from a
8 second predefined region of said substrate; and

9 c) contacting said substrate with a first amino acid to couple said first
10 amino acid to said polypeptide molecules in said first predefined region, said first amino acid
11 having an amino acid protecting group thereon, forming a first polypeptide on said substrate in
12 said first predefined region that is different from a polypeptide in said second predefined
13 region.

1 **184.** (New) The method as recited in claim **183**, wherein said step of
2 removing is an irradiation step.

1 **185.** (New) The method as recited in claim **184**, wherein said step of
2 irradiating is a step of masking a light source with a mask placed between said light source
3 and said substrate, said mask comprising first transparent regions and second opaque regions,

4 said transparent regions transmitting light from said source to at least said first predefined
5 region, and said opaque regions blocking light from said source to at least said second
6 predefined region.

1 **186.** (New) The method as recited in claim **183**, wherein said first and
2 second regions each have total areas less than about 1 cm^2 .

1 **187.** (New) The method as recited in claim **184**, wherein said steps of
2 irradiating are conducted with a monochromatic light.

1 **188.** (New) The method as recited in claim **184**, wherein said steps of
2 irradiating and contacting are repeated so as to synthesize 10^3 different polypeptides on said
3 substrate.

1 **189.** (New) The method as recited in claim **184**, wherein the step of
2 irradiating a first predefined region is a step of irradiating half of a region of said substrate
3 irradiated in a prior synthesis step, and not irradiating half of said region irradiated in a prior
4 synthesis step.

1 **190.** (New) The method as recited in claim **183**, wherein said steps a) and
2 b) are repeated to synthesize more than 1,000 different polypeptides on different synthesis
3 regions of said substrate, each of said different polypeptides occupying an area of less than
4 about 10^{-2} cm^2 to about $1 \times 10^{-5} \text{ cm}^2$.

1 **191.** (New) The method as recited in claim **190**, wherein said steps a) and b)
2 are repeated to synthesize more than 1,000 different polypeptides on different synthesis
3 regions of said substrate, each of said different polypeptides occupying an area of less than
4 about 10^{-2} cm^2 to about $1 \times 10^{-4} \text{ cm}^2$.

1 **192.** (New) The method as recited in claim **191**, wherein said steps a) and b)
2 are repeated to synthesize more than 1,000 different polypeptides on different synthesis

regions of said substrate, each of said different polypeptides occupying an area of less than about 10^{-2} cm² to about 1×10^{-3} cm².

193. (New) A method of synthesizing polypeptides, said method comprising:

a) generating a pattern of light and dark areas by selectively irradiating at least a first area of a surface of a substrate, said surface comprising immobilized amino acids on said surface, said amino acids coupled to a photoremovable protective group, without irradiating at least a second area of said surface, to remove said protective group from said amino acids in said first area;

b) simultaneously contacting said first area and said second area of said surface with a first amino acid to couple said first amino acid to said immobilized amino acids in said first area, and not in said second area, said first amino acid having said photoremovable protective group;

c) generating another pattern of light and dark areas by selectively irradiating with light at least a part of said first area of said surface and at least a part of said second area to remove said protective group in said at least a part of said first area and said at least a part of said second area;

d) simultaneously contacting said first area and said second area of said surface with a second amino acid to couple said second amino acid to said immobilized amino acids in at least a part of said first area and at least a part of said second area; and

e) performing additional irradiating and amino acid contacting and coupling steps so that a matrix array of at least 100 different polypeptides is formed on said surface, each different polypeptide synthesized in an area of less than 0.1 cm², whereby said different polypeptides have sequences and locations on said surface defined by the patterns of light and dark areas formed during the irradiating steps and the amino acids coupled in said contacting steps.

194. (New) The method as recited in claim **193**, wherein said substrate is selected from the group consisting of Langmuir Blodgett film, glass, germanium, silicon,

(poly)tetrafluorethylene, polystyrene, gallium arsenide, gallium phosphide, silicon oxide, silicon nitride, and combinations thereof.

195. (New) The method as recited in claim **193**, wherein said protective group is selected from the group consisting of 6-nitroveratryloxycarbonyl, 2-nitrobenzyloxy carbonyl, dimethyl dimethoxybenzyloxy carbonyl, 5-bromo-7-nitroindoliny, o-hydroxyalpha-methyl cinnamoyl, 2-oxymethylene anthriquinone, and mixtures thereof.

196. (New) The method as recited in claim **193**, wherein each different polypeptide synthesized is in an area of less than about 0.1 cm^2 to about $1 \times 10^{-5} \text{ cm}^2$.

197. (New) The method as recited in claim **196**, wherein each different polypeptide synthesized is in an area of less than about $1 \times 10^{-1} \text{ cm}^2$ to about $1 \times 10^{-4} \text{ cm}^2$.

198. (New) The method as recited in claim **197**, wherein each different polypeptide synthesized is in an area of less than about $1 \times 10^{-2} \text{ cm}^2$ to about $1 \times 10^{-3} \text{ cm}^2$.

199. (New) The method as recited in claim **193**, wherein said matrix array is at least 400 different polypeptides/ cm^2 .

200. (New) The method as recited in claim **193**, wherein said matrix array is at least 1000 different polypeptides / cm^2 .

201. (New) The method as recited in claim **193**, wherein said method produces a substrate that contains more than 1,000 different polypeptides per 0.1 cm^2 .

202. (New) The method as recited in claim **193**, wherein said method produces a substrate that contains more than 10,000 different polypeptides per 0.1 cm^2 .

203. (New) The method as recited in claim **193**, wherein said method produces a substrate that contains more than 100,000 different polypeptides per 0.1 cm^2 .

204. (New) The method as recited in claim **193**, wherein the irradiating step c) further comprises:

3 i) placing a mask adjacent to said substrate, said mask having
4 substantially transparent regions and substantially opaque regions at a wavelength of light;
5 and

6 ii) illuminating said mask with a light source, said light source producing
7 at least said wavelength of light, said mask permitting illumination of half of said substrate
8 which was illuminated and half of said substrate which was not illuminated in said step a).

1 **205.** (New) The method as recited in claim **204**, wherein said additional
2 steps are performed so as to synthesize 10^3 different polypeptides in 10^3 respective
3 preselected regions on said substrate.

1 **206.** (New) The method as recited in claim **204**, wherein said additional
2 steps are performed so as to synthesize 10^6 different polypeptides in 10^6 respective
3 preselected regions on said substrate.

1 **207.** (New) The method of claim **204**, wherein said additional steps are
2 performed so that at least 1,000 different polypeptides are synthesized on said surface, and
3 each different polypeptide is contained within an area less than about $1 \times 10^{-3} \text{ cm}^2$.

1 **208.** (New) The method of claim **193**, wherein said immobilized amino
2 acids are attached to said surface by a linker selected from the group consisting of aryl
3 acetylene and ethylene glycol.

1 **209.** (New) The method of claim **193**, wherein said immobilized amino
2 acids are attached to said surface by polyethylene glycol.

REMARKS

Claims 172-209 are pending in this application and presented for examination.
Claims 1-171 have been canceled without prejudice or disclaimer. Early examination on the
merits is respectfully requested.

THE APPLICATION

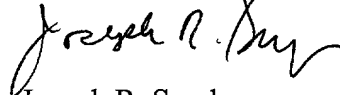
The present application is a continuation of 09/557,875, filed April 24, 2000, which is a continuation of 09/056,927 filed April 8, 1998, which is a continuation of 08/670,118 filed June 25, 1996, now U.S. Patent No. 5,800,992, which is a divisional of 08/168,904 filed December 15, 1993, which is a continuation of 07/624,114, filed December 6, 1990. The present application is also a continuation-in-part of USSN 07/362,901, filed June 7, 1989. This application is also a continuation-in-part of USSN 08/348,471 filed November 30, 1994, which is a continuation of USSN 07/805,727 filed December 6, 1991 (now U.S. Patent No. 5,424,186), which is a continuation-in-part of USSN 07/492,462, filed March 7, 1990 (now U.S. Patent No. 5,143,854).

The specification of the present case differs from that of 07/624,114 (the '114 application") in that portions of certain patent filings cited and incorporated by reference in the '114 application have been reproduced in the present application. Specifically, the incorporated portions are from US Application Serial Nos.: 07/362,901, 07/492,462 and 07/624,120. These applications are all cited in the cross-reference to related applications in the '114 application. The incorporated portions provide further details of methods for array synthesis. Figures from incorporated text have been renumbered for conformity with existing text. The cross-reference to related applications in the present application has also been updated relative to that in the '114 application to reflect issuance of certain patents from cited applications. The cross-reference has also been amended to include an additional priority claim to application 07/805,727, filed December 6, 1991 and predecessor cases. Further, the title of the present application has been amended relative to '114 to conform to the pending claims. No new matter is involved in any amendments. In brief, other than for the noted clerical changes and the update of the cross-reference, the specification of that of the present application is that of the '114 application and prior applications incorporated by reference in the same. Accordingly, it submitted that the present application has the same effective filing date as the '114 application (*i.e.*, December 6, 1990), and that at least some of the present claims have priority to the earlier '462 and/or '901 predecessor applications filed March 7, 1990 and June 7, 1989 respectively.

CONCLUSION

In view of the foregoing, Applicants respectfully request early examination on the merits. If the Examiner believes a telephone conference would expedite prosecution of this application, please telephone the undersigned at 415-576-0200.

Respectfully submitted,



Joseph R. Snyder
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JS
SF 1098245 v1

007.060-84675930

PATENT APPLICATION

**SUPPORT BOUND PROBES AND METHODS OF ANALYSIS USING
THE SAME**

Inventors: Stephen P.A. Fodor
Lubert Stryer
Michael C. Pirrung
J. Leighton Read

Assignee: Affymetrix, Inc.

Entity: Large

5

SEQUENCING BY HYBRIDIZATION OF A TARGET NUCLEIC ACID TO A
MATRIX OF DEFINED OLIGONUCLEOTIDES

10

CROSS-REFERENCE TO RELATED APPLICATION

15

The present application is a continuation of 09/056,927 filed April 8, 1998, which is a continuation of 08/670,118 filed June 25, 1996, now US 5,800,992, which is a divisional of 08/168,904 filed December 15, 1993, which is a continuation of 07/624,114, filed December 6, 1990 (incorporated by reference). This application is a continuation-in-part application of commonly assigned patent applications Pirrung et al., U.S.S.N.

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07/362,901 (VLSIPS parent) filed on June 7, 1989; and Pirrung et al., U.S.S.N. 07/492,462 (VLSIPS CIP), filed on March 7, 1990 (now US 5,143,854), which are hereby incorporated herein by reference. This application is also a continuation-in-part of USSN 08/348,471 filed November 30, 1994, which is a continuation of USSN 07/805,727 filed December 6, 1991 (now US 5,424,186), which is a continuation-in-part of USSN

25

07/492,462, filed March 7, 1990 (now US 5,143,854), which is a continuation-in-part of USSN 07/362,901, filed June 7, 1989. Additional commonly assigned applications Barrett et al., U.S.S.N. 07/435,316 (caged biotin parent) filed November 13, 1989; and Barrett et al., U.S.S.N. 07/612,671 (caged biotin CIP), filed November 13, 1990 are also incorporated herein by reference. Additional applications Pirrung et al., U.S.S.N.

30

07/624,120 (now abandoned) a divisional of which has issued as US 5,744,101 and Dower et al., U.S.S.N. 07/626,730 (now US 5,547,839), which are also commonly assigned and filed on the same day as this application, are also hereby incorporated herein by reference.

5 SEQUENCING BY HYBRIDIZATION OF A TARGET NUCLEIC ACID
TO A MATRIX OF DEFINED OLIGONUCLEOTIDES

BACKGROUND OF THE INVENTION

The present invention relates to the sequencing, fingerprinting, and mapping of polymers, particularly biological polymers. The inventions may be applied, for example, in the sequencing, fingerprinting, or mapping of nucleic acids, polypeptides, oligosaccharides, and synthetic polymers.

15 The relationship between structure and function of macromolecules is of fundamental importance in the understanding of biological systems. These relationships are important to understanding, for example, the functions of enzymes, structural proteins, and signalling proteins, ways in which cells communicate with each other, as well as mechanisms of cellular control and metabolic feedback.

Genetic information is critical in continuation of life processes. Life is substantially informationally based and its genetic content controls the growth and reproduction of the organism and its complements. Polypeptides, which are critical features of all living systems, are encoded by the genetic material of the cell. In particular, the properties of enzymes, functional proteins, and structural proteins are determined by the sequence of amino acids which make them up. As structure and function are integrally related, many biological functions may be explained by elucidating the underlying the structural features which provide those functions. For this reason, it has become very important to determine the genetic sequences of nucleotides which encode the enzymes, structural proteins, and other effectors of biological functions. In addition to segments of nucleotides which encode polypeptides, there are many nucleotide sequences which are involved in control and regulation of gene expression.

The human genome project is directed toward determining the complete sequence the genome of the human

organism. Although such a sequence would not correspond to the sequence of any specific individual, it would provide significant information as to the general organization and specific sequences contained within segments from particular individuals. It would also provide mapping information which is very useful for further detailed studies. However, the need for highly rapid, accurate, and inexpensive sequencing technology is nowhere more apparent than in a demanding sequencing project such as this. To complete the sequencing of a human genome would require the determination of approximately 3×10^9 , or 3 billion base pairs.

The procedures typically used today for sequencing include the Sanger dideoxy method, see, e.g., Sanger et al. (1977) Proc. Natl. Acad. Sci. USA, 74:5463-5467, or the Maxam and Gilbert method, see, e.g., Maxam et al., (1980) Methods in Enzymology, 65:499-559. The Sanger method utilizes enzymatic elongation procedures with chain terminating nucleotides. The Maxam and Gilbert method uses chemical reactions exhibiting specificity of reaction to generate nucleotide specific cleavages. Both methods require a practitioner to perform a large number of complex manual manipulations. These manipulations usually require isolating homogeneous DNA fragments, elaborate and tedious preparing of samples, preparing a separating gel, applying samples to the gel, electrophoresing the samples into this gel, working up the finished gel, and analyzing the results of the procedure.

Thus, a less expensive, highly reliable, and labor efficient means for sequencing biological macromolecules is needed. A substantial reduction in cost and increase in speed of nucleotide sequencing would be very much welcomed. In particular, an automated system would improve the reproducibility and accuracy of procedures. The present invention satisfies these and other needs.

SUMMARY OF THE INVENTION

The present invention provides improved methods useful for de novo sequencing of an unknown polymer sequence, for verification of known sequences, for fingerprinting

polymers, and for mapping homologous segments within a sequence. By reducing the number of manual manipulations required and automating most of the steps, the speed, accuracy, and reliability of these procedures are greatly enhanced.

5 The production of a substrate having a matrix of positionally defined regions with attached reagents exhibiting known recognition specificity can be used for the sequence analysis of a polymer. Although most directly applicable to sequencing, the present invention is also applicable to
10 fingerprinting, mapping, and general screening of specific interactions. The VLSIPS substrates will be applied to evaluating other polymers, e.g., carbohydrates, polypeptides, hydrocarbon synthetic polymers, and the like. For these non-polynucleotides, the sequence specific reagents will usually be
15 antibodies specific for a particular subunit sequence.

 The present invention also provides a means to automate sequencing manipulations. The automation of the substrate production method and of the scan and analysis steps minimizes the need for human intervention. This simplifies the
20 tasks and promotes reproducibility.

 The present invention provides a composition comprising a plurality of positionally distinguishable sequence specific reagents attached to a solid substrate, which reagents are capable of specifically binding to a predetermined subunit
25 sequence of a preselected multi-subunit length having at least three subunits, said reagents representing substantially all possible sequences of said preselected length. In some embodiments, the subunit sequence is a polynucleotide or a polypeptide, in others the preselected multi-subunit length is
30 five subunits and the subunit sequence is a polynucleotide sequence. In other embodiments, the specific reagent is an oligonucleotide of at least about five nucleotides. Alternatively, the specific reagent is a monoclonal antibody. Usually the specific reagents are all attached to a single
35 solid substrate, and the reagents comprise about 3000 different sequences. In other embodiments, the reagents represents at least about 25% of the possible subsequences of said preselected length. Usually, the reagents are localized in

regions of the substrate having a density of at least 25 regions per square centimeter, and often the substrate has a surface area of less than about 4 square centimeters.

The present invention also provides methods for
5 analyzing a sequence of a polynucleotide or a polypeptide, said method comprising the step of:

- a) exposing said polynucleotide or polypeptide to a composition as described.

It also provides useful methods for identifying or
10 comparing a target sequence with a reference, said method comprising the step of:

- a) exposing said target sequence to a composition as described;
- b) determining the pattern of positions of the
15 reagents which specifically interact with the target sequence; and
- c) comparing the pattern with the pattern exhibited by the reference when exposed to the composition.

20 The present invention also provides methods for sequencing a segment of a polynucleotide comprising the steps of:

- a) combining:
 - i) a substrate comprising a plurality of
25 chemically synthesized and positionally distinguishable oligonucleotides capable of recognizing defined oligonucleotide sequences; and
 - ii) a target polynucleotide; thereby forming
30 high fidelity matched duplex structures of complementary subsequences of known sequence; and
- b) determining which of said reagents have specifically interacted with subsequences in
35 said target polynucleotide.

In one embodiment, the segment is substantially the entire length of said polynucleotide.

The invention also provides methods for sequencing a polymer, said method comprising the steps of:

- 5 a) preparing a plurality of reagents which each specifically bind to a subsequence of preselected length;
- b) positionally attaching each of said reagents to one or more solid phase substrates, thereby producing substrates of positionally definable sequence specific probes;
- 10 c) combining said substrates with a target polymer whose sequence is to be determined; and
- d) determining which of said reagents have specifically interacted with subsequences in said target polymer.

15 In one embodiment, the substrates are beads.
Preferably, the plurality of reagents comprise substantially
all possible subsequences of said preselected length found in
said target. In another embodiment, the solid phase substrate
is a single substrate having attached thereto reagents
20 recognizing substantially all possible subsequences of
preselected length found in said target.

In another embodiment, the method further comprises the step of analyzing a plurality of said recognized subsequences to assemble a sequence of said target polymer. In a bead embodiment, at least some of the plurality of substrates have one subsequence specific reagent attached thereto, and the substrates are coded to indicate the sequence specificity of said reagent.

The present invention also embraces a method of using
30 a fluorescent nucleotide to detect interactions with
oligonucleotide probes of known sequence, said method
comprising:

- a) attaching said nucleotide to a target unknown polynucleotide sequence, and
- b) exposing said target polynucleotide sequence to a collection of positionally defined oligonucleotide probes of known sequences to

determine the sequences of said probes which interact with said target.

In a further refinement, an additional step is included of:

- 5 a) collating said known sequences to determine the overlaps of said known sequences to determine the sequence of said target sequence.

10 A method of mapping a plurality of sequences relative to one another is also provided, the method comprising:

- a) preparing a substrate having a plurality of positionally attached sequence specific probes are attached;
- 15 b) exposing each of said sequences to said substrate, thereby determining the patterns of interaction between said sequence specific probes and said sequences; and
- 20 c) determining the relative locations of said sequence specific probe interactions on said sequences to determine the overlaps and order of said sequences.

In one refinement, the sequence specific probes are oligonucleotides, applicable to where the target sequences are nucleic acid sequences.

25 In the nucleic acid sequencing application, the steps of the sequencing process comprise:

- a) producing a matrix substrate having known positionally defined regions of known sequence specific oligonucleotide probes;
- 30 b) hybridizing a target polynucleotide to the positions on the matrix so that each of the positions which contain oligonucleotide probes complementary to a sequence on the target hybridize to the target molecule;
- 35

c) detecting which positions have bound the target, thereby determining sequences which are found on the target; and

5 d) analyzing the known sequences contained in the target to determine sequence overlaps and assembling the sequence of the target therefrom.

The enablement of the sequencing process by hybridization is based in large part upon the ability to
10 synthesize a large number (e.g., to virtually saturate) of the possible overlapping sequence segments and distinguishing those probes which hybridize with fidelity from those which have mismatched bases, and to analyze a highly complex pattern of hybridization results to determine the overlap regions.

15 The detecting of the positions which bind the target sequence would typically be through a fluorescent label on the target. Although a fluorescent label is probably most convenient, other sorts of labels, e.g., radioactive, enzyme linked, optically detectable, or spectroscopic labels may be
20 used. Because the oligonucleotide probes are positionally defined, the location of the hybridized duplex will directly translate to the sequences which hybridize. Thus, upon analysis of the positions provides a collection of subsequences found within the target sequence. These subsequences are
25 matched with respect to their overlaps so as to assemble an intact target sequence.

In one preferred embodiment, linker molecules are provided on a substrate. A terminal end of the linker molecules is provided with a reactive functional group protected with a photoremovable protective group.

5 Using lithographic methods, the photoremovable protective group is exposed to light and removed from the linker molecules in first selected regions. The substrate is then washed or otherwise contacted with a first monomer that reacts with exposed functional groups on the linker
10 molecules. In a preferred embodiment, the monomer is an amino acid containing a photoremovable protective group at its amino or carboxy terminus and the linker molecule terminates in an amino or carboxy acid group bearing a photoremovable protective group.

15 A second set of selected regions is, thereafter, exposed to light and the photoremovable protective group on the linker molecule/protected amino acid is removed at the second set of regions. The substrate is then contacted with a second monomer
20 containing a photoremovable protective group for reaction with exposed functional groups. This process is repeated to selectively apply monomers until polymers of a desired length and desired chemical sequence are obtained.
25 Photolabile groups are then optionally removed and the sequence is, thereafter, optionally capped. Side chain protective groups, if present, are also removed.

An improved method and apparatus for the preparation of polymers is disclosed. The method and
30 apparatus may be applied to synthesize a variety of polymers at known locations on a substrate. The method could be used to synthesize up to about 10^6 or more different sequences per cm^2 at known locations in some
35 embodiments.

000000-000400

The method enables greater ease in peptide synthesis because the physical separation of reagents is not required when growing polymer chains. The chains themselves are separated by different physical locations on the substrate, but the entire substrate is exposed to the various reagents as the synthesis is conducted. Differential reaction is achieved by selectively exposing reactive functional groups to, e.g., light, electric currents, or another spatially localized activator. Remaining areas on the substrate remain unreacted.

By using the lithographic techniques disclosed herein, it is possible to direct light to relatively small and precisely known locations on the substrate. It is, therefore, possible to synthesize polymers of a known chemical sequence at known locations on the substrate.

The resulting substrate will have a variety of uses including, for example, screening large numbers of polymers for biological activity. To screen for biological activity, the substrate is exposed to one or more receptors such as antibody whole cells, receptors on vesicles, lipids, or any one of a variety of other receptors. The receptors are preferably labeled with, for example, a fluorescent marker, radioactive marker, or a labeled antibody reactive with the receptor. The location of the marker on the substrate is detected with, for example, photon detection or autoradiographic techniques. Through knowledge of the sequence of the material at the location where binding is detected, it is possible to quickly determine which sequence binds with the receptor and, therefore, the technique can be used to screen large numbers of peptides. Other possible applications of the inventions herein include diagnostics in which various antibodies for particular receptors would be placed on a substrate and, for example, blood sera would be screened for immune deficiencies. Still further applications include, for example, selective "doping" of organic materials in semiconductor devices, and the like.

In connection with one aspect of the invention an improved reactor system for synthesizing polymers is also disclosed. The reactor system includes a substrate mount which engages a substrate around a periphery thereof. The substrate mount provides for a reactor space between the substrate and the mount through or into which reaction fluids are pumped or flowed. A mask is placed on or focused on the substrate and illuminated so as to deprotect selected regions of the substrate in the reactor space. A monomer is pumped through the reactor space or otherwise contacted with the substrate and reacts with the deprotected regions. By selectively deprotecting regions on the substrate and flowing predetermined monomers through the reactor space, desired polymers at known locations may be synthesized.

Improved detection apparatus and methods are also disclosed. The detection method and apparatus utilize a substrate having a large variety of polymer sequences at known locations on a surface thereof. The substrate is exposed to a fluorescently labeled receptor which binds to one or more of the polymer sequences. The substrate is placed in a microscope detection apparatus for identification of locations where binding takes place. The microscope detection apparatus includes a monochromatic or polychromatic light source for directing light at the substrate, means for detecting fluoresced light from the substrate, and means for determining a location of the fluoresced light. The means for detecting light fluoresced on the substrate may in some embodiments include a photon counter. The means for determining a location of the fluoresced light may include an x/y translation table for the substrate. Translation of the slide and data collection are recorded and managed by an appropriately programmed digital computer.

A further understanding of the nature and advantages of the inventions herein may be realized by reference to the remaining portions of the specification and the attached drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 illustrates a flow chart for sequence, fingerprint, or mapping analysis.

Fig. 2 illustrates the proper function of a VLSIPS peptide synthesis.

Fig. 3 illustrates the proper function of a VLSIPS dipeptide synthesis.

Fig. 4 illustrates the process of a VLSIPS trinucleotide synthesis.

Fig. 5 illustrates masking and irradiation of a substrate at a first location. The substrate is shown in cross-section;

Fig. 6 illustrates the substrate after application of a monomer "A";

Fig. 7 illustrates irradiation of the substrate at a second location;

Fig. 8 illustrates the substrate after application of monomer "B";

Fig. 9 illustrates irradiation of the "A" monomer;

Fig. 10 illustrates the substrate after a second application of "B";

Fig. 11 illustrates a completed substrate;

Figs. 12A and 12B illustrate alternative embodiments of a reactor system for forming a plurality of polymers on a substrate;

Fig. 13 illustrates a detection apparatus for locating fluorescent markers on the substrate;

Figs. 14A-14M illustrate the method as it is applied to the production of the trimers of monomers "A" and "B";

Figs. 15 A and 15 B are fluorescence traces for standard fluorescent beads;

Figs. 16A and 16B are fluorescence curves for NVOC slides not exposed and exposed to light respectively;

Figs. 17 A to 17 D are fluorescence plots of slides exposed through 100 μm , 50 μm , 20 μm , and 10 μm masks;

Fig. 18 illustrates fluorescence of a slide with the peptide YGGFL on selected regions of its surface which has been exposed to labeled Herz antibody specific for this sequence;

Figs. 19A to 19 D illustrate formation of and a fluorescence plot of a slide with a checkerboard pattern of YGGFL and GGFL exposed to labeled Herz antibody.

Fig. 19 C illustrates a 500x500 μm mask which has been focused on the substrate according to Fig. 12 A while Fig. 19 D illustrates a 50x50 μm mask placed in direct contact with the substrate in accord with Fig. 12 B;

Fig. 20 is a fluorescence plot of YGGFL and PGGFL synthesized in a 50 μm checkerboard pattern;

Fig. 21 is a fluorescence plot of YPGGFL and YGGFL synthesized in a 50 μm checkerboard pattern;

Figs. 22 A and 22 B illustrate the mapping of sixteen sequences synthesized on two different glass slides;

Fig. 23 is a fluorescence plot of the slide illustrated in Fig. 22 A ; and

Fig. 24 is a fluorescence plot of the slide illustrated in Fig. 14 B.

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* * *

30 I. OVERALL DESCRIPTION

A. General

The present invention relies in part on the ability to synthesize or attach specific recognition reagents at known locations on a substrate, typically a single substrate. In particular, the present invention provides the ability to prepare a substrate having a very high density matrix pattern of positionally defined specific recognition reagents. The reagents are capable of interacting with their specific targets while attached to the substrate, e.g., solid phase interactions, and by appropriate labeling of these targets, the sites of the interactions between the target and the specific reagents may be derived. Because the reagents are positionally defined, the sites of the interactions will define the specificity of each interaction. As a result, a map of the patterns of interactions with specific reagents on the substrate is convertible into information on the specific interactions taking place, e.g., the recognized features.

Where the specific reagents recognize a large number of possible features, this system allows the determination of the combination of specific interactions which exist on the target molecule. Where the number of features is sufficiently large, the identical same combination, or pattern, of features is sufficiently unlikely that a particular target molecule may often be uniquely defined by its features. In the extreme, the features may actually be the subunit sequence of the target molecule, and a given target sequence may be uniquely defined by its combination of features.

In particular, the methodology is applicable to sequencing polynucleotides. The specific sequence recognition reagents will typically be oligonucleotide probes which hybridize with specificity to subsequences found on the target sequence. A sufficiently large number of those probes allows the fingerprinting of a target polynucleotide or the relative mapping of a collection of target polynucleotides, as described in greater detail below.

In the high resolution fingerprinting provided by a saturating collection of probes which include all possible subsequences of a given size, e.g., 10-mers, collating of all the subsequences and determination of specific overlaps will be derived and the entire sequence can usually be reconstructed.

Although a polynucleotide sequence analysis is a preferred embodiment, for which the specific reagents are most easily accessible, the invention is also applicable to analysis of other polymers, including polypeptides, carbohydrates, and synthetic polymers, including α -, β -, and ω -amino acids, polyurethanes, polyesters, polycarbonates, polyureas, polyamides, polyethyleneimines, polyarylene sulfides, polysiloxanes, polyimides, polyacetates, and mixed polymers. Various optical isomers, e.g., various D- and L- forms of the monomers, may be used.

Sequence analysis will take the form of complete sequence determination, to the level of the sequence of individual subunits along the entire length of the target sequence. Sequence analysis also takes the form of sequence homology, e.g., less than absolute subunit resolution, where

"similarity" in the sequence will be detectable, or the form of selective sequences of homology interspersed at specific or irregular locations.

In either case, the sequence is determinable at selective resolution or at particular locations. Thus, the hybridization method will be useful as a means for identification, e.g., a "fingerprint", much like a Southern hybridization method is used. It is also useful to map particular target sequences.

B. VLSIPS Substrates

The invention is enabled by the development of technology to prepare substrates on which specific reagents may be either positionally attached or synthesized. In particular, the very large scale immobilized polymer synthesis (VLSIPS) technology allows for the very high density production of an enormous diversity of reagents mapped out in a known matrix pattern on a substrate. These reagents specifically recognize subsequences in a target polymer and bind thereto, producing a map of positionally defined regions of interaction. These map positions are convertible into actual features recognized, and thus would be present in the target molecule of interest.

As indicated, the sequence specific recognition reagents will often be oligonucleotides which hybridize with fidelity and discrimination to the target sequence. For use with other polymers, monoclonal or polyclonal antibodies having high sequence specificity will often be used.

In the generic sense, the VLSIPS technology allows the production of a substrate with a high density matrix of positionally mapped regions with specific recognition reagents attached at each distinct region. By use of protective groups which can be positionally removed, or added, the regions can be activated or deactivated for addition of particular reagents or compounds. Details of the protection are described below and in related application U.S.S.N. 07/492,462 (VLSIPS CIP). In a preferred embodiment, photosensitive protecting agents will be used and the regions of activation or deactivation may be controlled by electro-optical and optical methods, similar to

many of the processes used in semiconductor wafer and chip fabrication.

In the nucleic acid nucleotide sequencing application, a VLSIPS substrate is synthesized having positionally defined oligonucleotide probes. See U.S.S.N. 07/492,462 (VLSIPS CIP); and U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS). By use of masking technology and photosensitive synthetic subunits, the VLSIPS apparatus allows for the stepwise synthesis of polymers according to a positionally defined matrix pattern. Each oligonucleotide probe will be synthesized at known and defined positional locations on the substrate. This forms a matrix pattern of known relationship between position and specificity of interaction. The VLSIPS technology allows the production of a very large number of different oligonucleotide probes to be simultaneously and automatically synthesized including numbers in excess of about 10^2 , 10^3 , 10^4 , 10^5 , 10^6 , or even more, and at densities of at least about 10^2 , $10^3/\text{cm}^2$, $10^4/\text{cm}^2$, $10^5/\text{cm}^2$ and up to $10^6/\text{cm}^2$ or more. This application discloses methods for synthesizing polymers on a silicon or other suitably derivatized substrate, methods and chemistry for synthesizing specific types of biological polymers on those substrates, apparatus for scanning and detecting whether interaction has occurred at specific locations on the substrate, and various other technologies related to the use of a high density very large scale immobilized polymer substrate. In particular, sequencing, fingerprinting, and mapping applications are discussed herein in detail, though related technologies are described in simultaneously filed applications U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS) and U.S.S.N. / , , attorney docket number 11509-16 (sequencing by synthesis), each of which is hereby incorporated herein by reference.

In other embodiments, antibody probes will be generated which specifically recognize particular subsequences found on a polymer. Antibodies would be generated which are specific for recognizing a three contiguous amino acid sequence, and monoclonal antibodies may be preferred.

Optimally, these antibodies would not recognize any sequences other than the specific three amino acid stretch desired and the binding affinity should be insensitive to flanking or remote sequences found on a target molecule. Likewise, antibodies specific for particular carbohydrate linkages or sequences will be generated. A similar approach could be used for preparing specific reagents which recognize other polymer subunit sequences. These reagents would typically be site specifically localized to a substrate matrix pattern where the regions are closely packed.

These reagents could be individually attached at specific sites on the substrate in a matrix by an automated procedure where the regions are positionally targeted by some other specific mechanism, e.g., one which would allow the entire collection of reagents to be attached to the substrate in a single reaction. Each reagent could be separately attached to a specific oligonucleotide sequence by an automated procedure. This would produce a collection of reagents where, e.g., each monoclonal antibody would have a unique oligonucleotide sequence attached to it. By virtue of a VLSIPS substrate which has different complementary oligonucleotides synthesized on it, each monoclonal antibody would specifically be bound only at that site on the substrate where the complementary oligonucleotide has been synthesized. A crosslinking step would fix the reagent to the substrate. See, e.g., Dattagupta et al. (1985) U.S. Patent No. 4,542,102 and (1987) U.S. Pat. No. 4,713,326; and Chatterjee, M. et al. (1990) J. Am. Chem. Soc. 112:6997-____, which are hereby incorporated herein by reference. This allows a high density positionally specific collection of specific recognition reagents, e.g., monoclonal antibodies, to be immobilized to a solid substrate using an automated system.

The regions which define particular reagents will usually be generated by selective protecting groups which may be activated or deactivated. Typically the protecting group will be bound to a monomer subunit or spatial region, and can be spatially affected by an activator, such as electromagnetic radiation. Examples of protective groups with utility herein

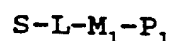
include nitroveratryl oxycarbonyl (NVOC), nitrobenzyl oxycarbonyl (NBOC), dimethyl dimethoxy benzyloxy carbonyl, 5-bromo-7-nitroindolinyl, O-hydroxy- α -methyl cinnamoyl, and 2-oxymethylene anthraquinone. Examples of activators include ion
5 beams, electric fields, magnetic fields, electron beams, x-ray, and other forms of electromagnetic radiation.

The present invention provides methods and apparatus for the preparation and use of a substrate having a plurality of polymer sequences in predefined
10 regions. The invention is described herein primarily with regard to the preparation of molecules containing sequences of amino acids, but could readily be applied in the preparation of other polymers. Such polymers include, for example, both linear and cyclic polymers
15 of nucleic acids, polysaccharides, phospholipids, and peptides having either α -, β -, or ω -amino acids, heteropolymers in which a known drug is covalently bound to any of the above, polyurethanes, polyesters, polycarbonates, polyureas, polyamides, polyethyleneimines, polyarylene
20 sulfides, polysiloxanes, polyimides, polyacetates, or other polymers which will be apparent upon review of this disclosure. In a preferred embodiment, the invention herein is used in the synthesis of peptides.

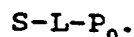
The prepared substrate may, for example, be
25 used in screening a variety of polymers as ligands for binding with a receptor, although it will be apparent that the invention could be used for the synthesis of a receptor for binding with a ligand. The substrate disclosed herein will have a wide variety of other uses.
30 Merely by way of example, the invention herein can be used in determining peptide and nucleic acid sequences which bind to proteins, finding sequence-specific binding drugs, identifying epitopes recognized by antibodies, and evaluation of a variety of drugs for clinical and
35 diagnostic applications, as well as combinations of the above.

Concurrently or after exposure of a known region of the substrate to light, the surface is contacted with a first monomer unit M_1 which reacts with the functional group which has been exposed by the deprotection step. The first monomer includes a protective group P_1 . P_1 may or may not be the same as P_0 .

Accordingly, after a first cycle, known first regions of the surface may comprise the sequence:



while remaining regions of the surface comprise the sequence:

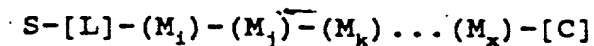


Thereafter, second regions of the surface (which may include the first region) are exposed to light and contacted with a second monomer M_2 (which may or may not be the same as M_1) having a protective group P_2 . P_2 may or may not be the same as P_0 and P_1 . After this second cycle, different regions of the substrate may comprise one or more of the following sequences:



The above process is repeated until the substrate includes desired polymers of desired lengths. By controlling the locations of the substrate exposed to light and the reagents exposed to the substrate following exposure, the location of each sequence will be known.

Thereafter, the protective groups are removed from some or all of the substrate and the sequences are, optionally, capped with a capping unit C. The process results in a substrate having a surface with a plurality of polymers of the following general formula:



where square brackets indicate optional groups, and $M_1\cdots M_x$ indicates any sequence of monomers. The number of monomers could cover a wide variety of values, but in a preferred embodiment they will range from 2 to 100.

In some embodiments a plurality of locations on the substrate polymers are to contain a common monomer subsequence. For example, it may be desired to synthesize a sequence $S-M_1-M_2-M_3$ at first locations and a sequence $S-M_4-M_2-M_3$ at second locations. The process would commence with irradiation of the first locations followed by contacting with M_1-P , resulting in the sequence $S-M_1-P$ at the first location. The second locations would then be irradiated and contacted with M_4-P , resulting in the sequence $S-M_4-P$ at the second locations. Thereafter both the first and second locations would be irradiated and contacted with the dimer M_2-M_3 , resulting in the sequence $S-M_1-M_2-M_3$ at the first locations and $S-M_4-M_2-M_3$ at the second locations. Of course, common subsequences of any length could be utilized including those in a range of 2 or more monomers, 2 to 100 monomers, 2 to 20 monomers, and a most preferred range of 2 to 3 monomers.

According to other embodiments, a set of masks is used for the first monomer layer and, thereafter, varied light wavelengths are used for selective deprotection. For example, in the process discussed

above, first regions are first exposed through a mask and reacted with a first monomer having a first protective group P_1 , which is removable upon exposure to a first wavelength of light (e.g., IR). Second regions are masked and reacted with a second monomer having a second protective group P_2 , which is removable upon exposure to a second wavelength of light (e.g., UV). Thereafter, masks become unnecessary in the synthesis because the entire substrate may be exposed alternatively to the first and second wavelengths of light in the deprotection cycle.

The polymers prepared on a substrate according to the above methods will have a variety of uses including, for example, screening for biological activity. In such screening activities, the substrate containing the sequences is exposed to an unlabeled or labeled receptor such as an antibody, receptor on a cell, phospholipid vesicle, or any one of a variety of other receptors. In one preferred embodiment the polymers are exposed to a first, unlabeled receptor of interest and, thereafter, exposed to a labeled receptor-specific recognition element, which is, for example, an antibody. This process will provide signal amplification in the detection stage.

The receptor molecules may bind with one or more polymers on the substrate. The presence of the labeled receptor and, therefore, the presence of a sequence which binds with the receptor is detected in a preferred embodiment through the use of autoradiography, detection of fluorescence with a charge-coupled device, fluorescence microscopy, or the like. The sequence of the polymer at the locations where the receptor binding is detected may be used to determine all or part of a sequence which is complementary to the receptor.

Use of the invention herein is illustrated primarily with reference to screening for biological activity. The invention will, however, find many other uses. For example, the invention may be used in

information storage (e.g., on optical disks), production of molecular electronic devices, production of stationary phases in separation sciences, production of dyes and brightening agents, photography, and in immobilization of cells, proteins, lectins, nucleic acids, polysaccharides and the like in patterns on a surface via molecular recognition of specific polymer sequences. By synthesizing the same compound in adjacent, progressively differing concentrations, a gradient will be established to control chemotaxis or to develop diagnostic dipsticks which, for example, titrate an antibody against an increasing amount of antigen. By synthesizing several catalyst molecules in close proximity, more efficient multistep conversions may be achieved by "coordinate immobilization." Coordinate immobilization also may be used for electron transfer systems, as well as to provide both structural integrity and other desirable properties to materials such as lubrication, wetting, etc.

According to alternative embodiments, molecular biodistribution or pharmacokinetic properties may be examined. For example, to assess resistance to intestinal or serum proteases, polymers may be capped with a fluorescent tag and exposed to biological fluids of interest.

III. Polymer Synthesis

Fig. 1 illustrates one embodiment of the invention disclosed herein in which a substrate 2 is shown in cross-section. Essentially, any conceivable substrate may be employed in the invention. The substrate may be biological, nonbiological, organic, inorganic, or a combination of any of these, existing as particles, strands, precipitates, gels, sheets, tubing, spheres, containers, capillaries, pads, slices, films, plates, slides, etc. The substrate may have any convenient shape, such as a disc, square, sphere, circle, etc. The substrate is preferably flat but may take on a

variety of alternative surface configurations. For example, the substrate may contain raised or depressed regions on which the synthesis takes place. The substrate and its surface preferably form a rigid support on which to carry out the reactions described herein.

The substrate and its surface is also chosen to provide appropriate light-absorbing characteristics. For instance, the substrate may be a polymerized Langmuir Blodgett film, functionalized glass, Si, Ge, GaAs, GaP, SiO₂, SiN₄, modified silicon, or any one of a wide variety of gels or polymers such as (poly)tetrafluoroethylene, (poly)vinylidenedifluoride, polystyrene, polycarbonate, or combinations thereof. Other substrate materials will be readily apparent to those of skill in the art upon review of this disclosure. In a preferred embodiment the substrate is flat glass or single-crystal silicon with surface relief features of less than 10 Å.

According to some embodiments, the surface of the substrate is etched using well known techniques to provide for desired surface features. For example, by way of the formation of trenches, v-grooves, mesa structures, or the like, the synthesis regions may be more closely placed within the focus point of impinging light, be provided with reflective "mirror" structures for maximization of light collection from fluorescent sources, or the like.

Surfaces on the solid substrate will usually, though not always, be composed of the same material as the substrate. Thus, the surface may be composed of any of a wide variety of materials, for example, polymers, plastics, resins, polysaccharides, silica or silica-based materials, carbon, metals, inorganic glasses, membranes, or any of the above-listed substrate materials. In some embodiments the surface may provide for the use of caged binding members which are attached firmly to the surface of the substrate in accord with the teaching of copending application Serial No. 404,920, previously incorporated

herein by reference. Preferably, the surface will contain reactive groups, which could be carboxyl, amino, hydroxyl, or the like. Most preferably, the surface will be optically transparent and will have surface Si-OH functionalities, such as are found on silica surfaces.

The surface 4 of the substrate is preferably provided with a layer of linker molecules 6, although it will be understood that the linker molecules are not required elements of the invention. The linker molecules are preferably of sufficient length to permit polymers in a completed substrate to interact freely with molecules exposed to the substrate. The linker molecules should be 6-50 atoms long to provide sufficient exposure. The linker molecules may be, for example, aryl acetylene, ethylene glycol oligomers containing 2-10 monomer units, diamines, diacids, amino acids, or combinations thereof. Other linker molecules may be used in light of this disclosure.

According to alternative embodiments, the linker molecules are selected based upon their hydrophilic/hydrophobic properties to improve presentation of synthesized polymers to certain receptors. For example, in the case of a hydrophilic receptor, hydrophilic linker molecules will be preferred so as to permit the receptor to more closely approach the synthesized polymer.

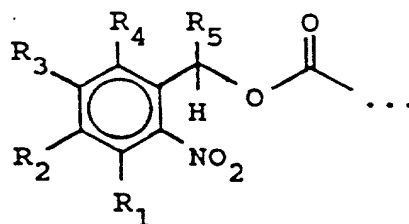
According to another alternative embodiment, linker molecules are also provided with a photocleavable group at an intermediate position. The photocleavable group is preferably cleavable at a wavelength different from the protective group. This enables removal of the various polymers following completion of the synthesis by way of exposure to the different wavelengths of light.

The linker molecules can be attached to the substrate via carbon-carbon bonds using, for example, (poly)trifluorochloroethylene surfaces, or preferably, by siloxane bonds (using, for example, glass or silicon

oxide surfaces). Siloxane bonds with the surface of the substrate may be formed in one embodiment via reactions of linker molecules bearing trichlorosilyl groups. The linker molecules may optionally be attached in an ordered array, i.e., as parts of the head groups in a polymerized Langmuir Blodgett film. In alternative embodiments, the linker molecules are adsorbed to the surface of the substrate. ,

The linker molecules and monomers used herein are provided with a functional group to which is bound a protective group. Preferably, the protective group is on the distal or terminal end of the linker molecule opposite the substrate. The protective group may be either a negative protective group (i.e., the protective group renders the linker molecules less reactive with a monomer upon exposure) or a positive protective group (i.e., the protective group renders the linker molecules more reactive with a monomer upon exposure). In the case of negative protective groups an additional step of reactivation will be required. In some embodiments, this will be done by heating.

The protective group on the linker molecules may be selected from a wide variety of positive light-reactive groups preferably including nitro aromatic compounds such as o-nitrobenzyl derivatives or benzyloxycarbonyl. In a preferred embodiment, 6-nitroveratryloxycarbonyl (NVOC), 2-nitrobenzyloxycarbonyl (NBOC) or α,α -dimethyl-dimethoxybenzyloxycarbonyl (DDZ) is used. In one embodiment, a nitro aromatic compound containing a benzylic hydrogen ortho to the nitro group is used, i.e., a chemical of the form:



where R₁ is alkoxy, alkyl, halo, aryl, alkenyl, or hydrogen; R₂ is alkoxy, alkyl, halo, aryl, nitro, or hydrogen; R₃ is alkoxy, alkyl, halo, nitro, aryl, or hydrogen; R₄ is alkoxy, alkyl, hydrogen, aryl, halo, or nitro; and R₅ is alkyl, alkynyl, cyano, alkoxy, hydrogen, halo, aryl, or alkenyl. Other materials which may be used include o-hydroxy- α -methyl cinnamoyl derivatives. Photoremovable protective groups are described in, for example, Patchornik, *J. Am. Chem. Soc.* (1970) 92:6333 and Amit et al., *J. Org. Chem.* (1974) 39:192, both of which are incorporated herein by reference.

In an alternative embodiment the positive reactive group is activated for reaction with reagents in solution. For example, a 5-bromo-7-nitro indoline group, when bound to a carbonyl, undergoes reaction upon exposure to light at 420 nm.

In a second alternative embodiment, the reactive group on the linker molecule is selected from a wide variety of negative light-reactive groups including a cinnamate group.

Alternatively, the reactive group is activated or deactivated by electron beam lithography, x-ray lithography, or any other radiation. Suitable reactive groups for electron beam lithography include sulfonyl. Other methods may be used including, for example, exposure to a current source. Other reactive groups and methods of activation may be used in light of this disclosure.

As shown in Fig. 5, the linking molecules are preferably exposed to, for example, light through a suitable mask 8 using photolithographic techniques of the type known in the semiconductor industry and described in, for example, Sze, VLSI Technology, McGraw-Hill (1983), and Mead et al., Introduction to VLSI Systems, Addison-Wesley (1980), which are incorporated herein by reference for all purposes. The light may be directed at either the surface containing the protective groups or at the back of the substrate, so long as the substrate is transparent to the wavelength of light needed for removal of the protective groups. In the embodiment shown in Fig. 5, light is directed at the surface of the substrate containing the protective groups. Fig. 5 illustrates the use of such masking techniques as they are applied to a positive reactive group so as to activate linking molecules and expose functional groups in areas 10a and 10b.

The mask 8 is in one embodiment a transparent support material selectively coated with a layer of opaque material. Portions of the opaque material are removed, leaving opaque material in the precise pattern desired on the substrate surface. The mask is brought into close proximity with, imaged on, or brought directly into contact with the substrate surface as shown in Fig. 5. "Openings" in the mask correspond to locations on the substrate where it is desired to remove photoremovable protective groups from the substrate. Alignment may be performed using conventional alignment techniques in which alignment marks (not shown) are used to accurately overlay successive masks with previous patterning steps, or more sophisticated techniques may be used. For example, interferometric techniques such as the one described in Flanders et al., "A New Interferometric Alignment Technique," App. Phys. Lett. (1977) 31:426-428, which is incorporated herein by reference, may be used.

To enhance contrast of light applied to the substrate, it is desirable to provide contrast enhancement materials between the mask and the substrate according to some embodiments. This contrast enhancement layer may comprise a molecule which is decomposed by light such as quinone diazid or a material which is transiently bleached at the wavelength of interest. Transient bleaching of materials will allow greater penetration where light is applied, thereby enhancing contrast. Alternatively, contrast enhancement may be provided by way of a cladded fiber optic bundle.

The light may be from a conventional incandescent source, a laser, a laser diode, or the like. If non-collimated sources of light are used it may be desirable to provide a thick- or multi-layered mask to prevent spreading of the light onto the substrate. It may, further, be desirable in some embodiments to utilize groups which are sensitive to different wavelengths to control synthesis. For example, by using groups which are sensitive to different wavelengths, it is possible to select branch positions in the synthesis of a polymer or eliminate certain masking steps. Several reactive groups along with their corresponding wavelengths for deprotection are provided in Table 1.

Table 1

Group	Approximate Deprotection Wavelength
Nitroveratryloxy carbonyl (NVOC)	UV (300-400 nm)
Nitrobenzyloxy carbonyl (NBOC)	UV (300-350 nm)
Dimethyl dimethoxybenzyloxy carbonyl	UV (280-300 nm)
5-Bromo-7-nitroindolinyl	UV (420 nm)
o-Hydroxy- α -methyl cinnamoyl	UV (300-350 nm)
2-Oxymethylene anthraquinone	UV (350 nm)

While the invention is illustrated primarily herein by way of the use of a mask to illuminate selected regions the substrate, other techniques may also be used. For example, the substrate may be translated under a modulated laser or diode light source. Such techniques are discussed in, for example, U.S. Patent No. 4,719,615 (Feyrer et al.), which is incorporated herein by reference. In alternative embodiments a laser galvanometric scanner is utilized. In other embodiments, the synthesis may take place on or in contact with a conventional liquid crystal (referred to herein as a "light valve") or fiber optic light sources. By appropriately modulating liquid crystals, light may be selectively controlled so as to permit light to contact selected regions of the substrate. Alternatively, synthesis may take place on the end of a series of optical fibers to which light is selectively applied. Other means of controlling the location of light exposure will be apparent to those of skill in the art.

The substrate may be irradiated either in contact or not in contact with a solution (not shown) and is, preferably, irradiated in contact with a solution. The solution contains reagents to prevent the by-products formed by irradiation from interfering with synthesis of the polymer according to some embodiments. Such by-products might include, for example, carbon dioxide, nitrosocarbonyl compounds, styrene derivatives, indole derivatives, and products of their photochemical reactions. Alternatively, the solution may contain reagents used to match the index of refraction of the substrate. Reagents added to the solution may further include, for example, acidic or basic buffers, thiols, substituted hydrazines and hydroxylamines, reducing agents (e.g., NADH) or reagents known to react with a given functional group (e.g., aryl nitroso + glyoxylic acid \rightarrow aryl formhydroxamate + CO₂).

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Either concurrently with or after the irradiation step, the linker molecules are washed or otherwise contacted with a first monomer, illustrated by "A" in regions 12a and 12b in Fig. 6. The first monomer reacts with the activated functional groups of the linkage molecules which have been exposed to light. The first monomer, which is preferably an amino acid, is also provided with a photoprotective group. The photoprotective group on the monomer may be the same as or different than the protective group used in the linkage molecules, and may be selected from any of the above-described protective groups. In one embodiment, the protective groups for the A monomer is selected from the group NBOC and NVOC.

As shown in Fig. 7, the process of irradiating is thereafter repeated, with a mask repositioned so as to remove linkage protective groups and expose functional groups in regions 14a and 14b which are illustrated as being regions which were protected in the previous masking step. As an alternative to repositioning of the first mask, in many embodiments a second mask will be utilized. In other alternative embodiments, some steps may provide for illuminating a common region in successive steps. As shown in Fig. 7, it may be desirable to provide separation between irradiated regions. For example, separation of about 1-5 μm may be appropriate to account for alignment tolerances.

As shown in Fig. 8, the substrate is then exposed to a second protected monomer "B," producing B regions 16a and 16b. Thereafter, the substrate is again masked so as to remove the protective groups and expose reactive groups on A region 12a and B region 16b. The substrate is again exposed to monomer B, resulting in the formation of the structure shown in Fig. 10. The dimers B-A and B-B have been produced on the substrate.

A subsequent series of masking and contacting steps similar to those described above with A (not shown)

provides the structure shown in Fig. 11. The process provides all possible dimers of B and A, i.e., B-A, A-B, A-A, and B-B.

The substrate, the area of synthesis, and the area for synthesis of each individual polymer could be of any size or shape. For example, squares, ellipsoids, rectangles, triangles, circles, or portions thereof, along with irregular geometric shapes, may be utilized. Duplicate synthesis areas may also be applied to a single substrate for purposes of redundancy.

In one embodiment the regions 12 and 16 on the substrate will have a surface area of between about 1 cm^2 and 10^{-10} cm^2 . In some embodiments the regions 12 and 16 have areas of less than about 10^{-1} cm^2 , 10^{-2} cm^2 , 10^{-3} cm^2 , 10^{-4} cm^2 , 10^{-5} cm^2 , 10^{-6} cm^2 , 10^{-7} cm^2 , 10^{-8} cm^2 , or 10^{-10} cm^2 . In a preferred embodiment, the regions 12 and 16 are between about $10 \times 10 \text{ }\mu\text{m}$ and $500 \times 500 \text{ }\mu\text{m}$.

In some embodiments a single substrate supports more than about 10 different monomer sequences and preferably more than about 100 different monomer sequences, although in some embodiments more than about 10^3 , 10^4 , 10^5 , 10^6 , 10^7 , or 10^8 different sequences are provided on a substrate. Of course, within a region of the substrate in which a monomer sequence is synthesized, it is preferred that the monomer sequence be substantially pure. In some embodiments, regions of the substrate contain polymer sequences which are at least about 1%, 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 60%, 70%, 80%, 90%, 95%, 96%, 97%, 98%, or 99% pure.

According to some embodiments, several sequences are intentionally provided within a single region so as to provide an initial screening for biological activity, after which materials within regions exhibiting significant binding are further evaluated.

IV. Details of One Embodiment of a Reactor System

Fig. 12 A schematically illustrates a preferred embodiment of a reactor system 100 for synthesizing polymers on the prepared substrate in accordance with one aspect of the invention. The reactor system includes a body 102 with a cavity 104 on a surface thereof. In preferred embodiments the cavity 104 is between about 50 and 1000 μm deep with a depth of about 500 μm preferred.

The bottom of the cavity is preferably provided with an array of ridges 106 which extend both into the plane of the Figure and parallel to the plane of the Figure. The ridges are preferably about 50 to 200 μm deep and spaced at about 2 to 3mm. The purpose of the ridges is to generate turbulent flow for better mixing. The bottom surface of the cavity is preferably light absorbing so as to prevent reflection of impinging light.

A substrate 112 is mounted above the cavity 104. The substrate is provided along its bottom surface 114 with a photoremovable protective group such as NVOC with or without an intervening linker molecule. The substrate is preferably transparent to a wide spectrum of light, but in some embodiments is transparent only at a wavelength at which the protective group may be removed (such as UV in the case of NVOC). The substrate in some embodiments is a conventional microscope glass slide or cover slip. The substrate is preferably as thin as possible, while still providing adequate physical support. Preferably, the substrate is less than about 1 mm thick, more preferably less than 0.5 mm thick, more preferably less than 0.1 mm thick, and most preferably less than 0.05 mm thick. In alternative preferred embodiments, the substrate is quartz or silicon.

The substrate and the body serve to seal the cavity except for an inlet port 108 and an outlet port 110. The body and the substrate may be mated for sealing in some embodiments with one or more gaskets. According to a preferred embodiment, the body is provided with two

concentric gaskets and the intervening space is held at vacuum to ensure mating of the substrate to the gaskets.

Fluid is pumped through the inlet port into the cavity by way of a pump 116 which may be, for example, a model no. B-120-S made by Eldex Laboratories. Selected fluids are circulated into the cavity by the pump, through the cavity, and out the outlet for recirculation or disposal. The reactor may be subjected to ultrasonic radiation and/or heated to aid in agitation in some embodiments.

Above the substrate 112, a lens 120 is provided which may be, for example, a 2" 100mm focal length fused silica lens. For the sake of a compact system, a reflective mirror 122 may be provided for directing light from a light source 124 onto the substrate. Light source 124 may be, for example, a Xe(Hg) light source manufactured by Oriel and having model no. 66024. A second lens 126 may be provided for the purpose of projecting a mask image onto the substrate in combination with lens 112. This form of lithography is referred to herein as projection printing. As will be apparent from this disclosure, proximity printing and the like may also be used according to some embodiments.

Light from the light source is permitted to reach only selected locations on the substrate as a result of mask 128. Mask 128 may be, for example, a glass slide having etched chrome thereon. The mask 128 in one embodiment is provided with a grid of transparent locations and opaque locations. Such masks may be manufactured by, for example, Photo Sciences, Inc. Light passes freely through the transparent regions of the mask, but is reflected from or absorbed by other regions. Therefore, only selected regions of the substrate are exposed to light.

As discussed above, light valves (LCD's) may be used as an alternative to conventional masks to selectively expose regions of the substrate.

Alternatively, fiber optic faceplates such as those available from Schott Glass, Inc, may be used for the purpose of contrast enhancement of the mask or as the sole means of restricting the region to which light is applied. Such faceplates would be placed directly above or on the substrate in the reactor shown in Fig. 8A. In still further embodiments, flys-eye lenses, tapered fiber optic faceplates, or the like, may be used for contrast enhancement.

In order to provide for illumination of regions smaller than a wavelength of light, more elaborate techniques may be utilized. For example, according to one preferred embodiment, light is directed at the substrate by way of molecular microcrystals on the tip of, for example, micropipettes. Such devices are disclosed in Lieberman *et al.*, "A Light Source Smaller Than the Optical Wavelength," *Science* (1990) 247:59-61, which is incorporated herein by reference for all purposes.

In operation, the substrate is placed on the cavity and sealed thereto. All operations in the process of preparing the substrate are carried out in a room lit primarily or entirely by light of a wavelength outside of the light range at which the protective group is removed. For example, in the case of NVOC, the room should be lit with a conventional dark room light which provides little or no UV light. All operations are preferably conducted at about room temperature.

A first, deprotection fluid (without a monomer) is circulated through the cavity. The solution preferably is of 5 mM sulfuric acid in dioxane solution which serves to keep exposed amino groups protonated and decreases their reactivity with photolysis by-products. Absorptive materials such as N,N-diethylamino 2,4-dinitrobenzene, for example, may be included in the deprotection fluid which serves to absorb light and prevent reflection and unwanted photolysis.

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The slide is, thereafter, positioned in a light raypath from the mask such that first locations on the substrate are illuminated and, therefore, deprotected. In preferred embodiments the substrate is illuminated for between about 1 and 15 minutes with a preferred illumination time of about 10 minutes at 10-20 mW/cm² with 365 nm light. The slides are neutralized (i.e., brought to a pH of about 7) after photolysis with, for example, a solution of di-isopropylethylamine (DIEA) in methylene chloride for about 5 minutes.

The first monomer is then placed at the first locations on the substrate. After irradiation, the slide is removed, treated in bulk, and then reinstalled in the flow cell. Alternatively, a fluid containing the first monomer, preferably also protected by a protective group, is circulated through the cavity by way of pump 116. If, for example, it is desired to attach the amino acid Y to the substrate at the first locations, the amino acid Y (bearing a protective group on its α -nitrogen), along with reagents used to render the monomer reactive, and/or a carrier, is circulated from a storage container 118, through the pump, through the cavity, and back to the inlet of the pump.

The monomer carrier solution is, in a preferred embodiment, formed by mixing of a first solution (referred to herein as solution "A") and a second solution (referred to herein as solution "B"). Table 2 provides an illustration of a mixture which may be used for solution A.

Table 2
Representative Monomer Carrier Solution "A"

5	100 mg NVOC amino protected amino acid
	37 mg HOBt (1-Hydroxybenzotriazole)
	250 μ l DMF (Dimethylformamide)
	86 μ l DIEA (Diisopropylethylamine)

10	The composition of solution B is illustrated in Table 3. Solutions A and B are mixed and allowed to react at room temperature for about 8 minutes, then diluted with 2 ml of DMF, and 500 μ l are applied to the surface of the slide or the solution is circulated
15	
	through the reactor system and allowed to react for about 2 hours at room temperature. The slide is then washed with DMF, methylene chloride and ethanol.

Table 3
Representative Monomer Carrier Solution "B"

	250 μ l DMF
25	111 mg BOP (Benzotriazolyl-n-oxy-tris(dimethylamino) phosphoniumhexafluorophosphate)

30	As the solution containing the monomer to be attached is circulated through the cavity, the amino acid or other monomer will react at its carboxy terminus with amino groups on the regions of the substrate which have been deprotected. Of course, while the invention is illustrated by way of circulation of the monomer through the cavity, the invention could be practiced by way of removing the slide from the reactor and submersing it in an appropriate monomer solution.
35	

After addition of the first monomer, the solution containing the first amino acid is then purged from the system. After circulation of a sufficient amount of the DMF/methylene chloride such that removal of the amino acid can be assured (e.g., about 50x times the volume of the cavity and carrier lines), the mask or substrate is repositioned, or a new mask is utilized such that second regions on the substrate will be exposed to light and the light 124 is engaged for a second exposure. This will deprotect second regions on the substrate and the process is repeated until the desired polymer sequences have been synthesized.

The entire derivatized substrate is then exposed to a receptor of interest, preferably labeled with, for example, a fluorescent marker, by circulation of a solution or suspension of the receptor through the cavity or by contacting the surface of the slide in bulk. The receptor will preferentially bind to certain regions of the substrate which contain complementary sequences.

Antibodies are typically suspended in what is commonly referred to as "supercocktail," which may be, for example, a solution of about 1% BSA (bovine serum albumin), 0.5% Tween in PBS (phosphate buffered saline) buffer. The antibodies are diluted into the supercocktail buffer to a final concentration of, for example, about 0.1 to 4 $\mu\text{g/ml}$.

Fig. 12 B illustrates an alternative preferred embodiment of the reactor shown in Fig. 8A. According to this embodiment, the mask 128 is placed directly in contact with the substrate. Preferably, the etched portion of the mask is placed face down so as to reduce the effects of light dispersion. According to this embodiment, the imaging lenses 120 and 126 are not necessary because the mask is brought into close proximity with the substrate.

For purposes of increasing the signal-to-noise ratio of the technique, some embodiments of the invention provide for exposure of the substrate to a first labeled or unlabeled receptor followed by exposure of a labeled,
5 second receptor (e.g., an antibody) which binds at multiple sites on the first receptor. If, for example, the first receptor is an antibody derived from a first species of an animal, the second receptor is an antibody derived from a second species directed to epitopes
10 associated with the first species. In the case of a mouse antibody, for example, fluorescently labeled goat antibody or antiserum which is antimouse may be used to bind at multiple sites on the mouse antibody, providing several times the fluorescence compared to the attachment
15 of a single mouse antibody at each binding site. This process may be repeated again with additional antibodies (e.g., goat-mouse-goat, etc.) for further signal amplification.

In preferred embodiments an ordered sequence of masks is utilized. In some embodiments it is possible to use as few as a single mask to synthesize all of the possible polymers of a given monomer set.

If, for example, it is desired to synthesize all 16 dinucleotides from four bases, a 1 cm square
25 synthesis region is divided conceptually into 16 boxes, each 0.25 cm wide. Denote the four monomer units by A, B, C, and D. The first reactions are carried out in four vertical columns, each 0.25 cm wide. The first mask exposes the left-most column of boxes, where A is
30 coupled. The second mask exposes the next column, where B is coupled; followed by a third mask, for the C column; and a final mask that exposes the right-most column, for D. The first, second, third, and fourth masks may be a single mask translated to different
35 locations.

5 The process is repeated in the horizontal direction for the second unit of the dimer. This time, the masks allow exposure of horizontal rows, again 0.25 cm wide. A, B, C, and D are sequentially coupled using masks that expose horizontal fourths of the reaction area. The resulting substrate contains all 16 dinucleotides of four bases.

10 The eight masks used to synthesize the dinucleotide are related to one another by translation or rotation. In fact, one mask can be used in all eight steps if it is suitably rotated and translated. For example, in the example above, a mask with a single transparent region could be sequentially used to expose each of the vertical columns, translated 90°, and then sequentially used to allow exposure of the horizontal rows.

15 Tables 4 and 5 provide a simple computer program in Quick Basic for planning a masking program and a sample output, respectively, for the synthesis of a polymer chain of three monomers ("residues") having three different monomers in the first level, four different monomers in the second level, and five different monomers in the third level in a striped pattern. The output of the program is the number of cells, the number of "stripes" (light regions) on each mask, and the amount of translation required for each exposure of the mask.

Table 4
Mask Strategy Program

```

DEFINT A-Z
DIM b(20), w(20), l(500)
F$ = "LPT1:"
OPEN f$ FOR OUTPUT AS #1

jmax = 3          'Number of residues
b(1) = 3: b(2) = 4: b(3) = 5      'Number of building blocks for res 1,2,3
g = 1: lmax(1) = 1

FOR j = 1 TO jmax: g = g * b(j): NEXT j

w(0) = 0: w(1) = g / b(1)

PRINT #1, "MASK2.BAS ", DATE$, TIME$: PRINT #1,
PRINT #1, USING "Number of residues-##"; jmax
FOR j = 1 TO jmax
PRINT #1, USING "      Residue ##      ## building blocks"; j; b(j)
NEXT j
PRINT #1, "
PRINT #1, USING "Number of cells-####"; g: PRINT #1,

FOR j = 2 TO jmax
lmax(j) = lmax(j - 1) * b(j - 1)
w(j) = w(j - 1) / b(j)
NEXT j

FOR j = 1 TO jmax
PRINT #1, USING "Mask for residue ##"; j: PRINT #1,
PRINT #1, USING "      Number of stripes-####"; lmax(j)
PRINT #1, USING "      Width of each stripe-####"; w(j)
FOR l = 1 TO lmax(j)
a = 1 + (l - 1) * w(j - 1)
ae = a + w(j) - 1
PRINT #1, USING "      Stripe ## begins at location ### and ends at ###"; l; a; ae
NEXT l
PRINT #1,
PRINT #1, USING "      For each of ## building blocks, translate mask by ##
cell(s)"; b(j); w(j),
PRINT #1, : PRINT #1, : PRINT #1,
NEXT j

```

Table 5
Masking Strategy Output

Number of residues= 3

Residue 1	3 building blocks
Residue 2	4 building blocks
Residue 3	5 building blocks

Number of cells= 60

Mask for residue 1

Number of stripes= 1

Width of each stripe= 20

Stripe 1 begins at location 1 and ends at 20

For each of 3 building blocks, translate mask by 20 cell(s)

Mask for residue 2

Number of stripes= 3

Width of each stripe= 5

Stripe 1 begins at location 1 and ends at 5

Stripe 2 begins at location 21 and ends at 25

Stripe 3 begins at location 41 and ends at 45

For each of 4 building blocks, translate mask by 5 cell(s)

Mask for residue 3

Number of stripes= 12

Width of each stripe= 1

Stripe 1 begins at location 1 and ends at 1

Stripe 2 begins at location 6 and ends at 6

Stripe 3 begins at location 11 and ends at 11

Stripe 4 begins at location 16 and ends at 16

Stripe 5 begins at location 21 and ends at 21

Stripe 6 begins at location 26 and ends at 26

Stripe 7 begins at location 31 and ends at 31

Stripe 8 begins at location 36 and ends at 36

Stripe 9 begins at location 41 and ends at 41

Stripe 10 begins at location 46 and ends at 46

Stripe 11 begins at location 51 and ends at 51

Stripe 12 begins at location 56 and ends at 56

For each of 5 building blocks, translate mask by 1 cell(s)

V. Details of One Embodiment of
A Fluorescent Detection Device

Fig. 13 illustrates a fluorescent detection device for detecting fluorescently labeled receptors on a substrate. A substrate 112 is placed on an x/y translation table 202. In a preferred embodiment the x/y translation table is a model no. PM500-A1 manufactured by Newport Corporation. The x/y translation table is connected to and controlled by an appropriately programmed digital computer 204 which may be, for example, an appropriately programmed IBM PC/AT or AT compatible computer. Of course, other computer systems, special purpose hardware, or the like could readily be substituted for the AT computer used herein for illustration. Computer software for the translation and data collection functions described herein can be provided based on commercially available software including, for example, "Lab Windows" licensed by National Instruments, which is incorporated herein by reference for all purposes.

The substrate and x/y translation table are placed under a microscope 206 which includes one or more objectives 208. Light (about 488 nm) from a laser 210, which in some embodiments is a model no. 2020-05 argon ion laser manufactured by Spectraphysics, is directed at the substrate by a dichroic mirror 207 which passes greater than about 520 nm light but reflects 488 nm light. Dichroic mirror 207 may be, for example, a model no. FT510 manufactured by Carl Zeiss. Light reflected from the mirror then enters the microscope 206 which may be, for example, a model no. Axioscop 20 manufactured by Carl Zeiss. Fluorescein-marked materials on the substrate will fluoresce >488 nm light, and the fluoresced light will be collected by the microscope and passed through the mirror. The fluorescent light from the substrate is then directed through a wavelength filter 209 and, thereafter through an aperture plate 211.

Wavelength filter 209 may be, for example, a model no. OG530 manufactured by Melles Griot and aperture plate 211 may be, for example, a model no. 477352/477380 manufactured by Carl Zeiss.

5 The fluoresced light then enters a
photomultiplier tube 212 which in some embodiments is a
model no. R943-02 manufactured by Hamamatsu, the signal
is amplified in preamplifier 214 and photons are counted
by photon counter 216. The number of photons is recorded
10 as a function of the location in the computer 204.
Pre-Amp 214 may be, for example, a model no. SR440
manufactured by Stanford Research Systems and photon
counter 216 may be a model no. SR400 manufactured by
Stanford Research Systems. The substrate is then moved
15 to a subsequent location and the process is repeated.
In preferred embodiments the data are acquired every 1 to
100 μm with a data collection diameter of about 0.8 to
10 μm preferred. In embodiments with sufficiently high
fluorescence, a CCD detector with broadfield illumination
20 is utilized.

By counting the number of photons generated in
a given area in response to the laser, it is possible to
determine where fluorescent marked molecules are located
on the substrate. Consequently, for a slide which has a
25 matrix of polypeptides, for example, synthesized on the
surface thereof, it is possible to determine which of the
polypeptides is complementary to a fluorescently marked
receptor.

According to preferred embodiments, the
30 intensity and duration of the light applied to the
substrate is controlled by varying the laser power and
scan stage rate for improved signal-to-noise ratio by
maximizing fluorescence emission and minimizing
background noise.

35 While the detection apparatus has been
illustrated primarily herein with regard to the detection
of marked receptors, the invention will find application

in other areas. For example, the detection apparatus disclosed herein could be used in the fields of catalysis, DNA or protein gel scanning, and the like.

5 VI. Determination of Relative
Binding Strength of Receptors

The signal-to-noise ratio of the present invention is sufficiently high that not only can the presence or absence of a receptor on a ligand be
10 detected, but also the relative binding affinity of receptors to a variety of sequences can be determined.

In practice it is found that a receptor will bind to several peptide sequences in an array, but will bind much more strongly to some sequences than others.
15 Strong binding affinity will be evidenced herein by a strong fluorescent or radiographic signal since many receptor molecules will bind in a region of a strongly bound ligand. Conversely, a weak binding affinity will be evidenced by a weak fluorescent or radiographic signal
20 due to the relatively small number of receptor molecules which bind in a particular region of a substrate having a ligand with a weak binding affinity for the receptor. Consequently, it becomes possible to determine relative binding avidity (or affinity in the case of univalent
25 interactions) of a ligand herein by way of the intensity of a fluorescent or radiographic signal in a region containing that ligand.

Semiquantitative data on affinities might also be obtained by varying washing conditions and
30 concentrations of the receptor. This would be done by comparison to known ligand receptor pairs, for example.

VII. Examples

The following examples are provided to
35 illustrate the efficacy of the inventions herein. All operations were conducted at about ambient temperatures and pressures unless indicated to the contrary.

A. Slide Preparation

Before attachment of reactive groups it is preferred to clean the substrate which is, in a preferred embodiment a glass substrate such as a microscope slide or cover slip. According to one embodiment the slide is soaked in an alkaline bath consisting of, for example, 1 liter of 95% ethanol with 120 ml of water and 120 grams of sodium hydroxide for 12 hours. The slides are then washed under running water and allowed to air dry, and rinsed once with a solution of 95% ethanol.

The slides are then aminated with, for example, aminopropyltriethoxysilane for the purpose of attaching amino groups to the glass surface on linker molecules, although any omega functionalized silane could also be used for this purpose. In one embodiment 0.1% aminopropyltriethoxysilane is utilized, although solutions with concentrations from $10^{-7}\%$ to 10% may be used, with about $10^{-3}\%$ to 2% preferred. A 0.1% mixture is prepared by adding to 100 ml of a 95% ethanol/5% water mixture, 100 microliters (μ l) of aminopropyltriethoxysilane. The mixture is agitated at about ambient temperature on a rotary shaker for about 5 minutes. 500 μ l of this mixture is then applied to the surface of one side of each cleaned slide. After 4 minutes, the slides are decanted of this solution and rinsed three times by dipping in, for example, 100% ethanol.

After the plates dry, they are placed in a 110-120°C vacuum oven for about 20 minutes, and then allowed to cure at room temperature for about 12 hours in an argon environment. The slides are then dipped into DMF (dimethylformamide) solution, followed by a thorough washing with methylene chloride.

The aminated surface of the slide is then exposed to about 500 μ l of, for example, a 30 millimolar (mM) solution of NVOC-GABA (gamma amino butyric acid) NHS (N-hydroxysuccinimide) in DMF for attachment of a NVOC-GABA to each of the amino groups.

The surface is washed with, for example, DMF, methylene chloride, and ethanol.

Any unreacted aminopropyl silane on the surface--that is, those amino groups which have not had the NVOC-GABA attached--are now capped with acetyl groups (to prevent further reaction) by exposure to a 1:3 mixture of acetic anhydride in pyridine for 1 hour. Other materials which may perform this residual capping function include trifluoroacetic anhydride, formicacetic anhydride, or other reactive acylating agents. Finally, the slides are washed again with DMF, methylene chloride, and ethanol.

B. Synthesis of Eight Trimers of "A" and "B"

Fig.14 illustrates a possible synthesis of the eight trimers of the two-monomer set: gly, phe (represented by "A" and "B," respectively). A glass slide bearing silane groups terminating in 6-nitro-veratryloxycarboxamide (NVOC-NH) residues is prepared as a substrate. Active esters (pentafluorophenyl, OBt, etc.) of gly and phe protected at the amino group with NVOC are prepared as reagents. While not pertinent to this example, if side chain protecting groups are required for the monomer set, these must not be photoreactive at the wavelength of light used to protect the primary chain.

For a monomer set of size n , $n \times \ell$ cycles are required to synthesize all possible sequences of length ℓ . A cycle consists of:

1. Irradiation through an appropriate mask to expose the amino groups at the sites where the next residue is to be added, with appropriate washes to remove the by-products of the deprotection.
2. Addition of a single activated and protected (with the same photochemically-removable group) monomer, which will react

only at the sites addressed in step 1, with appropriate washes to remove the excess reagent from the surface.

5 The above cycle is repeated for each member of the monomer set until each location on the surface has been extended by one residue in one embodiment. In other
10 embodiments, several residues are sequentially added at one location before moving on to the next location. Cycle times will generally be limited by the coupling reaction rate, now as short as 20 min in automated
15 peptide synthesizers. This step is optionally followed by addition of a protecting group to stabilize the array for later testing. For some types of polymers (e.g., peptides), a final deprotection of the entire surface (removal of photoprotective side chain groups) may be required.

20 More particularly, as shown in Fig. 14 A, the glass 20 is provided with regions 22, 24, 26, 28, 30, 32, 34, and 36. Regions 30, 32, 34, and 36 are masked, as shown in Fig. 14 B and the glass is irradiated and exposed to a reagent containing "A" (e.g., gly), with the resulting structure shown in Fig. 14 C. Thereafter, regions 22, 24, 26, and 28 are masked, the glass is irradiated (as shown in Fig. 14 D) and exposed to a
25 reagent containing "B" (e.g., phe), with the resulting structure shown in Fig. 14 E. The process proceeds, consecutively masking and exposing the sections as shown until the structure shown in Fig. 14 F is obtained. The glass is irradiated and the terminal groups are,
30 optionally, capped by acetylation. As shown, all possible trimers of gly/phe are obtained.

35 In this example, no side chain protective group removal is necessary. If it is desired, side chain deprotection may be accomplished by treatment with ethanedithiol and trifluoroacetic acid.

In general, the number of steps needed to obtain a particular polymer chain is defined by:

$$n \times \ell \quad (1)$$

where:

5 n = the number of monomers in the basis set of
monomers, and
 ℓ = the number of monomer units in a polymer
chain.

10 Conversely, the synthesized number of sequences
of length ℓ will be:

$$n^{\ell}. \quad (2)$$

15 Of course, greater diversity is obtained by
using masking strategies which will also include the
synthesis of polymers having a length of less than ℓ .
If, in the extreme case, all polymers having a length
less than or equal to ℓ are synthesized, the number of
polymers synthesized will be:

20
$$n^{\ell} + n^{\ell-1} + \dots + n^1. \quad (3)$$

25 The maximum number of lithographic steps needed
will generally be n for each "layer" of monomers, i.e.,
the total number of masks (and, therefore, the number of
lithographic steps) needed will be $n \times \ell$. The size of
the transparent mask regions will vary in accordance with
the area of the substrate available for synthesis and the
number of sequences to be formed. In general, the size
30 of the synthesis areas will be:

$$\text{size of synthesis areas} = (A)/(S)$$

where:

35 A is the total area available for synthesis;
and

S is the number of sequences desired in the area.

5 It will be appreciated by those of skill in the art that the above method could readily be used to simultaneously produce thousands or millions of oligomers on a substrate using the photolithographic techniques disclosed herein. Consequently, the method results in the ability to practically test large numbers of, for
10 example, di, tri, tetra, penta, hexa, hepta, octapeptides, dodecapeptides, or larger polypeptides (or correspondingly, polynucleotides).

The above example has illustrated the method by way of a manual example. It will of course be
15 appreciated that automated or semi-automated methods could be used. The substrate would be mounted in a flow cell for automated addition and removal of reagents, to minimize the volume of reagents needed, and to more carefully control reaction conditions. Successive masks
20 could be applied manually or automatically.

C. Synthesis of a Dimer of an Aminopropyl Group and a Fluorescent Group

In synthesizing the dimer of an aminopropyl
25 group and a fluorescent group, a functionalized durapore membrane was used as a substrate. The durapore membrane was a polyvinylidene difluoride with aminopropyl groups. The aminopropyl groups were protected with the DDZ group by reaction of the carbonyl chloride with the amino
30 groups, a reaction readily known to those of skill in the art. The surface bearing these groups was placed in a solution of THF and contacted with a mask bearing a checkerboard pattern of 1 mm opaque and transparent regions. The mask was exposed to ultraviolet light
35 having a wavelength down to at least about 280 nm for about 5 minutes at ambient temperature, although a wide range of exposure times and temperatures may be

appropriate in various embodiments of the invention. For example, in one embodiment, an exposure time of between about 1 and 5000 seconds may be used at process temperatures of between -70 and +50°C.

5 In one preferred embodiment, exposure times of between about 1 and 500 seconds at about ambient pressure are used. In some preferred embodiments, pressure above ambient is used to prevent evaporation.

10 The surface of the membrane was then washed for about 1 hour with a fluorescent label which included an active ester bound to a chelate of a lanthanide. Wash times will vary over a wide range of values from about a few minutes to a few hours. These materials fluoresce in the red and the green visible region. After the
15 reaction with the active ester in the fluorophore was complete, the locations in which the fluorophore was bound could be visualized by exposing them to ultraviolet light and observing the red and the green fluorescence. It was observed that the derivatized regions of the
20 substrate closely corresponded to the original pattern of the mask.

D. Demonstration of Signal Capability

25 Signal detection capability was demonstrated using a low-level standard fluorescent bead kit manufactured by Flow Cytometry Standarda and having model no. 824. This kit includes 5.8 μm diameter beads, each impregnated with a known number of fluorescein molecules.

30 One of the beads was placed in the illumination field on the scan stage as shown in Fig. 9 in a field of a laser spot which was initially shuttered. After being positioned in the illumination field, the photon detection equipment was turned on. The laser beam was unblocked and it interacted with the particle bead,
35 which then fluoresced. Fluorescence curves of beads impregnated with 7,000; 13,000; and 29,000 fluorescein molecules, are shown in Figs. 11A, 11B, and 11C

respectively. On each curve, traces for beads without fluorescein molecules are also shown. These experiments were performed with 488 nm excitation, with 100 μ W of laser power. The light was focused through a 40 power 0.75 NA objective.

The fluorescence intensity in all cases started off at a high value and then decreased exponentially. The fall-off in intensity is due to photobleaching of the fluorescein molecules. The traces of beads without fluorescein molecules are used for background subtraction. The difference in the initial exponential decay between labeled and nonlabeled beads is integrated to give the total number of photon counts, and this number is related to the number of molecules per bead. Therefore, it is possible to deduce the number of photons per fluorescein molecule that can be detected. For the curves illustrated in Fig. 11, this calculation indicates the radiation of about 40 to 50 photons per fluorescein molecule are detected.

E. Determination of the Number of Molecules Per Unit Area

Aminopropylated glass microscope slides prepared according to the methods discussed above were utilized in order to establish the density of labeling of the slides. The free amino termini of the slides were reacted with FITC (fluorescein isothiocyanate) which forms a covalent linkage with the amino group. The slide is then scanned to count the number of fluorescent photons generated in a region which, using the estimated 40-50 photons per fluorescent molecule, enables the calculation of the number of molecules which are on the surface per unit area.

A slide with aminopropyl silane on its surface was immersed in a 1 mM solution of FITC in DMF for 1 hour at about ambient temperature. After reaction, the slide was washed twice with DMF and then washed with

ethanol, water, and then ethanol again. It was then dried and stored in the dark until it was ready to be examined.

Through the use of curves similar to those shown in Fig. 15., and by integrating the fluorescent counts under the exponentially decaying signal, the number of free amino groups on the surface after derivitization was determined. It was determined that slides with labeling densities of 1 fluoroscein per $10^3 \times 10^3$ to $\sim 2 \times 2$ nm could be reproducibly made as the concentration of aminopropyltriethoxysilane varied from $10^{-5}\%$ to $10^{-1}\%$.

F. Removal of NVOC and Attachment of A Fluorescent Marker

NVOC-GABA groups were attached as described above. The entire surface of one slide was exposed to light so as to expose a free amino group at the end of the gamma amino butyric acid. This slide, and a duplicate which was not exposed, were then exposed to fluorescein isothiocyanate (FITC).

Fig. 16 A illustrates the slide which was not exposed to light, but which was exposed to FITC. The units of the x axis are time and the units of the y axis are counts. The trace contains a certain amount of background fluorescence. The duplicate slide was exposed to 350 nm broadband illumination for about 1 minute (12 mW/cm^2 , $\sim 350 \text{ nm}$ illumination), washed and reacted with FITC. The fluorescence curves for this slide are shown in Fig. 16 B. A large increase in the level of fluorescence is observed, which indicates photolysis has exposed a number of amino groups on the surface of the slides for attachment of a fluorescent marker.

G. Use of a Mask in Removal of NVOC

The next experiment was performed with a 0.1% aminopropylated slide. Light from a Hg-Xe arc lamp was imaged onto the substrate through a laser-ablated chrome-on-glass mask in direct contact with the substrate.

This slide was ~~illuminated~~ for approximately 5 minutes, with 12 mW of 350 nm broadband light and then reacted with the 1 mM FITC solution. It was put on the laser detection scanning stage and a graph was plotted as a two-dimensional representation of position color-coded for fluorescence intensity. The fluorescence intensity (in counts) as a function of location is given on the color scale to the right of Figure 17 A for a mask having 100x100 μm squares.

The experiment was repeated a number of times through various masks. The fluorescence pattern for a 50 μm mask is illustrated in Fig. 17 B, for a 20 μm mask in Fig. 17 C, and for a 10 μm mask in Fig. 17 D. The mask pattern is distinct down to at least about 10 μm squares using this lithographic technique.

H. Attachment of YGGFL and Subsequent Exposure to Herz Antibody and Goat Antimouse

In order to establish that receptors to a particular polypeptide sequence would bind to a surface-bound peptide and be detected, Leu enkephalin was coupled to the surface and recognized by an antibody. A slide was derivatized with 0.1% amino propyl-triethoxysilane and protected with NVOC. A 500 μm checkerboard mask was used to expose the slide in a flow cell using backside contact printing. The Leu enkephalin sequence (H_2N -tyrosine, glycine, glycine, phenylalanine, leucine- CO_2H , otherwise referred to herein as YGGFL) was attached via its carboxy end to the exposed amino groups on the surface of the slide. The peptide was added in DMF solution with the BOP/HOBT/DIEA coupling reagents and

recirculated through the flow cell for 2 hours at room temperature.

A first antibody, known as the Herz antibody, was applied to the surface of the slide for 45 minutes at 2 $\mu\text{g/ml}$ in a supercocktail (containing 1% BSA and 1% ovalbumin also in this case). A second antibody, goat anti-mouse fluorescein conjugate, was then added at 2 $\mu\text{g/ml}$ in the supercocktail buffer, and allowed to incubate for 2 hours.

The results of this experiment are provided in Fig. 18. Again, this figure illustrates fluorescence intensity as a function of position. The fluorescence scale is shown on the right, according to the color coding. This image was taken at 10 μm steps. This figure indicates that not only can deprotection be carried out in a well defined pattern, but also that (1) the method provides for successful coupling of peptides to the surface of the substrate, (2) the surface of a bound peptide is available for binding with an antibody, and (3) that the detection apparatus capabilities are sufficient to detect binding of a receptor.

I. Monomer-by-Monomer Formation of YGGFL and Subsequent Exposure to Labeled Antibody

Monomer-by-monomer synthesis of YGGFL and GGFL in alternate squares was performed on a slide in a checkerboard pattern and the resulting slide was exposed to the Herz antibody. This experiment and the results thereof are illustrated in Figs. 19 A, 19 B, 19 C, and 19 D.

In Fig. 19 A, a slide is shown which is derivatized with the aminopropyl group, protected in this case with t-BOC (t-butoxycarbonyl). The slide was treated with TFA to remove the t-BOC protecting group. E-aminocaproic acid, which was t-BOC protected at its amino group, was then coupled onto the aminopropyl groups. The aminocaproic acid serves as a spacer between the aminopropyl group and the peptide to be synthesized.

The amino end of the spacer was deprotected and coupled to NVOC-leucine. The entire slide was then illuminated with 12 mW of 325 nm broadband illumination. The slide was then coupled with NVOC-phenylalanine and washed. The entire slide was again illuminated, then coupled to NVOC-glycine and washed. The slide was again illuminated and coupled to NVOC-glycine to form the sequence shown in the last portion of Fig. 19 A.

As shown in Fig. 19 B, alternating regions of the slide were then illuminated using a projection print using a 500x500 μm checkerboard mask; thus, the amino group of glycine was exposed only in the lighted areas. When the next coupling chemistry step was carried out, NVOC-tyrosine was added, and it coupled only at those spots which had received illumination. The entire slide was then illuminated to remove all the NVOC groups, leaving a checkerboard of YGGFL in the lighted areas and in the other areas, GGFL. The Herz antibody (which recognizes the YGGFL, but not GGFL) was then added, followed by goat anti-mouse fluorescein conjugate.

The resulting fluorescence scan is shown in Fig. 19 C, and the color coding for the fluorescence intensity is again given on the right. Dark areas contain the tetrapeptide GGFL, which is not recognized by the Herz antibody (and thus there is no binding of the goat anti-mouse antibody with fluorescein conjugate), and in the red areas YGGFL is present. The YGGFL pentapeptide is recognized by the Herz antibody and, therefore, there is antibody in the lighted regions for the fluorescein-conjugated goat anti-mouse to recognize.

Similar patterns are shown for a 50 μm mask used in direct contact ("proximity print") with the substrate in Fig. 19 D. Note that the pattern is more distinct and the corners of the checkerboard pattern are touching when the mask is placed in direct contact with the substrate (which reflects the increase in resolution using this technique).

J. Monomer-by-Monomer Synthesis of YGGFL and PGGFL

A synthesis using a 50 μm checkerboard mask similar to that shown in Fig. 19 was conducted. However, P was added to the GGFL sites on the substrate through an additional coupling step. P was added by exposing protected GGFL to light through a mask, and subsequent exposure to P in the manner set forth above. Therefore, half of the regions on the substrate contained YGGFL and the remaining half contained PGGFL.

The fluorescence plot for this experiment is provided in Fig. 20. As shown, the regions are again readily discernable. This experiment demonstrates that antibodies are able to recognize a specific sequence and that the recognition is not length-dependent.

K. Monomer-by-Monomer Synthesis of YGGFL and YPGGFL

In order to further demonstrate the operability of the invention, a 50 μm checkerboard pattern of alternating YGGFL and YPGGFL was synthesized on a substrate using techniques like those set forth above. The resulting fluorescence plot is provided in Fig. 21. Again, it is seen that the antibody is clearly able to recognize the YGGFL sequence and does not bind significantly at the YPGGFL regions.

L. Synthesis of an Array of Sixteen Different Amino Acid Sequences and Estimation of Relative Binding Affinity to Herz Antibody

Using techniques similar to those set forth above, an array of 16 different amino acid sequences (replicated four times) was synthesized on each of two glass substrates. The sequences were synthesized by attaching the sequence NVOC-GFL across the entire surface of the slides. Using a series of masks, two layers of amino acids were then selectively applied

to the substrate. Each region had dimensions of 0.25 cm x 0.0625 cm. The first slide contained amino acid sequences containing only L amino acids while the second slide contained selected D amino acids. Figs. 18A and 18B illustrate a map of the various regions on the first and second slides, respectively. The patterns shown in Figs. 22A and 22 B were duplicated four times on each slide. The slides were then exposed to the Herz antibody and fluorescein-labeled goat anti-mouse.

Fig. 23 is a fluorescence plot of the first slide, which contained only L amino acids. Red indicates strong binding (149,000 counts or more) while black indicates little or no binding of the Herz antibody (20,000 counts or less). The bottom right-hand portion of the slide appears "cut off" because the slide was broken during processing. The sequence YGGFL is clearly most strongly recognized. The sequences YAGFL and YSGFL also exhibit strong recognition of the antibody. By contrast, most of the remaining sequences show little or no binding. The four duplicate portions of the slide are extremely consistent in the amount of binding shown therein.

Fig. 24 is a fluorescence plot of the second slide. Again, strongest binding is exhibited by the YGGFL sequence. Significant binding is also detected to YAGFL, YSGFL, and YpGFL. The remaining sequences show less binding with the antibody. Note the low binding efficiency of the sequence YGGFL.

Table 6 lists the various sequences tested in order of relative fluorescence, which provides information regarding relative binding affinity.

Table 6
Apparent Binding to Herz Ab

	<u>L-a.a. Set</u>	<u>D-a.a. Set</u>
5	YGGFL	YGGFL
	YAGFL	YaGFL
	YSGFL	YsGFL
	LGGFL	YpGFL
	FGGFL	fGGFL
10	YPGFL	yGGFL
	LAGFL	faGFL
	FAGFL	wGGFL
	WGGFL	yaGFL
		fpGFL
15		waGFL

VIII. Illustrative Alternative Embodiment

20 According to an alternative embodiment of the invention, the methods provide for attaching to the surface a caged binding member which in its caged form has a relatively low affinity for other potentially binding species, such as receptors and specific binding substances. Such techniques are more fully described in copending application Serial No. 404,920, filed September 8, 1989, and incorporated herein by reference for all purposes.

30 According to this alternative embodiment, the invention provides methods for forming predefined regions on a surface of a solid support, wherein the predefined regions are capable of immobilizing receptors. The methods make use of caged binding members attached to the surface to enable selective activation of the predefined regions. The caged binding members are liberated to act as binding members ultimately capable of binding receptors upon selective activation of the predefined

regions. The activated binding members are then used to immobilize specific molecules such as receptors on the predefined region of the surface. The above procedure is repeated at the same or different sites on the surface so as to provide a surface prepared with a plurality of regions on the surface containing, for example, the same or different receptors. When receptors immobilized in this way have a differential affinity for one or more ligands, screenings and assays for the ligands can be conducted in the regions of the surface containing the receptors.

The alternative embodiment may make use of novel caged binding members attached to the substrate. Caged (unactivated) members have a relatively low affinity for receptors of substances that specifically bind to uncaged binding members when compared with the corresponding affinities of activated binding members. Thus, the binding members are protected from reaction until a suitable source of energy is applied to the regions of the surface desired to be activated. Upon application of a suitable energy source, the caging groups labilize, thereby presenting the activated binding member. A typical energy source will be light.

Once the binding members on the surface are activated they may be attached to a receptor. The receptor chosen may be a monoclonal antibody, a nucleic acid sequence, a drug receptor, etc. The receptor will usually, though not always, be prepared so as to permit attaching it, directly or indirectly, to a binding member. For example, a specific binding substance having a strong binding affinity for the binding member and a strong affinity for the receptor or a conjugate of the receptor may be used to act as a bridge between binding members and receptors if desired. The method uses a receptor prepared such that the receptor retains its activity toward a particular ligand.

Preferably, the caged binding member attached to the solid substrate will be a photoactivatable biotin complex, i.e., a biotin molecule that has been chemically modified with photoactivatable protecting groups so that it has a significantly reduced binding affinity for avidin or avidin analogs than does natural biotin. In a preferred embodiment, the protecting groups localized in a predefined region of the surface will be removed upon application of a suitable source of radiation to give binding members, that are biotin or a functionally analogous compound having substantially the same binding affinity for avidin or avidin analogs as does biotin.

In another preferred embodiment, avidin or an avidin analog is incubated with activated binding members on the surface until the avidin binds strongly to the binding members. The avidin so immobilized on predefined regions of the surface can then be incubated with a desired receptor or conjugate of a desired receptor. The receptor will preferably be biotinylated, e.g., a biotinylated antibody, when avidin is immobilized on the predefined regions of the surface. Alternatively, a preferred embodiment will present an avidin/biotinylated receptor complex, which has been previously prepared, to activated binding members on the surface.

25

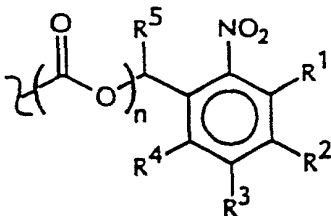
IX. Conclusion

The present inventions provide greatly improved methods and apparatus for synthesis of polymers on substrates. It is to be understood that the above description is intended to be illustrative and not restrictive. Many embodiments will be apparent to those of skill in the art upon reviewing the above description. By way of example, the invention has been described primarily with reference to the use of photoremovable protective groups, but it will be readily recognized by those of skill in the art that sources of radiation other than light could also be used. For example, in some

embodiments it may be desirable to use protective groups which are sensitive to electron beam irradiation, x-ray irradiation, in combination with electron beam lithograph, or x-ray lithography techniques.

- 5 Alternatively, the group could be removed by exposure to an electric current.

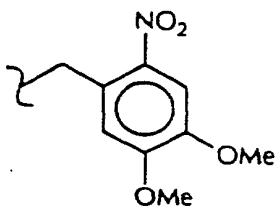
A preferred class of photoremovable protecting groups has the general formula:



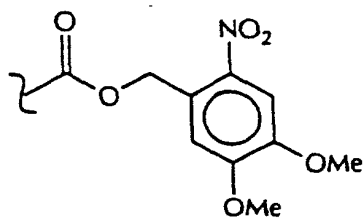
where R^1 , R^2 , R^3 , and R^4 independently are a hydrogen atom, a lower alkyl, aryl, benzyl, halogen, hydroxyl, alkoxyl, thiol, thioether, amino, nitro, carboxyl, formate, formamido or phosphido group, or adjacent substituents

(i.e., R^1-R^2 , R^2-R^3 , R^3-R^4) are substituted oxygen groups that together form a cyclic acetal or ketal; R^5 is a hydrogen atom, a alkoxyl, alkyl, hydrogen, halo, aryl, or alkenyl group, and $n = 0$ or 1 .

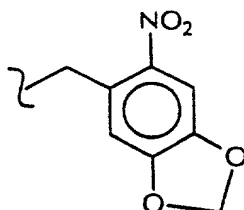
A preferred protecting group, 6-nitroveratryl (NV), which is used for protecting the carboxyl terminus of an amino acid or the hydroxyl group of a nucleotide, for example, is formed when R^2 and R^3 are each a methoxy group, R^1 , R^4 and R^5 are each a hydrogen atom, and $n = 0$:



A preferred protecting group, 6-nitroveratryloxycarbonyl (NVOC), which is used to protect the amino terminus of an amino acid, for example, is formed when R^2 and R^3 are each a methoxy group, R^1 , R^4 and R^5 are each a hydrogen atom, and $n = 1$:



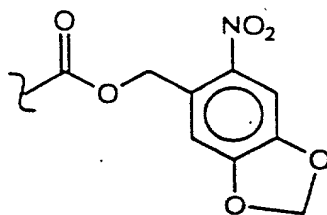
Another preferred protecting group,
 6-nitropiperonyl (NP), which is used for protecting the
 carboxyl terminus of an amino acid or the hydroxyl group
 of a nucleotide, for example, is formed when R^2 and R^3
 5 together form a methylene acetal, R^1 , R^4 and R^5 are each a
 hydrogen atom, and $n = 0$:



5

Another preferred protecting group,
6-nitropiperonyloxycarbonyl (NPOC), which is used to
protect the amino terminus of an amino acid, for example,
is formed when R^2 and R^3 together form a methylene acetal,
 R^1 , R^4 and R^5 are each a hydrogen atom, and $n = 1$:

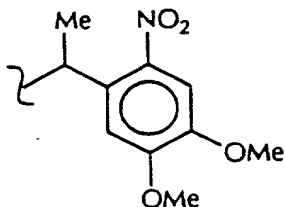
15



20

A most preferred protecting group,
methyl-6-nitroveratryl (MeNV), which is used for
protecting the carboxyl terminus of an amino acid or the
hydroxyl group of a nucleotide, for example, is formed
when R^2 and R^3 are each a methoxy group, R^1 and R^4 are
each a hydrogen atom, R^5 is a methyl group, and $n = 0$:

25

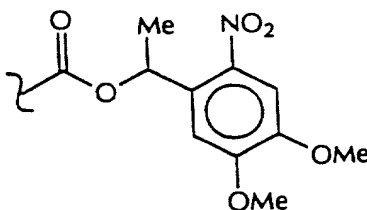


30

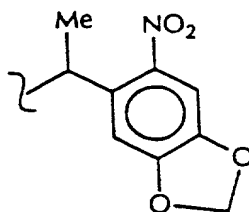
Another most preferred protecting group,
methyl-6-nitroveratryloxycarbonyl (MeNVOC), which is used
to protect the amino terminus of an amino acid, for
example, is formed when R^2 and R^3 are each a methoxy

35

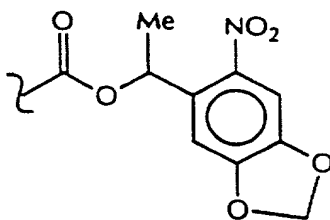
group, R^1 and R^4 are each a hydrogen atom, R^5 is a methyl group, and $n = 1$:



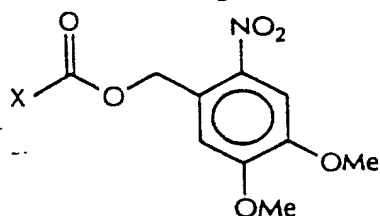
Another most preferred protecting group, methyl-6-nitropiperonyl (MeNP), which is used for protecting the carboxyl terminus of an amino acid or the hydroxyl group of a nucleotide, for example, is formed when R^2 and R^3 together form a methylene acetal, R^1 and R^4 are each a hydrogen atom, R^5 is a methyl group, and $n = 0$:



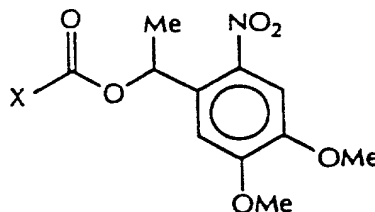
Another most preferred protecting group, methyl-6-nitropiperonyloxycarbonyl (MeNPOC), which is used to protect the amino terminus of an amino acid, for example, is formed when R^2 and R^3 together form a methylene acetal, R^1 and R^4 are each a hydrogen atom, R^5 is a methyl group, and $n = 1$:



A protected amino acid having a photoactivatable oxycarbonyl protecting group, such as NVOC or NPOC or their corresponding methyl derivatives, MeNVOC or MeNPOC, respectively, on the amino terminus is formed by acylating the amine of the amino acid with an activated oxycarbonyl ester of the protecting group. Examples of activated oxycarbonyl esters of NVOC and MeNVOC have the general formula:



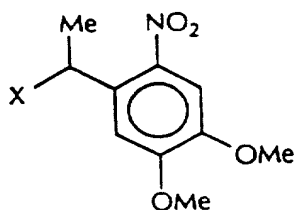
NVOC-X



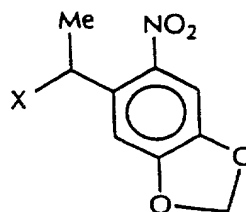
MeNVOC-X

where X is halogen, mixed anhydride, phenoxy, p-nitrophenoxy, N-hydroxysuccinimide, and the like.

A protected amino acid or nucleotide having a photoactivatable protecting group, such as NV or NP or their corresponding methyl derivatives, MeNV or MeNP, respectively, on the carboxy terminus of the amino acid or 5'-hydroxy terminus of the nucleotide, is formed by acylating the carboxy terminus or 5'-OH with an activated benzyl derivative of the protecting group. Examples of activated benzyl derivatives of MeNV and MeNP have the general formula:



MeNV-X



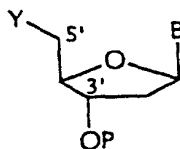
MeNP-X

where X is halogen, hydroxyl, tosyl, mesyl, trifluormethyl, diazo, azido, and the like.

Another method for generating protected monomers is to react the benzylic alcohol derivative of the protecting group with an activated ester of the

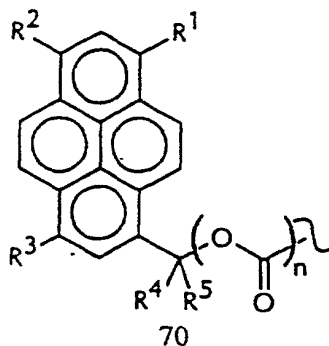
monomer. For example, to protect the carboxyl terminus of an amino acid, an activated ester of the amino acid is reacted with the alcohol derivative of the protecting group, such as 6-nitroveratrol (NVOH). Examples of activated esters suitable for such uses include halo-formate, mixed anhydride, imidazolyl formate, acyl halide, and also includes formation of the activated ester in situ the use of common reagents such as DCC and the like. See Atherton et al. for other examples of activated esters.

A further method for generating protected monomers is to react the benzylic alcohol derivative of the protecting group with an activated carbon of the monomer. For example, to protect the 5'-hydroxyl group of a nucleic acid, a derivative having a 5'-activated carbon is reacted with the alcohol derivative of the protecting group, such as methyl-6-nitropiperonol (MePyROH). Examples of nucleotides having activating groups attached to the 5'-hydroxyl group have the general formula:



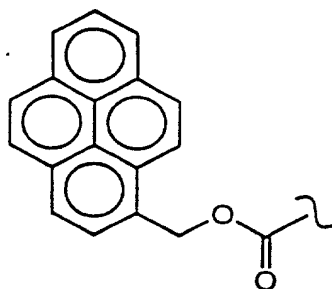
where Y is a halogen atom, a tosyl, mesyl, trifluoromethyl, azido, or diazo group, and the like.

Another class of preferred photochemical protecting groups has the formula:

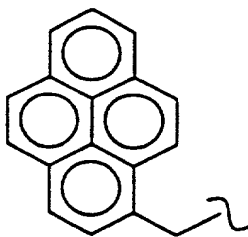


where R^1 , R^2 , and R^3 independently are a hydrogen atom, a lower alkyl, aryl, benzyl, halogen, hydroxyl, alkoxy, thiol, thioether, amino, nitro, carboxyl, formate, formamido, sulfanates, sulfido or phosphido group, R^4 and R^5 independently are a hydrogen atom, an alkoxy, alkyl, halo, aryl, hydrogen, or alkenyl group, and $n = 0$ or 1.

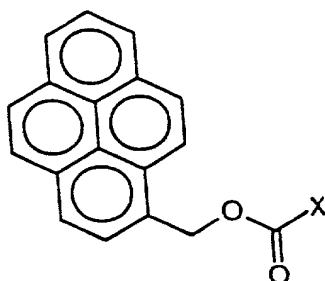
A preferred protecting group, 1-pyrenylmethyloxycarbonyl (PyROC), which is used to protect the amino terminus of an amino acid, for example, is formed when R^1 through R^5 are each a hydrogen atom and $n = 1$:



Another preferred protecting group, 1-pyrenylmethyl (PyR), which is used for protecting the carboxy terminus of an amino acid or the hydroxyl group of a nucleotide, for example, is formed when R^1 through R^5 are each a hydrogen atom and $n = 0$:

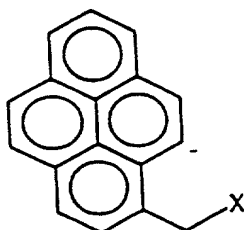


An amino acid having a pyrenylmethyloxycarbonyl protecting group on its amino terminus is formed by acylation of the free amine of amino acid with an activated oxycarbonyl ester of the pyrenyl protecting group. Examples of activated oxycarbonyl esters of PyROC have the general formula:



where X is halogen, or mixed anhydride, p-nitrophenoxy,
or N-hydroxysuccinimide group, and the like.

A protected amino acid or nucleotide having a photoactivatable protecting group, such as PyR, on the carboxy terminus of the amino acid or 5'-hydroxy terminus of the nucleic acid, respectively, is formed by acylating the carboxy terminus or 5'-OH with an activated pyrenylmethyl derivative of the protecting group. Examples of activated pyrenylmethyl derivatives of PyR have the general formula:



where X is a halogen atom, a hydroxyl, diazo, or azido group, and the like.

Another method of generating protected monomers is to react the pyrenylmethyl alcohol moiety of the protecting group with an activated ester of the monomer. For example, an activated ester of an amino acid can be reacted with the alcohol derivative of the protecting group, such as pyrenylmethyl alcohol (PyROH), to form the protected derivative of the carboxy terminus of the amino acid. Examples of activated esters include halo-formate, mixed anhydride, imidazolyl formate, acyl halide, and also

includes formation of the activated ester in situ and the use of common reagents such as DCC and the like.

Clearly, many photosensitive protecting groups are suitable for use in the present invention.

5 In preferred embodiments, the substrate is irradiated to remove the photoremovable protecting groups and create regions having free reactive moieties and side products resulting from the protecting group. The removal rate of the protecting groups depends on the
10 wavelength and intensity of the incident radiation, as well as the physical and chemical properties of the protecting group itself. Preferred protecting groups are removed at a faster rate and with a lower intensity of radiation. For example, at a given set of conditions,
15 MeNVOC and MeNPOC are photolytically removed from the N-terminus of a peptide chain faster than their unsubstituted parent compounds, NVOC and NPOC, respectively.

 Removal of the protecting group is accomplished
20 by irradiation to liberate the reactive group and degradation products derived from the protecting group. Not wishing to be bound by theory, it is believed that irradiation of an NVOC- and MeNVOC-protected oligomers occurs by the following reaction schemes:

25 NVOC-AA \rightarrow 3,4-dimethoxy-6-nitrosobenzaldehyde + CO₂ + AA
 MeNVOC-AA \rightarrow 3,4-dimethoxy-6-nitrosoacetophenone + CO₂ + AA

 where AA represents the N-terminus of the amino acid
30 oligomer.

 Along with the unprotected amino acid, other products are liberated into solution: carbon dioxide and a 2,3-dimethoxy-6-nitrosophenylcarbonyl compound, which
35 can react with nucleophilic portions of the oligomer to form unwanted secondary reactions. In the case of an NVOC-protected amino acid, the degradation product is a nitrosobenzaldehyde, while the degradation product for

the other is a nitrosophenyl ketone. For instance, it is believed that the product aldehyde from NVOC degradation reacts with free amines to form a Schiff base (imine) that affects the remaining polymer synthesis. Preferred photoremovable protecting groups react slowly or reversibly with the oligomer on the support.

Again not wishing to be bound by theory, it is believed that the product ketone from irradiation of a MeNVOC-protected oligomer reacts at a slower rate with nucleophiles on the oligomer than the product aldehyde from irradiation of the same NVOC-protected oligomer. Although not unambiguously determined, it is believed that this difference in reaction rate is due to the difference in general reactivity between aldehyde and ketones towards nucleophiles due to steric and electronic effects.

The photoremovable protecting groups of the present invention are readily removed. For example, the photolysis of N-protected L-phenylalanine in solution and having different photoremovable protecting groups was analyzed, and the results are presented in the following table:

Table
Photolysis of Protected L-Phe-OH

Solvent	<u>t_{1/2} in seconds</u>			
	NBOC	NVOC	MeNVOC	MeNPOC
Dioxane	1288	110	24	19
5mM H ₂ SO ₄ /Dioxane	1575	98	33	22

The half life, $t_{1/2}$, is the time in seconds required to remove 50% of the starting amount of protecting group. NBOC is the 6-nitrobenzyloxycarbonyl group, NVOC is the 6-nitroveratryloxycarbonyl group, MeNVOC is the methyl-6-nitroveratryloxycarbonyl group,

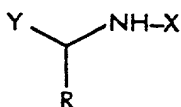
and MeNPOC is the methyl-6-nitropiperonyloxycarbonyl group. The photolysis was carried out in the indicated solvent with 362/364 nm-wavelength irradiation having an intensity of 10 mW/cm², and the concentration of each protected phenylalanine was 0.10 mM.

The table shows that deprotection of NVOC-, MeNVOC-, and MeNPOC-protected phenylalanine proceeded faster than the deprotection of NBOC. Furthermore, it shows that the deprotection of the two derivatives that are substituted on the benzylic carbon, MeNVOC and MeNPOC, were photolyzed at the highest rates in both dioxane and acidified dioxane.

1. Use of Photoremovable Groups During Solid-Phase Synthesis of Peptides

The formation of peptides on a solid-phase support requires the stepwise attachment of an amino acid to a substrate-bound growing chain. In order to prevent unwanted polymerization of the monomeric amino acid under the reaction conditions, protection of the amino terminus of the amino acid is required. After the monomer is coupled to the end of the peptide, the N-terminal protecting group is removed, and another amino acid is coupled to the chain. This cycle of coupling and deprotecting is continued for each amino acid in the peptide sequence. See Merrifield, J. Am. Chem. Soc. (1963) 85:2149, and Atherton et al., "Solid Phase Peptide Synthesis" 1989, IRL Press, London, both incorporated herein by reference for all purposes. As described above, the use of a photoremovable protecting group allows removal of selected portions of the substrate surface, via patterned irradiation, during the deprotection cycle of the solid phase synthesis. This selectively allows spatial control of the synthesis--the next amino acid is coupled only to the irradiated areas.

In one embodiment, the photoremovable protecting groups of the present invention are attached to an activated ester of an amino acid at the amino terminus:



where R is the side chain of a natural or unnatural amino acid, X is a photoremovable protecting group, and Y is an activated carboxylic acid derivative. The photoremovable protecting group, X, is preferably NVOC, NPOC, PyROC, MeNVOC, MeNPOC, and the like as discussed above. The activated ester, Y, is preferably a reactive derivative having a high coupling efficiency, such as an acyl halide, mixed anhydride, N-hydroxysuccinimide ester, perfluorophenyl ester, or urethane protected acid, and the like. Other activated esters and reaction conditions are well known (See Atherton et al.).

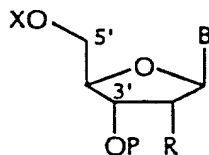
2. Use of Photoremovable Groups During Solid-Phase Synthesis of Oligonucleotides

The formation of oligonucleotides on a solid-phase support requires the stepwise attachment of a nucleotide to a substrate-bound growing oligomer. In order to prevent unwanted polymerization of the monomeric nucleotide under the reaction conditions, protection of the 5'-hydroxyl group of the nucleotide is required. After the monomer is coupled to the end of the oligomer, the 5'-hydroxyl protecting group is removed, and another nucleotide is coupled to the chain. This cycle of coupling and deprotecting is continued for each nucleotide in the oligomer sequence. See Gait, "Oligonucleotide Synthesis: A Practical Approach" 1984, IRL Press, London, incorporated herein by reference for

all purposes. As described above, the use of a photoremovable protecting group allows removal, via patterned irradiation, of selected portions of the substrate surface during the deprotection cycle of the solid phase synthesis. This selectively allows spatial control of the synthesis--the next nucleotide is coupled only to the irradiated areas.

Oligonucleotide synthesis generally involves coupling an activated phosphorous derivative on the 3'-hydroxyl group of a nucleotide with the 5'-hydroxyl group of an oligomer bound to a solid support. Two major chemical methods exist to perform this coupling: the phosphate-triester and phosphoamidite methods (See Gait). Protecting groups of the present invention are suitable for use in either method.

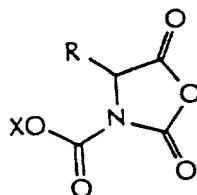
In a preferred embodiment, a photoremovable protecting group is attached to an activated nucleotide on the 5'-hydroxyl group:



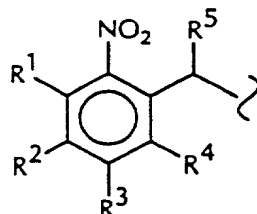
where B is the base attached to the sugar ring; R is a hydrogen atom when the sugar is deoxyribose or R is a hydroxyl group when the sugar is ribose; P represents an activated phosphorous group; and X is a photoremovable protecting group. The photoremovable protecting group, X, is preferably NV, NP, PyR, MeNV, MeNP, and the like as described above. The activated phosphorous group, P, is preferably a reactive derivative having a high coupling efficiency, such as a phosphate-triester, phosphoamidite or the like. Other activated phosphorous derivatives, as well as reaction conditions, are well known (See Gait).

E. Amino Acid N-Carboxy Anhydrides
Protected With a Photoremovable Group

During Merrifield peptide synthesis, an activated ester of one amino acid is coupled with the free amino terminus of a substrate-bound oligomer. Activated esters of amino acids suitable for the solid phase synthesis include halo-formate, mixed anhydride, imidazolyl formate, acyl halide, and also includes formation of the activated ester in situ and the use of common reagents such as DCC and the like (See Atherton et al.). A preferred protected and activated amino acid has the general formula:



where R is the side chain of the amino acid and X is a photoremovable protecting group. This compound is a urethane-protected amino acid having a photoremovable protecting group attach to the amine. A more preferred activated amino acid is formed when the photoremovable protecting group has the general formula:

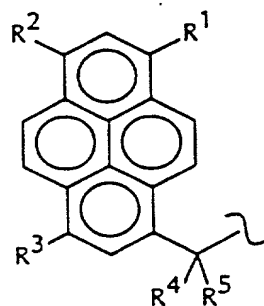


where R¹, R², R³, and R⁴ independently are a hydrogen atom, a lower alkyl, aryl, benzyl, halogen, hydroxyl, alkoxyl, thiol, thioether, amino, nitro, carboxyl, formate, formamido or phosphido group, or adjacent substituents (i.e., R¹-R², R²-R³, R³-R⁴) are substituted oxygen groups

that together form a cyclic acetal or ketal; and R⁵ is a hydrogen atom, a alkoxyl, alkyl, hydrogen, halo, aryl, or alkenyl group.

A preferred activated amino acid is formed when the photoremovable protecting group is 6-nitroveratryloxycarbonyl. That is, R¹ and R⁴ are each a hydrogen atom, R² and R³ are each a methoxy group, and R⁵ is a hydrogen atom. Another preferred activated amino acid is formed when the photoremovable group is 6-nitropiperonyl: R¹ and R⁴ are each a hydrogen atom, R² and R³ together form a methylene acetal, and R⁵ is a hydrogen atom. Other protecting groups are possible. Another preferred activated ester is formed when the photoremovable group is methyl-6-nitroveratryl or methyl-6-nitropiperonyl.

Another preferred activated amino acid is formed when the photoremovable protecting group has the general formula:



where R¹, R², and R³ independently are a hydrogen atom, a lower alkyl, aryl, benzyl, halogen, hydroxyl, alkoxyl, thiol, thioether, amino, nitro, carboxyl, formate, formamido, sulfanates, sulfido or phosphido group, and R⁴ and R⁵ independently are a hydrogen atom, an alkoxy, alkyl, halo, aryl, hydrogen, or alkenyl group. The resulting compound is a urethane-protected amino acid having a pyrenylmethyloxycarbonyl protecting group attached to the amine. A more preferred embodiment is formed when R¹ through R⁵ are each a hydrogen atom.

The urethane-protected amino acids having a photoremovable protecting group of the present invention

are prepared by condensation of an N-protected amino acid with an acylating agent such as an acyl halide, anhydride, chloroformate and the like (See Fuller et al., U.S. Patent No. 4,946,942 and Fuller et al., J. Amer. Chem. Soc. (1990) 112:7414-7416, both herein incorporated by reference for all purposes).

Urethane-protected amino acids having photoremovable protecting groups are generally useful as reagents during solid-phase peptide synthesis, and because of the spatially selectivity possible with the photoremovable protecting group, are especially useful for the spatially addressing peptide synthesis. These amino acids are difunctional: the urethane group first serves to activate the carboxy terminus for reaction with the amine bound to the surface and, once the peptide bond is formed, the photoremovable protecting group protects the newly formed amino terminus from further reaction. These amino acids are also highly reactive to nucleophiles, such as deprotected amines on the surface of the solid support, and due to this high reactivity, the solid-phase peptide coupling times are significantly reduced, and yields are typically higher.

1. Example

Light activated formation of a thymidine-cytidine dimer was carried out. A three dimensional representation of a fluorescence scan showing a checkboard pattern generated by the light-directed synthesis of a dinucleotide is shown in Fig. 8. 5'-nitroveratryl thymidine was attached to a synthesis substrate through the 3' hydroxyl group. The nitroveratryl protecting groups were removed by illumination through a 500 nm checkerboard mask. The substrate was then treated with phosphoramidite activated 2'-deoxycytidine. In order to follow the reaction fluorometrically, the deoxycytidine had been modified with an Fmoc protected aminohexyl linker attached to the exocyclic amine (5'-O-dimethoxytrityl-4-N-(6-N-fluorenylmethylcarbamoyl-hexylcarboxy)-2'-deoxycytidine). After removal of the Fmoc protecting group with base, the regions which contained the dinucleotide were fluorescently labelled by treatment of the substrate with 1 mM FITC in DMF for one hour.

The three-dimensional representation of the fluorescent intensity data in Fig. 14 clearly reproduces the checkerboard illumination pattern used during photolysis of the substrate. This result demonstrates that oligonucleotides as well as peptides can be synthesized by the light-directed method.

C. Binary Masking

In fact, the means for producing a substrate useful for these techniques are explained in U.S.S.N. 07/492,462 (VLSIPS CIP), which is hereby incorporated herein by reference.

5 However, there are various particular ways to optimize the synthetic processes. Many of these methods are described in U.S.S.N. ____/____,____, attorney docket number 11509-28 (automated VLSIPS).

10 Briefly, the binary synthesis strategy refers to an ordered strategy for parallel synthesis of diverse polymer sequences by sequential addition of reagents which may be represented by a reactant matrix, and a switch matrix, the product of which is a product matrix. A reactant matrix is a 1 x n matrix of the building blocks to be added. The switch
15 matrix is all or a subset of the binary numbers from 1 to n arranged in columns. In preferred embodiments, a binary strategy is one in which at least two successive steps illuminate half of a region of interest on the substrate. In most preferred embodiments, binary synthesis refers to a
20 synthesis strategy which also factors a previous addition step. For example, a strategy in which a switch matrix for a masking strategy halves regions that were previously illuminated, illuminating about half of the previously illuminated region and protecting the remaining half (while also protecting about
25 half of previously protected regions and illuminating about half of previously protected regions). It will be recognized that binary rounds may be interspersed with non-binary rounds and that only a portion of a substrate may be subjected to a binary scheme, but will still be considered to be a binary
30 masking scheme within the definition herein. A binary "masking" strategy is a binary synthesis which uses light to

remove protective groups from materials for addition of other materials such as nucleotides or amino acids.

In particular, this procedure provides a simplified and highly efficient method for saturating all possible
5 sequences of a defined length polymer. This masking strategy is also particularly useful in producing all possible oligonucleotide, sequence probes of a given length.

D. Applications

10 The technology provided by the present invention has very broad applications. Although described specifically for polynucleotide sequences, similar sequencing, fingerprinting, mapping, and screening procedures can be applied to
15 polypeptide, carbohydrate, or other polymers. In particular, the present invention may be used to completely sequence a given target sequence to subunit resolution. This may be for de novo sequencing, or may be used in conjunction with a second sequencing procedure to provide independent verification. See, e.g., (1988) Science 242:1245. For example, a large
20 polynucleotide sequence defined by either the Maxam and Gilbert technique or by the Sanger technique may be verified by using the present invention.

In addition, by selection of appropriate probes, a polynucleotide sequence can be fingerprinted. Fingerprinting
25 is a less detailed sequence analysis which usually involves the characterization of a sequence by a combination of defined features. Sequence fingerprinting is particularly useful because the repertoire of possible features which can be tested is virtually infinite. Moreover, the stringency of matching is
30 also variable depending upon the application. A Southern Blot analysis may be characterized as a means of simple fingerprint analysis.

Fingerprinting analysis may be performed to the resolution of specific nucleotides, or may be used to determine
35 homologies, most commonly for large segments. In particular, an array of oligonucleotide probes of virtually any workable size may be positionally localized on a matrix and used to probe a sequence for either absolute complementary matching, or

homology to the desired level of stringency using selected hybridization conditions.

In addition, the present invention provides means for mapping analysis of a target sequence or sequences. Mapping will usually involve the sequential ordering of a plurality of various sequences, or may involve the localization of a particular sequence within a plurality of sequences. This may be achieved by immobilizing particular large segments onto the matrix and probing with a shorter sequence to determine which of the large sequences contain that smaller sequence.

Alternatively, relatively shorter probes of known or random sequence may be immobilized to the matrix and a map of various different target sequences may be determined from overlaps. Principles of such an approach are described in some detail by Evans et al. (1989) "Physical Mapping of Complex Genomes by Cosmid Multiplex Analysis," Proc. Natl. Acad. Sci. USA 86:5030-5034; Michiels et al. (1987) "Molecular Approaches to Genome Analysis: A Strategy for the Construction of Ordered Overlap Clone Libraries," CABIOS 3:203-210; Olsen et al. (1986) "Random-Clone Strategy for Genomic Restriction Mapping in Yeast," Proc. Natl. Acad. Sci. USA 83:7826-7830; Craig, et al. (1990) "Ordering of Cosmid Clones Covering the Herpes Simplex Virus Type I (HSV-I) Genome: A Test Case for Fingerprinting by Hybridization," Nuc. Acids Res. 18:2653-2660; and Coulson, et al. (1986) "Toward a Physical Map of the Genome of the Nematode *Caenorhabditis elegans*," Proc. Natl. Acad. Sci. USA 83:7821-7825; each of which is hereby incorporated herein by reference.

Fingerprinting analysis also provides a means of identification. In addition to its value in apprehension of criminals from whom a biological sample, e.g., blood, has been collected, fingerprinting can ensure personal identification for other reasons. For example, it may be useful for identification of bodies in tragedies such as fire, flood, and vehicle crashes. In other cases the identification may be useful in identification of persons suffering from amnesia, or of missing persons. Other forensics applications include establishing the identity of a person, e.g., military identification "dog tags", or may be used in identifying the

source of particular biological samples. Fingerprinting technology is described, e.g., in Carrano, et al. (1989) "A High-Resolution, Fluorescence-Based, Semi-automated method for DNA Fingerprinting," Genomics 4: 129-136, which is hereby
5 incorporated herein by reference. See, e.g., table I, for nucleic acid applications, and corresponding applications may be accomplished using polypeptides.

TABLE I.

VLSIPS PROJECT IN NUCLEIC ACIDS

	I.	Construction of Chips
	II.	Applications
5	A.	Sequencing
	1.	Primary sequencing
	2.	Secondary sequencing (sequence checking)
	3.	Large scale mapping
	4.	Fingerprinting
10	B.	Duplex/Triplex formation
	1.	Antisense
	2.	Sequence specific function modulation (e.g. promoter inhibition)
	C.	Diagnosis
15	1.	Genetic markers
	2.	Type markers
	a.	Blood donors
	b.	Tissue transplants
	D.	Microbiology
20	1.	Clinical microbiology
	2.	Food microbiology
	III.	Instrumentation
	A.	Chip machines
25	B.	Detection
	IV.	Software Development
	A.	Instrumentation software
	B.	Data reduction software
30	C.	Sequence analysis software

appropriate. Design modifications may also be incorporated therein.

F. Data Analysis

5 Data is analyzed by processes similar to those described below in the section describing theoretical analysis. More efficient algorithms will be mathematically devised, and will usually be designed to be performed on a computer. Various computer programs which may more quickly or efficiently
10 make measurement samples and distinguish signal from noise will also be devised. See, particularly, U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS).

The initial data resulting from the detection system is an array of data indicative of fluorescent intensity versus
15 location on the substrate. The data are typically taken over regions substantially smaller than the area in which synthesis of a given polymer has taken place. Merely by way of example, if polymers were synthesized in squares on the substrate having dimensions of 500 microns by 500 microns, the data may be taken
20 over regions having dimensions of 5 microns by 5 microns. In most preferred embodiments, the regions over which fluorescence data are taken across the substrate are less than about 1/2 the area of the regions in which individual polymers are synthesized, preferably less than 1/10 the area in which a
25 single polymer is synthesized, and most preferably less than 1/100 the area in which a single polymer is synthesized. Hence, within any area in which a given polymer has been synthesized, a large number of fluorescence data points are collected.

30 A plot of number of pixels versus intensity for a scan should bear a rough resemblance to a bell curve, but spurious data are observed, particularly at higher intensities. Since it is desirable to use an average of fluorescent intensity over a given synthesis region in determining relative
35 binding affinity, these spurious data will tend to undesirably skew the data.

Accordingly, in one embodiment of the invention the data are corrected for removal of these spurious data points,

and an average of the data points is thereafter utilized in determining relative binding efficiency. In general the data are fitted to a base curve and statistically measures are used to remove spurious data.

5 In an additional analytical tool, various degeneracy reducing analogues may be incorporated in the hybridization probes. Various aspects of this strategy are described, e.g., in Macevitz, S. (1990) PCT publication number WO 90/04652, which is hereby incorporated herein by reference.

10 II. THEORETICAL ANALYSIS

The principle of the hybridization sequencing procedure is based, in part, upon the ability to determine overlaps of short segments. The VLSIPS technology provides the
15 ability to generate reagents which will saturate the possible short subsequence recognition possibilities. The principle is most easily illustrated by using a binary sequence, such as a sequence of zeros and ones. Once having illustrated the application to a binary alphabet, the principle may easily be
20 understood to encompass three letter, four letter, five or more letter, even 20 letter alphabets. A theoretical treatment of analysis of subsequence information to reconstruction of a target sequence is provided, e.e., in Lysov, Yu., et al. (1988) Doklady Akademi. Nauk. SSR 303:1508-1511; Khropko K., et al.
25 (1989) FEBS Letters 256:118-122; Pevzner, P. (1989) J. of Biomolecular Structure and Dynamics 7:63-69; and Drmanac, R. et al. (1989) Genomics 4:114-128; each of which is hereby incorporated herein by reference.

The reagents for recognizing the subsequences will
30 usually be specific for recognizing a particular polymer subsequence anywhere within a target polymer. It is preferable that conditions may be devised which allow absolute discrimination between high fidelity matching and very low levels of mismatching. The reagent interaction will preferably
35 exhibit no sensitivity to flanking sequences, to the subsequence position within the target, or to any other remote structure within the sequence. For polynucleotide sequencing, the specific reagents can be oligonucleotide probes; for

polypeptides and carbohydrates, antibodies will be useful reagents. Antibody reagents should also be useful for other types of polymers.

5 A. Simple n-mer Structure: Theory

 1. Simple two letter alphabet: example

 A simple example is presented below of how a sequence of ten digits comprising zeros and ones would be sequenceable using short segments of five digits. For example, consider the
10 sample ten digit sequence:

 1010011100.

A VLSIPS substrate could be constructed, as discussed elsewhere, which would have reagents attached in a defined matrix pattern which specifically recognize each of the
15 possible five digit sequences of ones and zeros. The number of possible five digit subsequences is $2^5 = 32$. The number of possible different sequences 10 digits long is $2^{10} = 1,024$. The five contiguous digit subsequences within a ten digit sequence number six, i.e., positioned at digits 1-5, 2-6, 3-7,
20 4-8, 5-9, and 6-10. It will be noted that the specific order of the digits in the sequence is important and that the order is directional, e.g., running left to right versus right to left. The first five digit sequence contained in the target sequence is 10100. The second is 01001, the third is 10011,
25 the fourth is 00111, the fifth is 01110, and the sixth is 11100.

 The VLSIPS substrate would have a matrix pattern of positionally attached reagents which recognize each of the different 5-mer subsequences. Those reagents which recognize
30 each of the 6 contained 5-mers will bind the target, and a label allows the positional determination of where the sequence specific interaction has occurred. By correlation of the position in the matrix pattern, the corresponding bound subsequences can be determined.

In the above-mentioned sequence, six different 5-mer sequences would be determined to be present. They would be:

5 10100
 01001
 10011
 00111
 01110
 11100

10 Any sequence which contains the first five digit sequence, 10100, already narrows the number of possible sequences (e.g., from 1024 possible sequences) which contain it to less than about 192 possible sequences.

15 This 192 is derived from the observation that with the subsequence 10100 at the far left of the sequence, in positions 1-5, there are only 32 possible sequences. Likewise, for that particular subsequence in positions 2-6, 3-7, 4-8, 5-9, and 6-10. So, to sum up all of the sequences that could contain 10100, there are 32 for each position and 6 positions for a total of about 192 possible sequences. However, some of
20 these 10 digit sequences will have been counted twice. Thus, by virtue of containing the 10100 subsequence, the number of possible 10-mer sequences has been decreased from 1024 sequences to less than about 192 sequences.

25 In this example, not only do we know that sequence contains 10100, but we also know that it contains the second five character sequence, 01001. By virtue of knowing that the sequence contains 10100, we can look specifically to determine whether the sequence contains a subsequence of five characters which contains the four leftmost digits plus a next digit to
30 the left. For example, we would look for a sequence of X1010, but we find that there is none. Thus, we know that the 10100 must be at the left end of the 10-mer. We would also look to see whether the sequence contains the rightmost four digits plus a next digit to the right, e.g., 0100X. We find that the
35 sequence also contains the sequence 01001, and that X is a 1. Thus, we know at least that our target sequence has an overlap of 0100 and has the left terminal sequence 101001.

40 Applying the same procedure to the second 5-mer, we also know that the sequence must include a sequence of five digits having the sequence 1001Y where Y must be either 0 or 1.

We look through the fragments and we see that we have a 10011 sequence within our target, thus Y is also 1. Thus, we would know that our sequence has a sequence of the first seven being 1010011.

5 Moving to the next 5-mer, we know that there must be a sequence of 0011Z, where Z must be either 0 or 1. We look at the fragments produced above and see that the target sequence contains a 00111 subsequence and Z is 1. Thus, we know the sequence must start with 10100111.

10 The next 5-mer must be of the sequence 0111W where W must be 0 or 1. Again, looking up at the fragments produced, we see that the target sequence contains a 01110 subsequence, and W is a 0. Thus, our sequence to this point is 101001110. We know that the last 5-mer must be either 11100 or 11101.

15 Looking above, we see that it is 11100 and that must be the last of our sequence. Thus, we have determined that our sequence must have been 1010011100.

 However, it will be recognized from the example above with the sequences provided therein, that the sequence analysis
20 can start with any known positive probe subsequence. The determination may be performed by moving linearly along the sequence checking the known sequence with a limited number of next positions. Given this possibility, the sequence may be determined, besides by scanning all possible oligonucleotide
25 probe positions, by specifically looking only where the next possible positions would be. This may increase the complexity of the scanning but may provide a longer time span dedicated towards scanning and detecting specific positions of interest relative to other sequence possibilities. Thus, the scanning
30 apparatus could be set up to work its way along a sequence from a given contained oligonucleotide to only look at those positions on the substrate which are expected to have a positive signal.

 It is seen that given a sequence, it can be de-
35 constructed into n-mers to produce a set of internal contiguous subsequences. From any given target sequence, we would be able to determine what fragments would result. The hybridization sequence method depends, in part, upon being able to work in

the reverse, from a set of fragments of known sequences to the full sequence. In simple cases, one is able to start at a single position and work in either or both directions towards the ends of the sequence as illustrated in the example.

5 The number of possible sequences of a given length increases very quickly with the length of that sequence. Thus, a 10-mer of zeros and ones has 1024 possibilities, a 12-mer has 4096. A 20-mer has over a million possibilities, and a 30-mer has over a billion. However, a given 30-mer has, at most, 26
10 different internal 5-mer sequences. Thus, a 30 character target sequence having over a million possible sequences can be substantially defined by only 26 different 5-mers. It will be recognized that the probe oligonucleotides will preferably, but need not necessarily, be of identical length, and that the
15 probe sequences need not necessarily be contiguous in that the overlapping subsequences need not differ by only a single subunit. Moreover, each position of the matrix pattern need not be homogeneous, but may actually contain a plurality of probes of known sequence. In addition, although all of the
20 possible subsequence specifications would be preferred, a less than full set of sequences specifications could be used. In particular, although a substantial fraction will preferably be at least about 70%, it may be less than that. About 20% would be preferred, more preferably at least about 30% would be
25 desired. Higher percentages would be especially preferred.

2. Example of four letter alphabet

A four letter alphabet may be conceptualized in at least two different ways from the two letter alphabet. One
30 way, is to consider the four possible values at each position and to analogize in a similar fashion to the binary example each of the overlaps. A second way is to group the binary digits into groups.

Using the first means, the overlap comparisons are
35 performed with a four letter alphabet rather than a two letter alphabet. Then, in contrast to the binary system with 10 positions where $2^{10} = 1024$ possible sequences, in a 4-character alphabet with 10 positions, there will actually be $4^{10} =$

representation of cytosine, G is the representation of guanine, and T is the representation of thymine or uracil.

3. Generalization to m-letter Alphabet

5 This reconstruction process may be applied to
polymers of virtually any number of possible characters in the
alphabet, and for virtually any length sequence to be
sequenced, though limitations, as discussed below, will limit
its efficiency at various extremes of length. It will be
10 recognized that the theory can be applied to a large diversity
of systems where sequence is important.

 For example, the method could be applied to
sequencing of a polypeptide. A polypeptide can have any of
twenty natural amino acid possibilities at each position. A
15 twenty letter alphabet is amenable to sequencing by this method
so long as reagents exist for recognizing shorter subsequences
therein. A preferred reagent for achieving that goal would be
a set of monoclonal antibodies each of which recognizes a
specific three contiguous amino acid subsequence. A complete
20 set of antibodies which recognize all possible subsequences of
a given length, e.g., 3 amino acids, and preferably with a
uniform affinity, would be $20^3 = 8000$ reagents.

 It will also be recognized that each target sequence
which is recognized by the specific reagents need not have
25 homogeneous termini. Thus, fragments of the entire target
sequence will also be useful for hybridizing appropriate
subsequences. It is, however, preferable that there not be a
significant amount of labeled homogeneous contaminating
extraneous sequences. This constraint does usually require the
30 purification of the target molecule to be sequenced, but a
specific label technique would dispense with a purification
requirement if the unlabeled extraneous sequences do not
interfere with the labeled sequences.

 In addition, conformational effects of target
35 polypeptide folding may, in certain embodiments, be negligible
if the polypeptide is fragmented into sufficiently small
peptides, or if the interaction is performed under conditions
where conformation, but not specific interaction, is disrupted.

B. Complications

Two obvious complications exist with the method of sequence analysis by hybridization. The first results from a probe of inappropriate length while the second relates to internally repeated sequences.

The first obvious complication is a problem which arises from an inappropriate length of recognition sequence, which causes problems with the specificity of recognition. For example, if the recognized sequence is too short, every sequence which is utilized will be recognized by every probe sequence. This occurs, e.g., in a binary system where the probes are each of sequences which occur relatively frequently, e.g., a two character probe for the binary system. Each possible two character probe would be expected to appear $\frac{1}{4}$ of the time in every single two character position. Thus, the above sequence example would be recognized by each of the 00, 10, 01, and 11. Thus, the sequence information is virtually lost because the resolution is too low and each recognition reagent specifically binds at multiple sites on the target sequence.

The number of different probes which bind to a target depends on the relationship between the probe length and the target length. At the extreme of short probe length, the just mentioned problem exists of excessive redundancy and lack of resolution. The lack of stability in recognition will also be a problem with extremely short probes. At the extreme of long probe length, each entire probe sequence is on a different position of a substrate. However, a problem arises from the number of possible sequences, which goes up dramatically with the length of the sequence. Also, the specificity of recognition begins to decrease as the contribution to binding by any particular subunit may become sufficiently low that the system fails to distinguish the fidelity of recognition. Mismatched hybridization may be a problem with the polynucleotide sequencing applications, though the fingerprinting and mapping applications may not be so strict in their fidelity requirements. As indicated above, a thirty

position binary sequence has over a million possible sequences, a number which starts to become unreasonably large in its required number of different sequences, even though the target length is still very short. Preparing a substrate with all
5 sequence-possibilities for a long target may be extremely difficult due to the many different oligomers which must be synthesized.

The above example illustrates how a long target sequence may be reconstructed with a reasonably small number of
10 shorter subsequences. Since the present day resolution of the regions of the substrate having defined oligomer probes attached to the substrate approaches about 10 microns by 10 microns for resolvable regions, about 10^6 , or 1 million, positions can be placed on a one centimeter square substrate.
15 However, high resolution systems may have particular disadvantages which may be outweighed using the lower density substrate matrix pattern. For this reason, a sufficiently large number of probe sequences can be utilized so that any given target sequence may be determined by hybridization to a
20 relatively small number of probes.

A second complication relates to convergence of sequences to a single subsequence. This will occur when a particular subsequence is repeated in the target sequence. This problem can be addressed in at least two different ways.
25 The first, and simpler way, is to separate the repeat sequences onto two different targets. Thus, each single target will not have the repeated sequence and can be analyzed to its end. This solution, however, complicates the analysis by requiring that some means for cutting at a site between the repeats can
30 be located. Typically a careful sequencer would want to have two intermediate cut points so that the intermediate region can also be sequenced in both directions across each of the cut points. This problem is inherent in the hybridization method for sequencing but can be minimized by using a longer known
35 probe sequence so that the frequency of probe repeats is decreased.

Knowing the sequence of flanking sequences of the repeat will simplify the use of polymerase chain reaction (PCR)

or a similar technique to further definitively determine the sequence between sequence repeats. Probes can be made to hybridize to those known sequences adjacent the repeat sequences, thereby producing new target sequences for analysis.

5 See, e.g., Innis et al. (eds.) (1990) PCR Protocols: A Guide to Methods and Applications, Academic Press; and methods for synthesis of oligonucleotide probes, see, e.g., Gait (1984) Oligonucleotide Synthesis: A Practical Approach, IRL Press, Oxford.

10 ... Other means for dealing with convergence problems include using particular longer probes, and using degeneracy reducing analogues, see, e.g., Macevicz, S. (1990) PCT publication number WO 90/04652, which is hereby incorporated herein by reference. By use of stretches of the degeneracy
15 reducing analogues with other probes in particular combinations, the number of probes necessary to fully saturate the possible oligomer probes is decreased. For example, with a stretch of 12-mers having the central 4-mer of degenerate nucleotides, in combination with all of the possible 8-mers,
20 the collection numbers twice the number of possible 8-mers, e.g. $65,536 + 65,536 = 131,072$, but the population provides screening equivalent to all possible 12-mers.

By way of further explanation, all possible oligonucleotide 8-mers may be depicted in the fashion:

25 N1-N2-N3-N4-N5-N6-N7-N8,

in which there are $4^8 = 65,536$ possible 8-mers. As described in U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS), producing all possible 8-mers requires $4 \times 8 = 32$ chemical binary synthesis steps to produce the entire
30 matrix pattern of 65,536 8-mer possibilities. By incorporating degeneracy reducing nucleotides, D's, which hybridize nonselectively to any corresponding complementary nucleotide, new oligonucleotides 12-mers can be made in the fashion:

N1-N2-N3-N4-D-D-D-D-N5-N6-N7-N8,

35 in which there are again, as above, only $4^8 = 65,536$ possible "12-mers", which in reality only have 8 different nucleotides.

However, it can be seen that each possible 12-mer probe could be represented by a group of the two 8-mer types.

Moreover, repeats of less than 12 nucleotides would not converge, or cause repeat problems in the analysis. Thus, instead of requiring a collection of probes corresponding to all 12-mers, or $4^{12} = 16,777,216$ different 12-mers, the same information can be derived by making 2 sets of "8-mers" consisting of the typical 8-mer collection of $4^8 = 65,536$ and the "12-mer" set with the degeneracy reducing analogues, also requiring making $4^8 = 65,536$. The combination of the two sets, requires making $65,536 + 65,536 = 131,072$ different molecules, but giving the information of 16,777,216 molecules. Thus, incorporating the degeneracy reducing analogue decreases the number of molecules necessary to get 12-mer resolution by a factor of about 128-fold.

C. Non-polynucleotide Embodiments

The above example is directed towards a polynucleotide embodiment. This application is relatively easily achieved because the specific reagents will typically be complementary oligonucleotides, although in certain embodiments other specific reagents may be desired. For example, there may be circumstances where other than complementary base pairing will be utilized. The polynucleotide targets, will usually be single strand, but may be double or triple stranded in various applications. However, a triple stranded specific interaction might be sometimes desired, or a protein or other specific binding molecule may be utilized. For example, various promoter or DNA sequence specific binding proteins might be used, including, e.g., restriction enzyme binding domains, other binding domains, and antibodies. Thus, specific recognition reagents besides oligonucleotides may be utilized.

For other polymer targets, the specific reagents will often be polypeptides. These polypeptides may be protein binding domains from enzymes or other proteins which display specificity for binding. Usually an antibody molecule may be used, and monoclonal antibodies may be particularly desired. Classical methods may be applied for preparing antibodies, see, e.g., Harlow and Lane (1988) Antibodies: A Laboratory Manual Cold Spring Harbor Press, New York; and Goding (1986)

Monoclonal Antibodies: Principles and Practice (2d Ed.)

Academic Press, San Diego. Other suitable techniques for in vitro exposure of lymphocytes to the antigens or selection of libraries of antibody binding sites are described, e.g., in

- 5 Huse et al. (1989) Science 246:1275-1281; and Ward et al. 91989) Nature 341:544-546, each of which is hereby incorporated herein by reference. Unusual antibody production methods are also described, e.g., in Hendricks et al. (1989) BioTechnology 7:1271-1274; and Hiatt et al. (1989) Nature 342:76-78, each of
10 which is hereby incorporated herein by reference. Other molecules which may exhibit specific binding interaction may be useful for attachment to a VLSIPS substrate by various methods, including the caged biotin methods, see, e.g., U.S.S.N. 07/435,316 (caged biotin parent), and U.S.S.N. 07/612,671
15 (caged biotin CIP).

- The antibody specific reagents should be particularly useful for the polypeptide, carbohydrate, and synthetic polymer applications. Individual specific reagents might be generated by an automated process to generate the number of reagents
20 necessary to advantageously use the high density positional matrix pattern. In an alternative approach, a plurality of hybridoma cells may be screened for their ability to bind to a VLSIPS matrix possessing the desired sequences whose binding specificity is desired. Each cell might be individually grown
25 up and its binding specificity determined by VLSIPS apparatus and technology. An alternative strategy would be to expose the same VLSIPS matrix to a polyclonal serum of high titer. By a successively large volume of serum and different animals, each region of the VLSIPS substrate would have attached to it a
30 substantial number of antibody molecules with specificity of binding. The substrate, with non-covalently bound antibodies could be derivatized and the antibodies transferred to an adjacent second substrate in the matrix pattern in which the antibody molecules had attached to the first matrix. If the
35 sensitivity of detection of binding interaction is sufficiently high, such a low efficiency transfer of antibody molecules may produce a sufficiently high signal to be useful for many purposes, including the sequencing applications.

possibility of attaching each and every one of these 10-mers to a separate specific position on a solid matrix. This attachment could be automated in any of a number of ways, particularly use of a caged biotin type linking. This would
5 produce a matrix having each of different possible 10-mers.

A batchwise hybridization is much preferred because of its reproducibility and simplicity. An automated process of attaching various reagents to positionally defined sites on a substrate is provided in U.S.S.N. 07/492,462 (VLSIPS CIP);

10 U.S.S.N. ___/___,___, attorney docket number 11509-28 (automated VLSIPS); and U.S.S.N. 07/612,671 (caged biotin CIP); each of which is hereby incorporated herein by reference.

Instead of separate synthesis of each oligonucleotide, these oligonucleotides are conveniently
15 synthesized in parallel by sequential synthetic processes on a defined matrix pattern as provided in U.S.S.N. 07/492,462 (VLSIPS CIP); and U.S.S.N. ___/___,___, attorney docket number 11509-28 (automated VLSIPS), which are incorporated herein by reference. Here, the oligonucleotides are synthesized stepwise
20 on a substrate at positionally separate and defined positions. Use of photosensitive blocking reagents allows for defined sequences of synthetic steps over the surface of a matrix pattern. By use of the binary masking strategy, the surface of the substrate can be positioned to generate a desired pattern
25 of regions, each having a defined sequence oligonucleotide synthesized and immobilized thereto.

Although the prior art technology can be used to generate the desired repertoire of oligonucleotide probes, an efficient and cost effective means would be to use the VLSIPS
30 technology described in U.S.S.N. 07/492,462 (VLSIPS CIP) and U.S.S.N. ___/___,___, attorney docket number 11509-28 (automated VLSIPS). In this embodiment, the photosensitive reagents involved in the production of such a matrix are described below.

35 The regions for synthesis may be very small, usually less than about 100 μm x 100 μm , more usually less than about 50 μm x 50 μm . The photolithography technology allows synthetic regions of less than about 10 μm x 10 μm , about

3 $\mu\text{m} \times 3 \mu\text{m}$, or less. The detection also may detect such sized regions, though larger areas are more easily and reliably measured.

At a size of about 30 microns by 30 microns, one million regions would take about 11 centimeters square or a single wafer of about 4 centimeters by 4 centimeters. Thus the present technology provides for making a single matrix of that size having all one million plus possible oligonucleotides. Region size are sufficiently small to correspond to densities of at least about 5 regions/cm², 20 regions/cm², 50 regions/cm², 100 regions/cm², and greater, including 300 regions/cm², 1000 regions/cm², 3K regions/cm², 10K regions/cm², 30K regions/cm², 100K regions/cm², 300K regions/cm² or more, even in excess of one million regions/cm².

Although the pattern of the regions which contain specific sequences is theoretically not important, for practical reasons certain patterns will be preferred in synthesizing the oligonucleotides. The application of binary masking algorithms for generating the pattern of known oligonucleotide probes is described in related U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS) which was filed simultaneously with this application. By use of these binary masks, a highly efficient means is provided for producing the substrate with the desired matrix pattern of different sequences. Although the binary masking strategy allows for the synthesis of all lengths of polymers, the strategy may be easily modified to provide only polymers of a given length. This is achieved by omitting steps where a subunit is not attached.

The strategy for generating a specific pattern may take any of a number of different approaches. These approaches are well described in related application U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS) and include a number of binary masking approaches which will not be exhaustively discussed herein. However, the binary masking and binary synthesis approaches provide a maximum of diversity with a minimum number of actual synthetic steps.

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The length of oligonucleotides used in sequencing applications will be selected on criteria determined to some extent by the practical limits discussed above. For example, if probes are made as oligonucleotides, there will be 65,536 possible eight nucleotide sequences. If a nine subunit oligonucleotide is selected, there are 262,144 possible permutations of sequences. If a ten-mer oligonucleotide is selected, there are 1,048,576 possible permutations of sequences. As the number gets larger, the required number of positionally defined subunits necessary to saturate the possibilities also increases. With respect to hybridization conditions, the length of the matching necessary to converse stability of the conditions selected can be compensated for. See, e.g., Kanehisa, M. (1984) Nuc. Acids Res. 12:203-213, which is hereby incorporated herein by reference.

Although not described in detail here, but below for oligonucleotide probes, the VLSIPS technology would typically use a photosensitive protective group on an oligonucleotide. Sample oligonucleotides are shown in Figure 1. In particular, the photoprotective group on the nucleotide molecules may be selected from a wide variety of positive light reactive groups preferably including nitro aromatic compounds such as o-nitrobenzyl derivatives or benzylsulfonyl. See, e.g., Gait (1984) Oligonucleotide Synthesis: A Practical Approach, IRL Press, Oxford, which is hereby incorporated herein by reference. In a preferred embodiment, 6-nitro-veratryl oxycarbony (NVOC), 2-nitrobenzyl oxycarbonyl (NBOC), or α,α -dimethyl-dimethoxybenzyl oxycarbonyl (DEZ) is used. Photoremovable protective groups are described in, e.g., Patchornik (1970) J. Amer. Chem. Soc. 92:6333-____; and Amit et al. (1974) J. Organic Chem. 39:192-____; each of which is hereby incorporated herein by reference.

A preferred linker for attaching the oligonucleotide to a silicon matrix is illustrated in Figure 2. A more detailed description is provided below. A photosensitive blocked nucleotide may be attached to specific locations of unblocked prior cycles of attachments on the substrate and can be successively built up to the correct length oligonucleotide probe.

It should be noted that multiple substrates may be simultaneously exposed to a single target sequence where each substrate is a duplicate of one another or where, in combination, multiple substrates together provide the complete or desired subset of possible subsequences. This provides the opportunity to overcome a limitation of the density of positions on a single substrate by using multiple substrates. In the extreme case, each probe might be attached to a single bead or substrate and the beads sorted by whether there is a binding interaction. Those beads which do bind might be encoded to indicate the subsequence specificity of reagents attached thereto.

Then, the target may be bound to the whole collection of beads and those beads that have appropriate specific reagents on them will bind to target. Then a sorting system may be utilized to sort those beads that actually bind the target from those that do not. This may be accomplished by presently available cell sorting devices or a similar apparatus. After the relatively small number of beads which have bound the target have been collected, the encoding scheme may be read off to determine the specificity of the reagent on the bead. An encoding system may include a magnetic system, a shape encoding system, a color encoding system, or a combination of any of these, or any other encoding system. Once again, with the collection of specific interactions that have occurred, the binding may be analyzed for sequence information, fingerprint information, or mapping information.

The parameters of polynucleotide sizes of both the probes and target sequences are determined by the applications and other circumstances. The length of the oligonucleotide probes used will depend in part upon the limitations of the VLSIPS technology to provide the number of desired probes. For example, in an absolute sequencing application, it is often useful to have virtually all of the possible oligonucleotides of a given length. As indicated above, there are 65,536 8-mers, 262,144 9-mers, 1,048,576 10-mers, 4,194,304 11-mers, etc. As the length of the oligomer increases the number of different probes which must be synthesized also increases at a

rate of a factor of 4 for every additional nucleotide.

Eventually the size of the matrix and the limitations in the resolution of regions in the matrix will reach the point where an increase in number of probes becomes disadvantageous.

5 However, this sequencing procedure requires that the system be able to distinguish, by appropriate selection of hybridization and washing conditions, between binding of absolute fidelity and binding of complementary sequences containing mismatches. On the other hand, if the fidelity is unnecessary, this
10 discrimination is also unnecessary and a significantly longer probe may be used. Significantly longer probes would typically be useful in fingerprinting or mapping applications.

The length of the probe is selected for a length that it will bind with specificity to possible targets. The
15 hybridization conditions are also very important in that they will determine how close the homology of complementary binding will be detected. In fact, a single target may be evaluated at a number of different conditions to determine its spectrum of specificity for binding particular probes. This may find use
20 in a number of other applications besides the polynucleotide sequencing fingerprinting or mapping. For example, it will be desired to determine the spectrum of binding affinities and specificities of cell surface antigens with binding by particular antibodies immobilized on the substrate surface,
25 particularly under different interaction conditions. In a related fashion, different regions with reagents having differing affinities or levels of specificity may allow such a spectrum to be defined using a single incubation, where various regions, at a given hybridization condition, show the binding
30 affinity. For example, fingerprint probes of various lengths, or with specific defined non-matches may be used. Unnatural nucleotides or nucleotides exhibiting modified specificity of complementary binding are described in greater detail in Macevicz (1990) PCT pub. No. WO 90/04652; and see the section
35 on modified nucleotides in the Sigma Chemical Company catalogue.

B. Labeling Target Nucleotide

The label used to detect the target sequences will be determined, in part, by the detection methods being applied. Thus, the labeling method and label used are selected in combination with the actual detecting systems being used.

Once a particular label has been selected, appropriate labeling protocols will be applied, as described below for specific embodiments. Standard labeling protocols for nucleic acids are described, e.g., in Sambrook et al.;

Kambara, H. et al. (1988) BioTechnology 6:816-821; Smith, L. et al. (1985) Nuc. Acids Res. 13:2399-2412; for polypeptides, see, e.g., Allen G. (1989) Sequencing of Proteins and Peptides, Elsevier, New York, especially chapter 5, and Greenstein and Winitz (1961) Chemistry of the Amino Acids, Wiley and Sons, New York. Carbohydrate labeling is described, e.g., in Chaplin and Kennedy (1986) Carbohydrate Analysis: A Practical Approach, IRL Press, Oxford. Labeling of other polymers will be performed by methods applicable to them as recognized by a person having ordinary skill in manipulating the corresponding polymer.

In some embodiments, the target need not actually be labeled if a means for detecting where interaction takes place is available. As described below, for a nucleic acid embodiment, such may be provided by an intercalating dye which intercalates only into double stranded segments, e.g., where interaction occurs. See, e.g., Sheldon et al. U.S. Pat. No. 4,582,789.

In many uses, the target sequence will be absolutely homogeneous, both with respect to the total sequence and with respect to the ends of each molecule. Homogeneity with respect to sequence is important to avoid ambiguity. It is preferable that the target sequences of interest not be contaminated with a significant amount of labeled contaminating sequences. The extent of allowable contamination will depend on the sensitivity of the detection system and the inherent signal to noise of the system. Homogeneous contamination sequences will be particularly disruptive of the sequencing procedure.

However, although the target polynucleotide must have a unique sequence, the target molecules need not have identical ends. In fact, the homogeneous target molecule preparation may be randomly sheared to increase the numerical number of molecules. Since the total information content remains the same, the shearing results only in a higher number of distinct sequences which may be labeled and bind to the probe. This fragmentation may give a vastly superior signal relative to a preparation of the target molecules having homogeneous ends. The signal for the hybridization is likely to be dependent on the numerical frequency of the target-probe interactions. If a sequence is individually found on a larger number of separate molecules a better signal will result. In fact, shearing a homogeneous preparation of the target may often be preferred before the labeling procedure is performed, thereby producing a large number of labeling groups associated with each subsequence.

C. Hybridization Conditions

The hybridization conditions between probe and target should be selected such that the specific recognition interaction, i.e., hybridization, of the two molecules is both sufficiently specific and sufficiently stable. See, e.g., Hames and Higgins (1985) Nucleic Acid Hybridisation: A Practical Approach, IRL Press, Oxford. These conditions will be dependent both on the specific sequence and often on the guanine and cytosine (GC) content of the complementary hybrid strands. The conditions may often be selected to be universally equally stable independent of the specific sequences involved. This typically will make use of a reagent such as an arylammonium buffer. See, Wood et al. (1985) "Base Composition-independent Hybridization in Tetramethylammonium Chloride: A Method for Oligonucleotide Screening of Highly Complex Gene Libraries," Proc. Natl. Acad. Sci. USA, 82:1585-1588; and Krupov et al. (1989) "An Oligonucleotide Hybridization Approach to DNA Sequencing," FEBS Letters, 256:118-122; each of which is hereby incorporated herein by reference. An arylammonium buffer tends to minimize

differences in hybridization rate and stability due to GC content. By virtue of the fact that sequences then hybridize with approximately equal affinity and stability, there is relatively little bias in strength or kinetics of binding for particular sequences. Temperature and salt conditions along with other buffer parameters should be selected such that the kinetics of renaturation should be essentially independent of the specific target subsequence or oligonucleotide probe involved. In order to ensure this, the hybridization reactions will usually be performed in a single incubation of all the substrate matrices together exposed to the identical same target probe solution under the same conditions.

Alternatively, various substrates may be individually treated differently. Different substrates may be produced, each having reagents which bind to target subsequences with substantially identical stabilities and kinetics of hybridization. For example, all of the high GC content probes could be synthesized on a single substrate which is treated accordingly. In this embodiment, the arylammonium buffers could be unnecessary. Each substrate is then treated in a manner that the collection of substrates show essentially uniform binding and the hybridization data of target binding to the individual substrate matrix is combined with the data from other substrates to derive the necessary subsequence binding information. The hybridization conditions will usually be selected to be sufficiently specific that the fidelity of base matching will be properly discriminated. Of course, control hybridizations should be included to determine the stringency and kinetics of hybridization.

D. Detection; VLSIPS Scanning

The next step of the sequencing process by hybridization involves labeling of target polynucleotide molecules. A quickly and easily detectable signal is preferred. The VLSIPS apparatus is designed to easily detect a fluorescent label, so fluorescent tagging of the target sequence is preferred. Other suitable labels include heavy metal labels, magnetic probes, chromogenic labels (e.g.,

phosphorescent labels, dyes, and fluorophores) spectroscopic labels, enzyme linked labels, radioactive labels, and labeled binding proteins. Additional labels are described in U.S. Pat. No. 4,366,241, which is incorporated herein by reference.

5 The detection methods used to determine where hybridization has taken place will typically depend upon the label selected above. Thus, for a fluorescent label a fluorescent detection step will typically be used. U.S.S.N. 07/492,462 (VLSIPS CIP) and U.S.S.N. __/__, __, attorney
10 docket number 11509-28 (automated VLSIPS) describe apparatus and mechanisms for scanning a substrate matrix using fluorescence detection, but a similar apparatus is adaptable for other optically detectable labels.

 The detection method provides a positional
15 localization of the region where hybridization has taken place. However, the position is correlated with the specific sequence of the probe since the probe has specifically been attached or synthesized at a defined substrate matrix position. Having collected all of the data indicating the subsequences present
20 in the target sequence, this data may be aligned by overlap to reconstruct the entire sequence of the target, as illustrated above.

 It is also possible to dispense with actual labeling if some means for detecting the positions of interaction
25 between the sequence specific reagent and the target molecule are available. This may take the form of an additional reagent which can indicate the sites either of interaction, or the sites of lack of interaction, e.g., a negative label. For the nucleic acid embodiments, locations of double strand
30 interaction may be detected by the incorporation of intercalating dyes, or other reagents such as antibody or other reagents that recognize helix formation, see, e.g., Sheldon, et al. (1986) U.S. Pat. No. 4,582,789, which is hereby incorporated herein by reference.

35 E. Analysis

 Although the reconstruction can be performed manually as illustrated above, a computer program will typically be used

to perform the overlap analysis. A program may be written and run on any of a large number of different computer hardware systems. The variety of operating systems and languages useable will be recognized by a computer software engineer.

5 Various different languages may be used, e.g., BASIC; C; PASCAL; etc. A simple flow chart of data analysis is illustrated in Figure 4.

F. Substrate Reuse

10 Finally, after a particular sequence has been hybridized and the pattern of hybridization analyzed, the matrix substrate should be reusable and readily prepared for exposure to a second or subsequent target polynucleotides. In order to do so, the hybrid duplexes are disrupted and the
15 matrix treated in a way which removes all traces of the original target. The matrix may be treated with various detergents or solvents to which the substrate, the oligonucleotide probes, and the linkages to the substrate are inert. This treatment may include an elevated temperature
20 treatment, treatment with organic or inorganic solvents, modifications in pH, and other means for disrupting specific interaction. Thereafter, a second target may actually be applied to the recycled matrix and analyzed as before.

25 G. Non-Polynucleotide Aspects

Although the sequencing, fingerprinting, and mapping functions will make use of the natural sequence recognition property of complementary nucleotide sequences, the non-polynucleotide sequences typically require other sequence
30 recognition reagents. These reagents will take the form, typically, of proteins exhibiting binding specificity, e.g., enzyme binding sites or antibody binding sites.

Enzyme binding sites may be derived from promoter proteins, restriction enzymes, and the like. See, e.g.,
35 Stryer, L. (1988) Biochemistry, W.H. Freeman, Palo Alto. Antibodies will typically be produced using standard procedures, see, e.g., Harlow and Lane (1988) Antibodies: A Laboratory Manual, Cold Spring Harbor Press, New York; and

Goding (1986) Monoclonal Antibodies: Principles and Practice, (2d Ed.) Academic Press, San Diego.

Typically, an antigen, or collection of antigens are presented to an immune system. This may take the form of synthesized short polymers produced by the VLSIPS technology, or by the other synthetic means, or from isolation of natural products. For example, antigen for the polypeptides may be made by the VLSIPS technology, by standard peptide synthesis, by isolation of natural proteins with or without degradation to shorter segments, or by expression of a collection of short nucleic acids of random or defined sequences. See, e.g., Tuerk and Gold (1990) Science 249:505-510, for generation of a collection of randomly mutagenized oligonucleotides useful for expression.

The antigen or collection is presented to an appropriate immune system, e.g., to a whole animal as in a standard immunization protocol, or to a collection of immune cells or equivalent. In particular, see Ward et al. (1989) Nature 341:544-546; and Huse et al. (1989) Science 246:1275-1281, each of which is hereby incorporated herein by reference.

A large diversity of antibodies will be generated, some of which have specificities for the desired sequences. Antibodies may be purified having the desired sequence specificities by isolating the cells producing them. For example, a VLSIPS substrate with the desired antigens synthesized thereon may be used to isolate cells with cell surface reagents which recognize the antigens. The VLSIPS substrate may be used as an affinity reagent to select and recover the appropriate cells. Antibodies from those cells may be attached to a substrate using the caged biotin methodology, or by attaching a targeting molecule, e.g., an oligonucleotide. Alternatively, the supernatants from antibody producing cells can be easily assayed using a VLSIPS substrate to identify the cells producing the appropriate antibodies.

Although cells may be isolated, specific antibody molecules which perform the sequence recognition will also be sufficient. Preferably populations of antibody with a known specificity can be isolated. Supernatants from a large

population of producing cells may be passed over a VLSIPS substrate to bind to the desired antigens attached to the substrate. When a sufficient density of antibody molecules are attached, they may be removed by an automated process, preferably as antibody populations exhibiting specificity of binding.

In one particular embodiment, a VLSIPS substrate, e.g., with a large plurality of fingerprint antigens attached thereto, is used to isolate antibodies from a supernatant of a population of cells producing antibodies to the antigens. Using the substrate as an affinity reagent, the antibodies will attach to the appropriate positionally defined antigens. The antibodies may be carefully removed therefrom, preferably by an automated system which retains their homogeneous specificities. The isolated antibodies can be attached to a new substrate in a positionally defined matrix pattern.

In a further embodiment, these spatially separated antibodies may be isolated using a specific targeting method for isolation. In this embodiment, a linker molecule which attaches to a particular portion of the antibody, preferably away from the binding site, can be attached to the antibodies. Various reagents will be used, including staphylococcus protein A or antibodies which bind to domains remote from the binding site. Alternatively, the antibodies in the population, before affinity purification, may be derivatized with an appropriate reagent compatible with new VLSIPS synthesis. A preferred reagent is a nucleotide which can serve as a linker to synthetic VLSIPS steps for synthesizing a specific sequence thereon. Then, by successive VLSIPS cycles, each of the antibodies attached to the defined antigen regions can have a defined oligonucleotide synthesized thereon and corresponding in area to the region of the substrate having each antigen attached. These defined oligonucleotides will be useful as targeting reagents to attach those antibodies possessing the same target sequence specificity at defined positions on a new substrate, by virtue of having bound to the antigen region, to a new VLSIPS substrate having the complementary target oligonucleotides positionally located on it. In this fashion,

a VLSIPS substrate having the desired antigens attached thereto can be used to generate a second VLSIPS substrate with positionally defined reagents which recognize those antigens.

5 The selected antigens will typically be selected to be those which define particular functionalities or properties, so as to be useful for fingerprinting and other uses. They will also be useful for mapping and sequencing embodiments.

IV. FINGERPRINTING

10 A. General

Many of the procedures and techniques used in the polynucleotide sequencing section are also appropriate for fingerprinting applications. See, e.g., Poustka, et al. (1986) Cold Spring Harbor Symposia on Quant. Biol., vol. LI, 131-139, 15 Cold Spring Harbor Press, New York; which is hereby incorporated herein by reference. The fingerprinting method provided herein is based, in part, upon the ability to positionally localize a large number of different specific probes onto a single substrate. This high density matrix 20 pattern provides the ability to screen for, or detect, a very large number of different sequences simultaneously. In fact, depending upon the hybridization conditions, fingerprinting to the resolution of virtually absolute matching of sequence is possible thereby approaching an absolute sequencing embodiment. 25 And the sequencing embodiment is very useful in identifying the probes useful in further fingerprinting uses. For example, characteristic features of genetic sequences will be identified as being diagnostic of the entire sequence. However, in most embodiments, longer probe and target will be used, and for 30 which slight mismatching may not need to be resolved.

B. Preparation of Substrate Matrix

A collection of specific probes may be produced by either of the methods described above in the section on 35 sequencing. Specific oligonucleotide probes of desired lengths may be individually synthesized on a standard oligonucleotide synthesizer. The length of these probes is limited only by the length of the ability of the synthesizer to continue to

accurately synthesize a molecule. Oligonucleotides or sequence fragments may also be isolated from natural sources.

Biological amplification methods may be coupled with synthetic synthesizing procedures such as, e.g., polymerase chain
5 reaction.

In one embodiment, the individually isolated probes may be attached to the matrix at defined positions. These probe reagents may be attached by an automated process making use of the caged biotin methodology described in U.S.S.N.

10 07/612,671 (caged biotin CIP), or using photochemical reagents, see, e.g., Dattagupta et al. (1985) U.S. Pat. No. 4,542,102 and (1987) U.S. Pat. No. 4,713,326. Each individual purified reagent can be attached individually at specific locations on a substrate.

15 In another embodiment, the VLSIPS synthesizing technique may be used to synthesize the desired probes at specific positions on a substrate. The probes may be synthesized by successively adding appropriate monomer subunits, e.g., nucleotides, to generate the desired sequences.

20 In another embodiment, a relatively short specific oligonucleotide is used which serves as a targeting reagent for positionally directing the sequence recognition reagent. For example, the sequence specific reagents having a separate additional sequence recognition segment (usually of a different
25 polymer from the target sequence) can be directed to target oligonucleotides attached to the substrate. By use of non-natural targeting reagents, e.g., unusual nucleotide analogues which pair with other unnatural nucleotide analogues and which do not interfere with natural nucleotide interactions, the
30 natural and non-natural portions can coexist on the same molecule without interfering with their individual functionalities. This can combine both a synthetic and biological production system analogous to the technique for targeting monoclonal antibodies to locations on a VLSIPS
35 substrate at defined positions. Unnatural optical isomers of nucleotides may be useful unnatural reagents subject to similar chemistry, but incapable of interfering with the natural biological polymers. See also, U.S.S.N. __/__, __, __, attorney

docket number 11509-26 (sequencing by synthesis); which is hereby incorporated herein by reference.

After the separate substrate attached reagents are attached to the targeting segment, the two are crosslinked, thereby permanently attaching them to the substrate. Suitable crosslinking reagents are known, see, e.g., Dattagupta et al. (1985) U.S. Pat., No. 4,542,102 and (1987) "Coupling of nucleic acids to solid support by photochemical methods," U.S. Pat. No. 4,713,326, each of which is hereby incorporated herein by reference. Similar linkages for attachment of proteins to a solid substrate are provided, e.g., in Merrifield (1986) Science 232:341-____, which is hereby incorporated herein by reference.

C. Labeling Target Nucleotides

The labeling procedures used in the sequencing embodiments will also be applicable in the fingerprinting embodiments. However, since the fingerprinting embodiments often will involve relatively large target molecules and relatively short oligonucleotide probes, the amount of signal necessary to incorporate into the target sequence may be less critical than in the sequencing applications. For example, a relatively long target with a relatively small number of labels per molecule may be easily amplified or detected because of the relatively large target molecule size.

In various embodiments, it may be desired to cleave the target into smaller segments as in the sequencing embodiments. The labeling procedures and cleavage techniques described in the sequencing embodiments would usually also be applicable here.

D. Hybridization Conditions

The hybridization conditions used in fingerprinting embodiments will typically be less critical than for the sequencing embodiments. The reason is that the amount of mismatching which may be useful in providing the fingerprinting information would typically be far greater than that necessary in sequencing uses. For example, Southern hybridizations do

not typically distinguish between slightly mismatched sequences. Under these circumstances, important and valuable information may be arrived at with less stringent hybridization conditions while providing valuable fingerprinting information.

5 However, since the entire substrate is typically exposed to the target molecule at one time, the binding affinity of the probes should usually be of approximately comparable levels. For this reason, if oligonucleotide probes are being used, their lengths should be approximately comparable and will be selected to
10 hybridize under conditions which are common for most of the probes on the substrate. Much as in a Southern hybridization, the target and oligonucleotide probes are of lengths typically greater than about 25 nucleotides. Under appropriate hybridization conditions, e.g., typically higher salt and lower
15 temperature, the probes will hybridize irrespective of imperfect complementarity. In fact, with probes of greater than, e.g., about fifty nucleotides, the difference in stability of different sized probes will be relatively minor.

Typically the fingerprinting is merely for probing
20 similarity or homology. Thus, the stringency of hybridization can usually be decreased to fairly low levels. See, e.g., Wetmur and Davidson (1968) "Kinetics of Renaturation of DNA," J. Mol. Biol., 31:349-370; and Kanehisa, M. (1984) Nuc. Acids Res., 12:203-213.

25

E. Detection; VLSIPS Scanning

Detection methods will be selected which are appropriate for the selected label. The scanning device need not necessarily be digitized or placed into a specific digital
30 database, though such would most likely be done. For example, the analysis in fingerprinting could be photographic. Where a standardized fingerprint substrate matrix is used, the pattern of hybridizations may be spatially unique and may be compared photographically. In this manner, each sample may have a
35 characteristic pattern of interactions and the likelihood of identical patterns will preferably be such low frequency that the fingerprint pattern indeed becomes a characteristic pattern virtually as unique as an individual's fingertip fingerprint.

With a standardized substrate, every individual could be, in theory, uniquely identifiable on the basis of the pattern of hybridizing to the substrate.

Of course, the VLSIPS scanning apparatus may also be
5 useful to generate a digitized version of the fingerprint pattern. In this way, the identification pattern can be provided in a linear string of digits. This sequence could also be used for a standardized identification system providing significant useful medical transferability of specific data.
10 In one embodiment, the probes used are selected to be of sufficiently high resolution to measure the antigens of the major histocompatibility complex, it might even be possible to provide transplantation matching data in a linear stream of data. The fingerprinting data may provide a condensed version,
15 or summary, of the linear genetic data, or any other information data base.

F. Analysis

The analysis of the fingerprint will often be much
20 simpler than a total sequence determination. However, there may be particular types of analysis which will be substantially simplified by a selected group of probes. For example, probes which exhibit particular populational heterogeneity may be selected. In this way, analysis may be simplified and
25 practical utility enhanced merely by careful selection of the specific probes and a careful matrix layout of those probes.

G. Substrate Reuse

As with the sequencing application, the
30 fingerprinting usages may also take advantage of the reusability of the substrate. In this way, the interactions can be disrupted, the substrate treated, and the renewed substrate is equivalent to an unused substrate.

35 H. Non-polynucleotide Aspects

Besides polynucleotide applications, the fingerprinting analysis may be applied to other polymers, especially polypeptides, carbohydrates, and other polymers,

both organic and inorganic. Besides using the fingerprinting method for analyzing a particular polymer, the fingerprinting method may be used to characterize various samples. For example, a cell or population of cells may be tested for their expression of specific antigens or their mRNA sequence intent. For example, a T-cell may be classified by virtue of its combination of expressed surface antigens. With specific reagents which interact with these antigens, a cell or a population of cells or a lysed cell may be exposed to a VLSIPS substrate. The biological sample may be classified or characterized by analyzing the pattern of specific interaction. This may be applicable to a cell or tissue type, to the expressed messenger RNA population expressed by a cell to the genetic content of a cell, or to virtually any sample which can be classified and/or identified by its combination of specific molecular properties.

The ability to generate a high density means for screening the presence or absence of specific interactions allows for the possibility of screening for, if not saturating, all of a very large number of possible interactions. This is very powerful in providing the means for testing the combinations of molecular properties which can define a class of samples. For example, a species of organism may be characterized by its DNA sequences, e.g., a genetic fingerprint. By using a fingerprinting method, it may be determined that all members of that species are sufficiently similar in specific sequences that they can be easily identified as being within a particular group. Thus, newly defined classes may be resolved by their similarity in fingerprint patterns. Alternatively, a non-member of that group will fail to share those many identifying characteristics. However, since the technology allows testing of a very large number of specific interactions, it also provides the ability to more finely distinguish between closely related different cells or samples. This will have important applications in diagnosing viral, bacterial, and other pathological on nonpathological infections.

In particular, cell classification may be defined by any of a number of different properties. For example, a cell class may be defined by its DNA sequences contained therein. This allows species identification for parasitic or other
5 infections. For example, the human cell is presumably genetically distinguishable from a monkey cell, but different human cells will share many genetic markers. At higher resolution, each individual human genome will exhibit unique sequences that can define it as a single individual.

10 Likewise, a developmental stage of a cell type may be definable by its pattern of expression of messenger RNA. For example, in particular stages of cells, high levels of ribosomal RNA are found whereas relatively low levels of other
15 types of messenger RNAs may be found. The high resolution distinguishability provided by this fingerprinting method allows the distinction between cells which have relatively minor differences in its expressed mRNA population. Where a pattern is shown to be characteristic of a stage, a stage may be defined by that particular pattern of messenger RNA
20 expression.

In a similar manner, the antigenic determinants found on a protein may very well define the cell class. For example, immunological T-cells are distinguishable from B-cells because, in part, the cell surface antigens on the cell types are
25 distinguishable. Different T-cell subclasses can be also distinguished from one another by whether they contain particular T-cell antigens. The present invention provides the possibility for high resolution testing of many different interactions simultaneously, and the definition of new cell
30 types will be possible.

The high resolution VLSIPS substrate may also be used as a very powerful diagnostic tool to test the combination of presence, of a plurality of different assays from a biological sample. For example, a cancerous condition may be indicated by
35 a combination of various different properties found in the blood. For example, a cancerous condition may be indicated by a combination of expression of various soluble antigens found in the blood along with a high number of various cellular

antigens found on lymphocytes and/or particular cell degradation products. With a substrate as provided herein, a large number of different features can be simultaneously performed on a biological sample. In fact, the high resolution
5 of the test will allow more complete characterization of parameters which define particular diseases. Thus, the power of diagnostic tests may be limited by the extent of statistical correlation with a particular condition rather than with the number of antigens or interactions which are tested. The
10 present invention provides the means to generate this large universe of possible reagents and the ability to actually accumulate that correlative data.

In another embodiment, a substrate as provided herein may be used for genetic screening. This would allow for
15 simultaneous screening of thousands of genetic markers. As the density of the matrix is increased, many more molecules can be simultaneously tested. Genetic screening then becomes a simpler method as the present invention provides the ability to screen for thousands, tens of thousands, and hundreds of
20 thousands, even millions of different possible genetic features. However, the number of high correlation genetic markers for conditions numbers only in the hundreds. Again, the possibility for screening a large number of sequences provides the opportunity for generating the data which can
25 provide correlation between sequences and specific conditions or susceptibility. The present invention provides the means to generate extremely valuable correlations useful for the genetic detection of the causative mutation leading to medical conditions. In still another embodiment, the present invention
30 would be applicable to distinguishing two individuals having identical genetic compositions. The antibody population within an individual is dependent both on genetic and historical factors. Each individual experiences a unique exposure to various infectious agents, and the combined antibody expression
35 is partly determined thereby. Thus, individuals may also be fingerprinted by their immunological content, either of actively expressed antibodies, or their immunological memory. Similar sorts of immunological and environmental histories may

be useful for fingerprinting, perhaps in combination with other screening properties. In particular, the present invention may be useful for screening allergic reactions or susceptibilities, a simple IgE specificity test may be useful in determining a spectrum of allergies.

With the definition of new classes of cells, a cell sorter will be used to purify them. Moreover, new markers for defining that class of cells will be identified. For example, where the class is defined by its RNA content, cells may be screened by antisense probes which detect the presence or absence of specific sequences therein. Alternatively, cell lysates may provide information useful in correlating intracellular properties with extracellular markers which indicate functional differences. Using standard cell sorter technology with a fluorescence or labeled antisense probe which recognizes the internal presence of the specific sequences of interest, the cell sorter will be able to isolate a relatively homogeneous population of cells possessing the particular marker. Using successive probes the sorting process should be able to select for cells having a combination of a large number of different markers.

In a non-polynucleotide embodiment, cells may be defined by the presence of other markers. The markers may be carbohydrates, proteins, or other molecules. Thus, a substrate having particular specific reagents, e.g., antibodies, attached to it should be able to identify cells having particular patterns of marker expression. Of course, combinations of these made be utilized and a cell class may be defined by a combination of its expressed mRNA, its carbohydrate expression, its antigens, and other properties. This fingerprinting should be useful in determining the physiological state of a cell or population of cells.

Having defined a cell type whose function or properties are defined by the reagents attachable to a VLSIPS substrate, such as cellular antigens, these structural manifestations of function may be used to sort cells to generate a relatively homogeneous population of that class of cells. Standard cell sorter technology may be applied to

purify such a population, see, e.g., Dengl, J. and Herzenberg (1982) "Selection of hybridomas and hybridoma variants using the fluorescence activated cell sorter," J. Immunological Methods 52:1-14; and Becton Dickinson, Fluorescence Activated Cell Sorter Division, San Jose, California, and Coulter Diagnostics, Hialeah, Florida.

With the fingerprinted method as in identification means arises from mosaicism problems in an organism. A mosaic organism is one whose genetic content in different cells is significantly different. Various clonal populations should have similar genetic fingerprints, though different clonal populations may have different genetic contents. See, for example, Suzuki et al. An Introduction to Genetic Analysis (4th Ed.), Freeman and Co., New York, which is hereby incorporated herein by reference. However, this problem should be a relatively rare problem and could be more carefully evaluated with greater experience using the fingerprinting methods.

The invention will also find use in detecting changes, both genetic and antigenic, e.g., in a rapidly "evolving" protozoa infection, or similarly changing organism.

V. MAPPING

A. General

The use of the present invention for mapping parallels its use for fingerprinting and sequencing. Where a polymer is a linear molecule, the mapping provides the ability to locate particular segments along the length of the polymer. Branched polymers can be treated as a series of individual linear polymers. The mapping provides the ability to locate, in a relative sense, the order of various subsequences. This may be achieved using at least two different approaches.

The first approach is to take the large sequence and fragment it at specific points. The fragments are then ordered and attached to a solid substrate. For example, the clones resulting from a chromosome walking process may be individually attached to the substrate by methods, e.g., caged biotin techniques, indicated earlier. Segments of unknown map position will be exposed to the substrate and will hybridize to

the segment which contains that particular sequence. This procedure allows the rapid determination of a number of different labeled segments, each mapping requiring only a single hybridization step once the substrate is generated. The substrate may be regenerated by removal of the interaction, and the next mapping segment applied.

In an alternative method, a plurality of subsequences can be attached to a substrate. Various short probes may be applied to determine which segments may contain particular overlaps. The theoretical basis and a description of this mapping procedure is contained in, e.g., Evans et al. 1989 "Physical Mapping of Complex Genomes by Cosmid Multiplex Analysis," Proc. Natl. Acad. Sci. USA 86:5030-5034, and other references cited above in the Section labeled "Overall Description." Using this approach, the details of the mapping embodiment are very similar to those used in the fingerprinting embodiment.

B. Preparation of Substrate Matrix

The substrate may be generated in either of the methods generally applicable in the sequencing and fingerprinting embodiments. The substrate may be made either synthetically, or by attaching otherwise purified probes or sequences to the matrix. The probes or sequences may be derived either from synthetic or biological means. As indicated above, the solid phase substrate synthetic methods may be utilized to generate a matrix with positionally defined sequences. In the mapping embodiment, the importance of saturation of all possible subsequences of a preselected length is far less important than in the sequencing embodiment, but the length of the probes used may be desired to be much longer. The processes for making a substrate which has longer oligonucleotide probes should not be significantly different from those described for the sequencing embodiments, but the optimization parameters may be modified to comply with the mapping needs.

C. Labeling

The labeling methods will be similar to those applicable in sequencing and fingerprinting embodiments. Again, the target sequences may be desired to be fragmented.

5

D. Hybridization/Specific Interaction

The specificity of interaction between the targets and probe would typically be closer to those used for fingerprinting embodiments, where homology is more important than absolute distinguishability of high fidelity complementary hybridization. Usually, the hybridization conditions will be such that merely homologous segments will interact and provide a positive signal. Much like the fingerprinting embodiment, it may be useful to measure the extent of homology by successive incubations at higher stringency conditions. Or, a plurality of different probes, each having various levels of homology may be used. In either way, the spectrum of homologies can be measured.

Where non-nucleic acid hybridization is involved, the specific interactions may also be compared in a fingerprint-like manner. The specific reagents may have less specificity, e.g., monoclonal antibodies which recognize a broader spectrum of sequences may be utilized relative to a sequencing embodiment. Again, the specificity of interaction may be measured under various conditions of increasing stringency to determine the spectrum of matching across the specific probes selected, or a number of different stringency reagents may be included to indicate the binding affinity.

E. Detection

The detection methods used in the mapping procedure will be virtually identical to those used in the fingerprinting embodiment. The detection methods will be selected in combination with the labeling methods.

35

F. Analysis

The analysis of the data in a mapping embodiment will typically be somewhat different from that in fingerprinting.

The fingerprinting embodiment will test for the presence or absence of specific or homologous segments. However, in the mapping embodiment, the existence of an interaction is coupled with some indication of the location of the interaction. The interaction is mapped in some manner to the physical polymer sequence. Some means for determining the relative positions of different probes, is performed. This may be achieved by synthesis of the substrate in pattern, or may result from analysis of sequences after they have been attached to the substrate.

-For example, the probes may be randomly positioned at various locations on the substrate. However, the relative positions of the various reagents in the original polymer may be determined by using short fragments, e.g., individually, as target molecules which determine the proximity of different probes. By an automated system of testing each different short fragment of the original polymer, coupled with proper analysis, it will be possible to determine which probes are adjacent one another on the original target sequence and correlate that with positions on the matrix. In this way, the matrix is useful for determining the relative locations of various new segments in the original target molecule. This sort of analysis is described in Evans, and the related references described above.

G. Substrate Reuse

The substrate should be reusable in the manner described in the fingerprinting section. The substrate is renewed by removal of the specific interactions and is washed and prepared for successive cycles of exposure to new target sequences.

H. Non-polynucleotide Aspects

The mapping procedure may be used on other molecules than polynucleotides. Although hybridization is one type of specific interaction which is clearly useful for use in this mapping embodiment, antibody reagents may also be very useful. In the same way that polypeptide sequencing or other polymers may be sequenced by the reagents and techniques described in

the sequencing section and fingerprinting section, the mapping embodiment may also be used similarly.

In another form of mapping, as described above in the fingerprinting section, the developmental map of a cell or biological system may be measured using fingerprinting type technology. Thus, the mapping may be along a temporal dimension rather than along a polymer dimension. The mapping or fingerprinting embodiments may also be used in determining the genetic rearrangements which may be genetically important, as in lymphocyte and B-cell development. In another example, various rearrangements or chromosomal dislocations may be tested by either the fingerprinting or mapping methods. These techniques are similar in many respects and the fingerprinting and mapping embodiments may overlap in many respects.

VI. ADDITIONAL SCREENING AND APPLICATIONS

A. Specific Interactions

As originally indicated in the parent filing of VLSIPS, the production of a high density plurality of spatially segregated polymers provides the ability to generate a very large universe or repertoire of individually and distinct sequence possibilities. As indicated above, particular oligonucleotides may be synthesized in automated fashion at specific locations on a matrix. In fact, these oligonucleotides may be used to direct other molecules to specific locations by linking specific oligonucleotides to other reagents which are in batch exposed to the matrix and hybridized in a complementary fashion to only those locations where the complementary oligonucleotide has been synthesized on the matrix. This allows for spatially attaching a plurality of different reagents onto the matrix instead of individually attaching each separate reagent at each specific location. Although the caged biotin method allows the automated attachment, the speed of the caged biotin attachment process is relatively slow and requires a separate reaction for each reagent being attached. By use of the oligonucleotide method, the specificity of position can be done in an automated and parallel fashion. As each reagent is produced, instead of

directly attaching each reagent at each desired position, the reagent may be attached to a specific desired complementary oligonucleotide which will ultimately be specifically directed toward locations on the matrix having a complementary
5 oligonucleotide attached thereat.

In addition, the technology allows screening for specificity of interaction with particular reagents. For example, the oligonucleotide sequence specificity of binding of a potential reagent may be tested by presenting to the reagent
10 all of the possible subsequences available for binding. Although secondary or higher order sequence specific features might not be easily screenable using this technology, it does provide a convenient, simple, quick, and thorough screen of interactions between a reagent and its target recognition
15 sequences. See, e.g., Pfeifer et al. (1989) Science 246:810-812.

For example, the interaction of a promoter protein with its target binding sequence may be tested for many different, or all, possible binding sequences. By testing the
20 strength of interactions under various different conditions, the interaction of the promoter protein with each of the different potential binding sites may be analyzed. The spectrum of strength of interactions with each different potential binding site may provide significant insight into the
25 types of features which are important in determining specificity.

An additional example of a sequence specific interaction between reagents is the testing of binding of a double stranded nucleic acid structure with a single stranded
30 oligonucleotide. Often, a triple stranded structure is produced which has significant aspects of sequence specificity. Testing of such interactions with either sequences comprising only natural nucleotides, or perhaps the testing of nucleotide analogs may be very important in screening for particularly
35 useful diagnostic or therapeutic reagents. See, e.g., Häner and Dervan (1990) Biochemistry 29:9761-6765, and references therein.

B. Sequence Comparisons

Once a gene is sequenced, the present invention provides means to compare alleles or related sequences to locate and identify differences from the control sequence.

- 5 This would be extremely useful in further analysis of genetic variability at a specific gene locus.

C. Categorizations

- 10 As indicated above in the fingerprinting and mapping embodiments, the present invention is also useful to define specific stages in the temporal sequence of cells, e.g., development, and the resulting tissues within an organism. For example, the developmental stage of a cell, or population of cells, can be dependent upon the expression of particular
15 messenger RNAs or cellular antigens. The screening procedures provided allow for high resolution definition of new classes of cells. In addition, the temporal development of particular cells will be characterized by the presence or expression of various mRNAs. Means to simultaneously screen a plurality or
20 very large number of different sequences as provided. The combination of different markers made available dramatically increases the ability to distinguish fairly closely related cell types. Other markers may be combined with markers and methods made available herein to define new classifications of
25 biological samples, e.g., based upon new combinations of markers.

- The presence or absence of particular marker sequences will be used to define temporal developmental stages. Once the stages are defined, fairly simple methods can be
30 applied to actually purify those particular cells. For example, antisense probes or recognition reagents may be used with a cell sorter to select those cells containing or expressing the critical markers. Alternatively, the expression of those sequences may result in specific antigens which may
35 also be used in defining cell classes and sorting those cells away from others. In this way, for example, it should be possible to select a class of omnipotent immune system cells which are able to completely regenerate a human immune system.

Based upon the cellular classes defined by the parameters made available by this technology, purified classes of cells having identifiable differences, structural or functional, are made available.

5 In an alternative embodiment, a plurality of antigens or specific binding proteins attached to the substrate may be used to define particular cell types. For example, subclasses of T-cells are defined, in part, upon the combination of expressed cell surface antigens. The present invention allows
10 for the simultaneous screening of a large plurality of different antigens together. Thus, higher resolution classification of different T-cell subclasses becomes possible and, with the definitions and functional differences which correlate with those antigenic or other parameters, the ability
15 to purify those cell types becomes available. This is applicable not only to T-cells, lymphocyte cells, or even to freely circulating cells. Many of the cells for which this would be most useful will be immobile cells found in particular tissues or organs. Tumor cells will be diagnosed or detected
20 using these fingerprinting techniques. Coupled with a temporal change in structure, developmental classes may also be selected and defined using these technologies. The present invention also provides the ability not only to define new classes of cells based upon functional or structural differences, but it
25 also provides the ability to select or purify populations of cells which share these particular properties. Standard cell sorting procedures using antibody markers may be used to detect extracellular features. Intracellular features would also be amendable by introducing the label reagents into the cell. In
30 particular, antisense DNA or RNA molecules may be introduced into a cell to detect RNA sequences therein. See, e.g., Weintraub (1990) Scientific American 262:40-46.

D. Statistical Correlations

35 In an additional embodiment, the present invention also allows for the high resolution correlation of medical conditions with various different markers. For example, the present technology, when applied to amniocentesis or other

genetic screening methods, typically screen for tens of different markers at most. The present invention allows simultaneous screening for tens, hundreds, thousands, tens of thousands, hundreds of thousands, and even millions of different genetic sequences. Thus, applying the fingerprinting methods of the present invention to a sufficiently large population allows detailed statistical analysis to be made, thereby correlating particular medical conditions with particular markers, typically antigenic or genetic. Tumor specific antigens will be identified using the present invention.

Various medical conditions may be correlated against an enormous data base of the sequences within an individual. Genetic propensities and correlations then become available and high resolution genetic predictability and correlation become much more easily performed. With the enormous data base, the reliability of the predictions also is better tested. Particular markers which are partially diagnostic of particular medical conditions or medical susceptibilities will be identified and provide direction in further studies and more careful analysis of the markers involved. Of course, as indicated above in the sequencing embodiment, the present invention will find much use in intense sequencing projects. For example, sequencing of the entire human genome in the human genome project will be greatly simplified and enabled by the present invention.

VI. FORMATION OF SUBSTRATE

The substrate is provided with a pattern of specific reagents which are positionally localized on the surface of the substrate. This matrix of positions is defined by the automated system which produces the substrate. The instrument will typically be one similar to that described in U.S.S.N. 07/492,462 (VLSIPS CIP), and U.S.S.N. __/__, __, __, attorney docket number 11509-28 (automated VLSIPS). The instrumentation described therein is directly applicable to the applications used here. In particular, the apparatus comprises a substrate, typically a silicon containing substrate, on which positions on

the surface may be defined by a coordinate system of positions. These positions can be individually addressed or detected by the VLSIPS apparatus.

Typically, the VLSIPS apparatus uses optical methods used in semiconductor fabrication applications. In this way, masks may be used to photo-activate positions for attachment or synthesis of specific sequences on the substrate. These manipulations may be automated by the types of apparatus described in U.S.S.N. 07/462,492 (VLSIPS CIP) and U.S.S.N.

10 , attorney docket number 11509-28 (automated VLSIPS).

Selectively removable protecting groups allow creation of well defined areas of substrate surface having differing reactivities. Preferably, the protecting groups are selectively removed from the surface by applying a specific activator, such as electromagnetic radiation of a specific wavelength and intensity. More preferably, the specific activator exposes selected areas of surface to remove the protecting groups in the exposed areas.

Protecting groups of the present invention are used in conjunction with solid phase oligomer syntheses, such as peptide syntheses using natural or unnatural amino acids, nucleotide syntheses using deoxyribonucleic and ribonucleic acids, oligosaccharide syntheses, and the like. In addition to protecting the substrate surface from unwanted reaction, the protecting groups block a reactive end of the monomer to prevent self-polymerization. For instance, attachment of a protecting group to the amino terminus of an activated amino acid, such as the N-hydroxysuccinimide-activated ester of the amino acid prevents the amino terminus of one monomer from reacting with the activated ester portion of another during peptide synthesis.

Alternatively, the protecting group may be attached to the carboxyl group of an amino acid to prevent reaction at this site. Most protecting groups can be attached to either the amino or the carboxyl group of an amino acid, and the nature of the chemical synthesis will dictate which reactive group will require a protecting group. Analogously, attachment of a protecting group to the 5'-hydroxyl group of a nucleoside

during synthesis using for example, phosphate-triester coupling chemistry, prevents the 5'-hydroxyl of one nucleoside from reacting with the 3'-activated phosphate-triester of another.

Regardless of the specific use, protecting groups are employed to protect a moiety on a molecule from reacting with another reagent. Protecting groups of the present invention have the following characteristics: they prevent selected reagents from modifying the group to which they are attached; they are stable (that is, they remain attached) to the synthesis reaction conditions; they are removable under conditions that do not adversely affect the remaining structure; and once removed, do not react appreciably with the surface or surface-bound oligomer. The selection of a suitable protecting group will depend, of course, on the chemical nature of the monomer unit and oligomer, as well as the specific reagents they are to protect against.

In a preferred embodiment, the protecting groups will be photoactivatable. The properties and uses of photoreactive protecting compounds have been reviewed. See, McCray et al., Ann. Rev. of Biophys. and Biophys. Chem. (1989) 18:239-270, which is incorporated herein by reference. Preferably, the photosensitive protecting groups will be removable by radiation in the ultraviolet (UV) or visible portion of the electromagnetic spectrum. More preferably, the protecting groups will be removable by radiation in the near UV or visible portion of the spectrum. In some embodiments, however, activation may be performed by other methods such as localized heating, electron beam lithography, laser pumping, oxidation or reduction with microelectrodes, and the like. Sulfonyl compounds are suitable reactive groups for electron beam lithography. Oxidative or reductive removal is accomplished by exposure of the protecting group to an electric current source, preferably using microelectrodes directed to the predefined regions of the surface which are desired for activation. A more detailed description of these protective groups is provided in U.S.S.N. / , , attorney docket number 11509-28 (automated VLSIPS), which is hereby incorporated herein by reference.

The density of reagents attached to a silicon substrate may be varied by standard procedures. The surface area for attachment of reagents may be increased by modifying the silicon surface. For example, a matte surface may be machined or etched on the substrate to provide more sites for attachment of the particular reagents. Another way to increase the density of reagent binding sites is to increase the derivitization density of the silicon. Standard procedures for achieving this are described, below.

One method to control the derivatization density is to highly-derivatize the substrate with photochemical groups at high density. The substrate is then photolyzed for various predetermined times, which photoactivate the groups at a measurable rate, and react then with a capping reagent. By this method, the density of linker groups may be modulated by using a desired time and intensity of photoactivation.

In many applications, the number of different sequences which may be provided may be limited by the density and the size of the substrate on which the matrix pattern is generated. In situations where the density is insufficiently high to allow the screening of the desired number of sequences, multiple substrates may be used to increase the number of sequences tested. Thus, the number of sequences tested may be increased by using a plurality of different substrates.

Because the VLSIPS apparatus is almost fully automated, increasing the number of substrates does not lead to a significant increase in the number of manipulations which must be performed by humans. This again leads to greater reproducibility and speed in the handling of these multiple substrates.

A. Instrumentation

The concept of using VLSIPS generally allows a pattern or a matrix of reagents to be generated. The procedure for making the pattern is performed by any of a number of different methods. An apparatus and instrumentation useful for generating a high density VLSIPS substrate is described in

detail in U.S.S.N. 07/492,462 (VLSIPS CIP) and U.S.S.N.
____/____,____, attorney docket number 11509-28 (automated VLSIPS).

B. Binary Masking

5 The details of the binary masking are described in an accompanying application filed simultaneously with this, U.S.S.N. ____/____,____, attorney docket number 11509-28 (automated VLSIPS) whose specification is incorporated herein by reference.

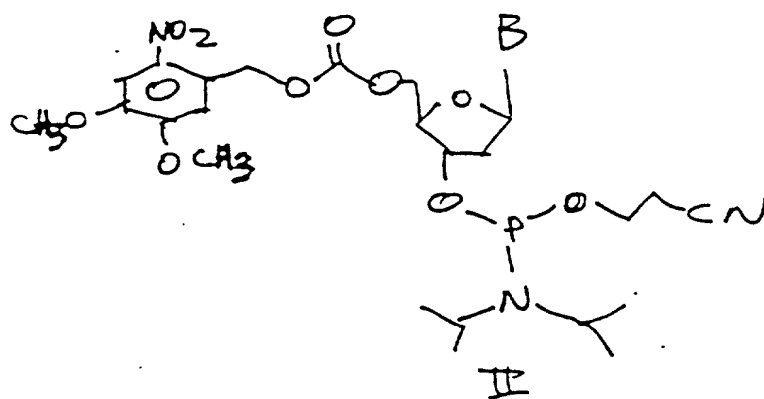
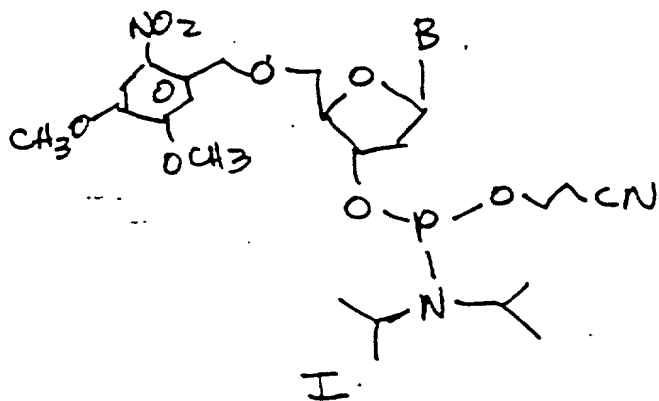
10 For example, the binary masking technique allows for producing a plurality of sequences based on the selection of either of two possibilities at any particular location. By a series of binary masking steps, the binary decision may be the determination, on a particular synthetic cycle, whether or not
15 to add any particular one of the possible subunits. By treating various regions of the matrix pattern in parallel, the binary masking strategy provides the ability to carry out spatially addressable parallel synthesis.

C. Synthetic Methods

20 The synthetic methods in making a substrate are described in the parent application, U.S.S.N. 07/492,462. The construction of the matrix pattern on the substrate will typically be generated by the use of photo-sensitive reagents.

25 By use of photo-lithographic optical methods, particular segments of the substrate can be irradiated with light to activate or deactivate blocking agents, e.g., to protect or deprotect particular chemical groups. By an appropriate sequence of photo-exposure steps at appropriate times with
30 appropriate masks and with appropriate reagents, the substrates can have known polymers synthesized at positionally defined regions on the substrate. Methods for synthesizing various substrates are described in U.S.S.N. 07/492,462 (VLSIPS CIP) and U.S.S.N. ____/____,____, attorney docket number 11509-28
35 (automated VLSIPS). By a sequential series of these photo-exposure and reaction manipulations, a defined matrix pattern of known sequences may be generated, and is typically referred to as a VLSIPS substrate. In the nucleic acid synthesis

embodiment, nucleosides used in the synthesis of DNA by photolytic methods will typically be one of the two forms shown below:



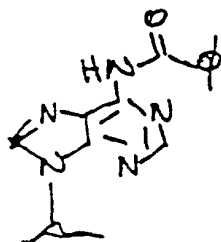
35

B = Adenine, Cytosine, Guanine, or Thymine

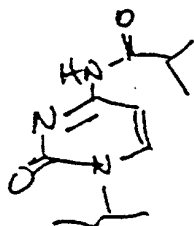
In I, the photolabile group at the 5' position is abbreviated NV (nitroveratryl) and in II, the group is

abbreviated NVOC (nitroveratryl oxycarbonyl). Although not shown in Fig. C the bases (adenine, cytosine, and guanine) contain exocyclic NH_2 groups which must be protected during DNA synthesis. Thymine contains no exocyclic NH_2 and therefore requires no protection. The standard protecting groups for these anaines are shown below:

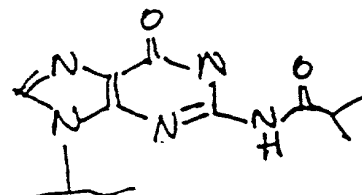
10



15



Cytosine (C)



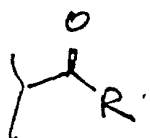
Guanine (G)

20

Adenine (A)

Other amides of the general formula

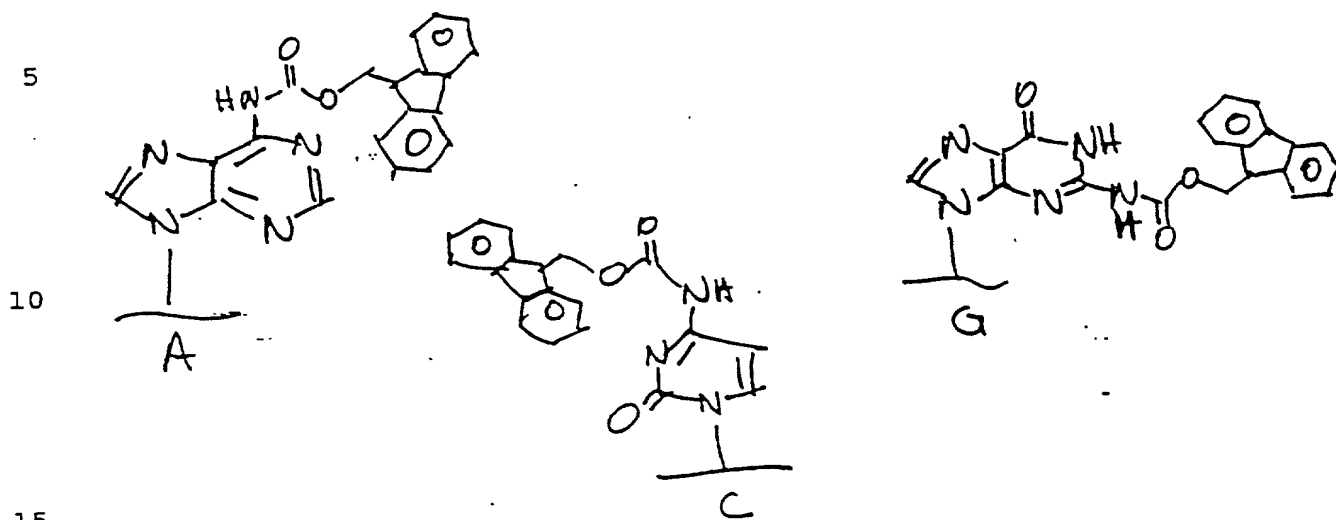
25.



R = alkyl
aryl

where R may be alkyl or aryl have been used.

Another type of protecting group FMOC (9-fluorenyl methoxycarbonyl) is currently being used to protect the exocyclic amines of the three bases:



Adenine (A)

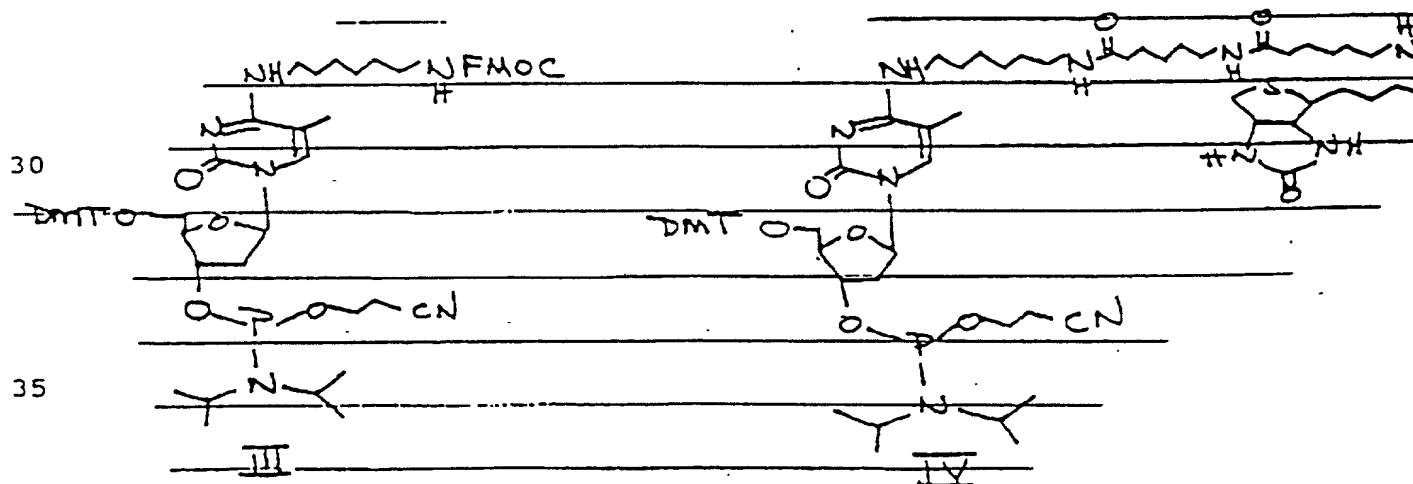
Cytosine (C)

Guanine (G)

20 The advantage of the Fmoc group is that it is removed under mild conditions (dilute organic bases) and can be used for all three bases. The amide protecting groups require more harsh conditions to be removed (NH_3/MeOH with heat).

Nucleosides used as 5'-OH probes, useful in verifying correct VLSIPS synthetic function, have been the following:

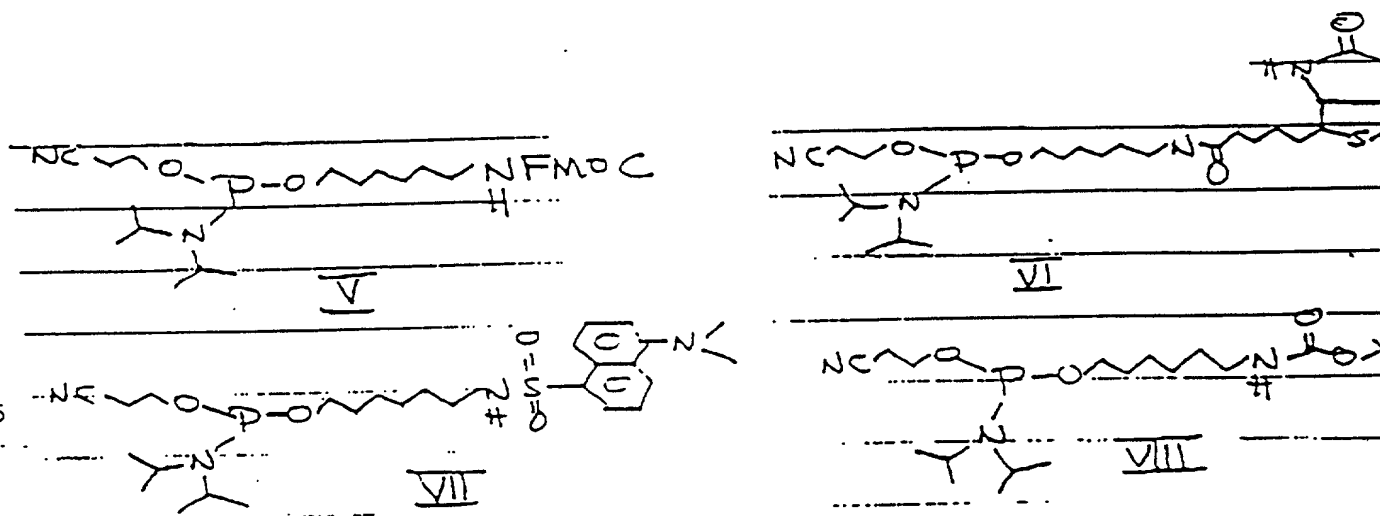
25



These compounds are used to detect where on a substrate photolysis has occurred by the attachment of either III or V to the newly generated 5'-OH. In the case of III, after the phosphate attachment is made, the substrate is

5 treated with a dilute base to remove the FMOC group. The resulting amine can be reacted with FITC and the substrate examined by fluorescence microscopy. This indicates the proper generation of a 5'-OH. In the case of compound IV, after the phosphate attachment is made, the substrate is treated with

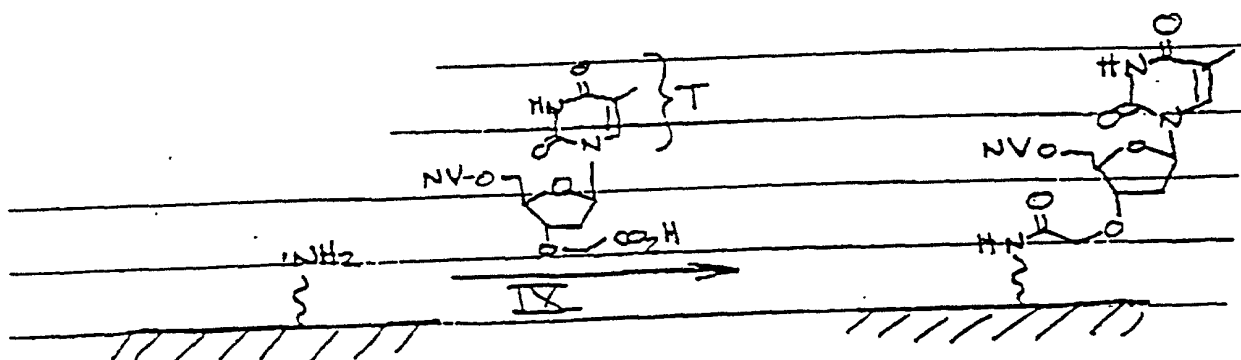
10 FITC labeled streptavidin and the substrate again may be examined by fluorescence microscopy. Other probes, although not nucleoside based, have included the following:



The method of attachment of the first nucleoside to the surface of the substrate depends on the functionality of the groups at the substrate surface. If the surface is amine functionalized, an amide bond is made (see example below).

5

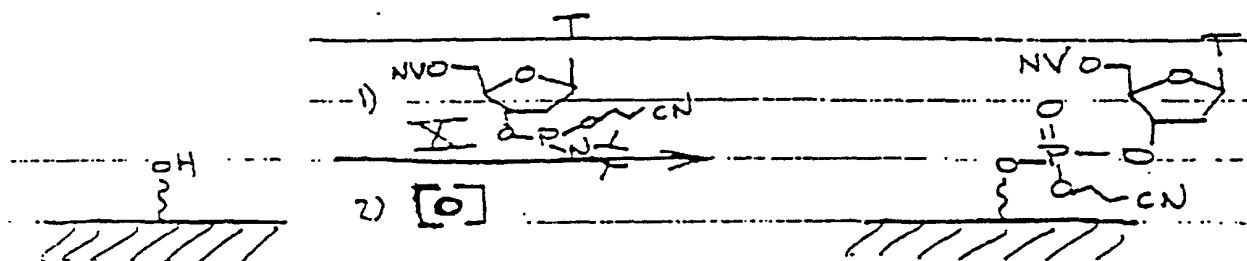
10



If the surface is hydroxy functionalized a phosphate bond is made (see example below)

15

20



25

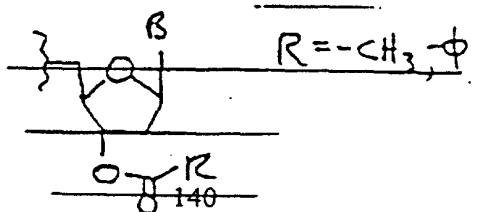
In both cases, the thymidine example is illustrated, but any one of the four phosphoramidite activated nucleosides can be used in the first step.

30

Photolysis of the photolabile group NV or NVOC on the 5' positions of the nucleosides is carried out at ~ 362 nm with an intensity of 14 mW/cm^2 for 10 minutes with the substrate side (side containing the photolabile group) immersed in dioxane. After the coupling of the next nucleoside is complete, the photolysis is repeated followed by another coupling until the desired oligomer is obtained.

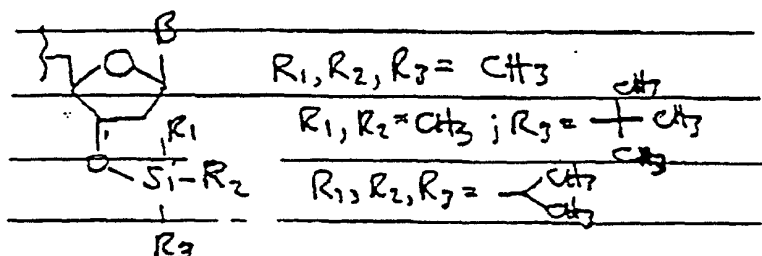
One of the most common 3'-O-protecting group is the ester, in particular the acetate

35



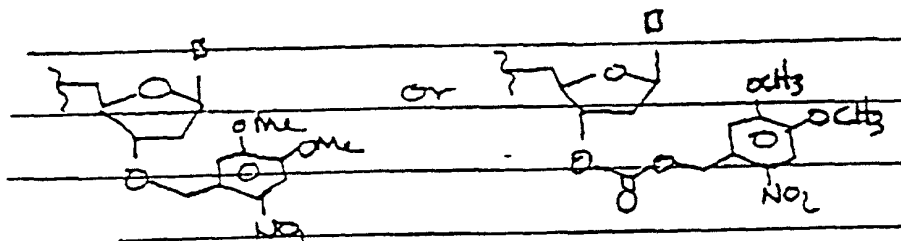
The groups can be removed by mild base treatment 0.1N NaOH/MeOH or $K_2CO_3/H_2O/MeOH$.

Another group used most often is the silyl ether.



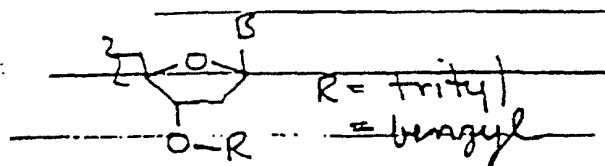
These groups can be removed by neutral conditions using 1 M tetra-n-butylammonium fluoride in THF or under acid conditions.

Related to photodeprotection, the nitroveratryl group could also be used to protect the 3'-position.



Here, light (photolysis) would be used to remove these protecting groups.

A variety of ethers can also be used in the protection of the 3'-O-position.



Removal of these groups usually involves acid or catalytic methods.

Note that corresponding linkages and photoblocked amino acids are described in detail in U.S.S.N. ____/____,____, attorney docket number 11509-28, which is hereby incorporated herein by reference.

Although the specificity of interactions at particular locations will usually be homogeneous due to a homogeneous polymer being synthesized at each defined location, for certain purposes, it may be useful to have mixed polymers with a commensurate mixed collection of interactions occurring at specific defined locations, or degeneracy reducing analogues, which have been discussed above and show broad specificity in binding. Then, a positive interaction signal may result from any of a number of sequences contained therein.

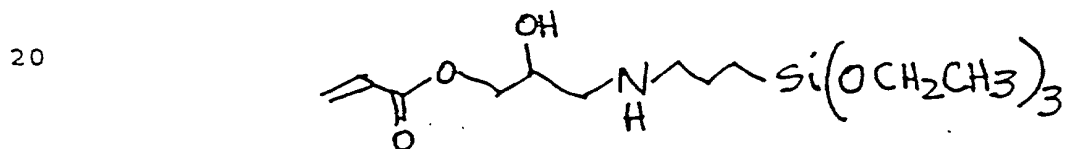
As an alternative method of generating a matrix pattern on a substrate, preformed polymers may be individually attached at particular sites on the substrate. This may be performed by individually attaching reagents one at a time to specific positions on the matrix, a process which may be automated. See, e.g., U.S.S.N. 07/435,316 (caged biotin parent), and U.S.S.N. 07/612,671 (caged biotin CIP). Another way of generating a positionally defined matrix pattern on a substrate is to have individually specific reagents which interact with each specific position on the substrate. For example, oligonucleotides may be synthesized at defined locations on the substrate. Then the substrate would have on its surface a plurality of regions having homogeneous oligonucleotides attached at each position.

In particular, at least four different substrate preparation procedures are available for treating a substrate surface. They are the standard VLSIPS method, polymeric substrates, DuraporeTM, and synthetic beads or fibers. The treatment labeled "standard VLSIPS" method is described in U.S.S.N. ____/____,____, attorney docket number 11509-28 (automated VLSIPS), and involves applying amino-propyltriethoxysilane to a glass surface.

The polymeric substrate approach involves either of two ways of generating a polymeric substrate. The first uses a high concentration of aminopropyltriethoxysilane (2-20%) in an aqueous ethanol solution (95%). This allows the silane

5 compound to polymerize both in solution and on the substrate surface, which provides a high density of amines on the surface of the glass. This density is contrasted with the standard VLSIPS method. This polymeric method allows for the deposition on the substrate surface of a monolayer due to the anhydrous
10 method used with the aforementioned silane.

The second polymeric method involves either the coating or covalent binding of an appropriate acrylic acid polymer onto the substrate surface.. In particular, e.g., in DNA synthesis, a monomer such as a hydroxypropylacrylate is
15 used to generate a high density of hydroxyl groups on the substrate surface, allowing for the formation of phosphate bonds. An example of such a compound is shown:



25 The method using a DuraporeTM membrane (Millipore) consists of a polyvinylidene difluoride coating with crosslinked polyhydroxylpropyl acrylate [PVDF-HPA]:



35 Here the building up of, e.g., a DNA oligomer, can be started immediately since phosphate bonds to the surface can be accomplished in the first step with no need for modification.

A nucleotide dimer (5'-C-T-3') has been successfully made on this substrate in our labs.

The fourth method utilizes synthetic beads or fibers.

This would use another substrate, such as a teflon copolymer graft bead or fiber, which is covalently coated with an organic layer (hydrophilic) terminating in hydroxyl sites (commercially available from Molecular Brosystems, Inc.) . This would offer the same advantage as the DuraporeTM membrane, allowing for immediate phosphate linkages, but would give additional contour by the 3-dimensional growth of oligomers.

A matrix pattern of new reagents may be targeted to each specific oligonucleotide position by attaching a complementary oligonucleotide to which the substrate bound form is complementary. For instance, a number of regions may have homogeneous oligonucleotides synthesized at various locations. Oligonucleotide sequences complementary to each of these can be individually generated and linked to a particular specific reagents. Often these specific reagents will be antibodies. As each of these is specific for finding its complementary oligonucleotide, each of the specific reagents will bind through the oligonucleotide to the appropriate matrix position. A single step having a combination of different specific reagents being attached specifically to a particular oligonucleotide will thereby bind to its complement at the defined matrix position. The oligonucleotides will typically then be covalently attached, using, e.g., an acridine dye, for photocrosslinking. Psoralen is a commonly used acridine dye for photocrosslinking purposes, see, e.g., Song et al. (1979) Photochem. Photobiol. 29:1177-1197; Cimino et al. (1985) Ann. Rev. Biochem. 54:1151-1193; Parsons (1980) Photochem. Photobiol. 32:813-821; and Dattagupta et al. (1985) U.S. Pat. No. 4,542,102, and (1987) U.S. Pat. No. 4,713,326; each of which is hereby incorporated herein by reference. This method allows a single attachment manipulation to attach all of the specific reagents to the matrix at defined positions and results in the specific reagents being homogeneously located at defined positions. In many embodiments, the specific reagents will be antibodies.

interaction. Because the avidin-biotin binding is very tight, this will usually be virtually irreversible binding.

2. crosslinked interactions

5 The surface immobilization may also take place by photo crosslinking of defined oligonucleotides linked to specific reagents. After hybridization of the complementary oligonucleotides, the oligonucleotides may be crosslinked by a reagent by psoralen or another similar type of acridine dye.
10 Other useful cross linking reagents are described in Dattagupta et al. (1985) U.S. Pat. No. 4,542,102, and (1987) U.S. Pat. No. 4,713,326.

In another embodiment, colony or phage plaque transfer of biological polymers may be transferred directly
15 onto a silicon substrate. For example, a colony plate may be transferred onto a substrate having a generic oligonucleotide sequence which hybridizes to another generic complementary sequence contained on all of the vectors into which inserts are cloned. This will specifically only bind those molecules which
20 are actually contained in the vectors containing the desired complementary sequence. This immobilization allows for producing a matrix onto which a sequence specific reagent can bind, or for other purposes. In a further embodiment, a plurality of different vectors each having a specific
25 oligonucleotide attached to the vector may be specifically attached to particular regions on a matrix having a complementary oligonucleotide attached thereto.

VIII. HYBRIDIZATION/SPECIFIC INTERACTION

30 A. General

As discussed previously in the VLSIPS parent applications, the VLSIPS substrates may be used for screening for specific interactions with sequence specific targets or probes.

35 In addition, the availability of substrates having the entire repertoire of possible sequences of a defined length opens up the possibility of sequencing by hybridization. This sequence may be de novo determination of an unknown sequence,

particularly of nucleic acid, verification of a sequence determined by another method, or an investigation of changes in a previously sequenced gene, locating and identifying specific changes. For example, often Maxam and Gilbert sequencing techniques are applied to sequences which have been determined by Sanger and Coulson. Each of those sequencing technologies have problems with resolving particular types of sequences. Sequencing by hybridization may serve as a third and independent method for verifying other sequencing techniques. See, e.g., (1988) Science 242:1245.

In addition, the ability to provide a large repertoire of particular sequences allows use of short subsequence and hybridization as a means to fingerprint a sample. This may be used in a nucleic acid, as well as other polymer embodiments. For example, fingerprinting to a high degree of specificity of sequence matching may be used for identifying highly similar samples, e.g., those exhibiting high homology to the selected probes. This may provide a means for determining classifications of particular sequences. This should allow determination of whether particular genomes of bacteria, phage, or even higher cells might be related to one another.

In addition, fingerprinting may be used to identify an individual source of biological sample. See, e.g., Lander, E. (1989) Nature, 339:501-505, and references therein. For example, a DNA fingerprint may be used to determine whether a genetic sample arose from another individual. This would be particularly useful in various sorts of forensic tests to determine, e.g., paternity or sources of blood samples.

Significant detail on the particulars of genetic fingerprinting for identification purposes are described in, e.g., Morris et al. (1989) "Biostatistical evolution of evidence from continuous allele frequency distribution DNA probes in reference to disputed paternity of identity," J. Forensic Science 34:1311-1317; and Neufeld et al. (1990) Scientific American 262:46-53; each of which is hereby incorporated herein by reference.

In another embodiment, a fingerprinting-like procedure may be used for classifying cell types by analyzing a pattern of specific nucleic acids present in the cell. A series of antibodies may be used to identify cell markers, e.g., proteins, usually on the cell surface, but intracellular markers may also be used. Antigens which are extracellularly expressed are preferred so cell lysis is unnecessary in the screening, but intracellular markers may also be useful. The markers will usually be proteins, but may be nucleic acids, lipids, metabolites, carbohydrates, or other cellular components. See, e.g., Winkelgren, I. (1990) Science News 136:234-237, which indicates extracellular DNA may be common, and suggesting that such might be characteristic of cell types, stage, or physiology. This may also be useful in defining the temporal stage of development of cells, e.g., stem cells or other cells which undergo temporal changes in development. For example, the stage of a cell, or group of cells, may be tested or defined by isolating a sample of mRNA from the population and testing to see what sequences are present in messenger populations. Direct samples, or amplified samples, may be used. Where particular mRNA or other nucleic acid sequences may be characteristic of or shown to be characteristic of particular developmental stages, physiological states, or other conditions, this fingerprinting method may define them. Similar sorts of fingerprinting may be used for determining T-cell classes or perhaps even to generate classification schemes for such proteins as major histocompatibility complex antigens. Thus, the ability to make these substrates allows both the generation of reagents which will be used for defining subclasses or classes of cells or other biological materials, but also provides the mechanisms for selecting those cells which may be found in defined population groups.

Cell classification defined by such a combination of properties, typically expression of extracellular antigens, the present invention also provides the means for isolating homogeneous population of cells. Once the antigenic determinants which define a cell class have been identified, these antigens may be used in a sequential selection process to

isolate only those cells which exhibit the combination of defining structural properties.

The present invention may also be used for mapping sequences within a larger segment. This may be performed by at least two methods, particularly in reference to nucleic acids. Often, enormous segments of DNA are subcloned into a large plurality of subsequences. Ordering these subsequences may be important in determining the overlaps of sequences upon nucleotide determinations. Mapping may be performed by immobilizing particularly large segments onto a matrix using the VLSIPS technology. Alternatively, sequences may be ordered by virtue of subsequences shared by overlapping segments. See, e.g., Craig et al. (1990) Nuc. Acids Res. 18:2653-2660; Michiels et al. (1987) CABIOS 3:203-210; and Olson et al. (1986) Proc. Natl. Acad. Sci. USA 83:7826-7830.

B. Important Parameters

The extent of specific interaction between reagents immobilized to the VLSIPS substrate and another sequence specific reagent may be modified by the conditions of the interaction. Sequencing embodiments typically require high fidelity hybridization and the ability to discriminate perfect matching from imperfect matching. Fingerprinting and mapping embodiments may be performed using less stringent conditions, depending upon the circumstances.

For example, the specificity of antibody/antigen interaction may depend upon such parameters as pH, salt concentration, ionic composition, solvent composition, detergent composition and concentration, and chaotropic agent concentration. See, e.g., Harlow and Lane (1988) Antibodies: A Laboratory Manual, Cold Spring Harbor Press, New York. By careful control of these parameters, the affinity of binding may be mapped across different sequences.

In a nucleic acid hybridization embodiment, the specificity and kinetics of hybridization have been described in detail by, e.g., Wetmur and Davidson (1968) J. Mol. Biol., 31:349-370, Britten and Kohne (1968) Science 161:529-530, and Kanehisa, (1984) Nuc. Acids Res. 12:203-213, each of which is

hereby incorporated herein by reference. Parameters which are well known to affect specificity and kinetics of reaction include salt conditions, ionic composition of the solvent, hybridization temperature, length of oligonucleotide matching sequences, guanine and cytosine (GC) content, presence of hybridization accelerators, pH, specific bases found in the matching sequences, solvent conditions, and addition of organic solvents.

In particular, the salt conditions required for driving highly mismatched sequences to completion typically include a high salt concentration. The typical salt used is sodium chloride (NaCl), however, other ionic salts may be utilized, e.g., KCl. Depending on the desired stringency hybridization, the salt concentration will often be less than about 3 molar, more often less than 2.5 molar, usually less than about 2 molar, and more usually less than about 1.5 molar. For applications directed towards higher stringency matching, the salt concentrations would typically be lower. Ordinary high stringency conditions will utilize salt concentration of less than about 1 molar, more often less than about 750 millimolar, usually less than about 500 millimolar, and may be as low as about 250 or 150 millimolar.

The kinetics of hybridization and the stringency of hybridization both depend upon the temperature at which the hybridization is performed and the temperature at which the washing steps are performed. Temperatures at which steps for low stringency hybridization are desired would typically be lower temperatures, e.g., ordinarily at least about 15°C, more ordinarily at least about 20°C, usually at least about 25°C, and more usually at least about 30°C. For those applications requiring high stringency hybridization, or fidelity of hybridization and sequence matching, temperatures at which hybridization and washing steps are performed would typically be high. For example, temperatures in excess of about 35°C would often be used, more often in excess of about 40°C, usually at least about 45°C, and occasionally even temperatures as high as about 50°C or 60°C or more. Of course, the hybridization of oligonucleotides may be disrupted by even

higher temperatures. Thus, for stripping of targets from substrates, as discussed below, temperatures as high as 80°C, or even higher may be used.

The base composition of the specific oligonucleotides involved in hybridization affects the temperature of melting, and the stability of hybridization as discussed in the above references. However, the bias of GC rich sequences to hybridize faster and retain stability at higher temperatures can be compensated for by the inclusion in the hybridization incubation or wash steps of various buffers. Sample buffers which accomplish this result include the triethyl- and trimethyl ammonium buffers. See, e.g., Wood et al. (1987) Proc. Natl. Acad. Sci. USA, 82:1585-1588, and Khrapko, K. et al. (1989) FEBS Letters 256:118-122.

The rate of hybridization can also be affected by the inclusion of particular hybridization accelerators. These hybridization accelerators include the volume exclusion agents characterized by dextran sulfate, or polyethylene glycol (PEG). Dextran sulfate is typically included at a concentration of between 1% and 40% by weight. The actual concentration selected depends upon the application, but typically a faster hybridization is desired in which the concentration is optimized for the system in question. Dextran sulfate is often included at a concentration of between 0.5% and 2% by weight or dextran sulfate at a concentration between about 0.5% and 5%. Alternatively, proteins which accelerate hybridization may be added, e.g., the recA protein found in *E. coli*) or other homologous proteins.

With respect to those embodiments where specific reagents are not oligonucleotides, the conditions of specific interaction would depend on the affinity of binding between the specific reagent and its target. Typically parameters which would be of particular importance would be pH, salt concentration anion and cation compositions, buffer concentration, organic solvent inclusion, detergent concentration, and inclusion of such reagents such as chaotropic agents. In particular, the affinity of binding may be tested over a variety of conditions by multiple washes and

repeat scans or by using reagents with differences in binding affinity to determine which reagents bind or do not bind under the selected binding and washing conditions. The spectrum of binding affinities may provide an additional dimension of information which may be very useful in identification purposes and mapping.

Of course, the specific hybridization conditions will be selected to correspond to a discriminatory condition which provides a positive signal where desired but fails to show a positive signal at affinities where interaction is not desired. This may be determined by a number of titration steps or with a number of controls which will be run during the hybridization and/or washing steps to determine at what point the hybridization conditions have reached the stage of desired specificity.

IX. DETECTION METHODS

Methods for detection depend upon the label selected. The criteria for selecting an appropriate label are discussed below, however, a fluorescent label is preferred because of its extreme sensitivity and simplicity. Standard labeling procedures are used to determine the positions where interactions between a sequence and a reagent take place. For example, if a target sequence is labeled and exposed to a matrix of different probes, only those locations where probes do interact with the target will exhibit any signal. Alternatively, other methods may be used to scan the matrix to determine where interaction takes place. Of course, the spectrum of interactions may be determined in a temporal manner by repeated scans of interactions which occur at each of a multiplicity of conditions. However, instead of testing each individual interaction separately, a multiplicity of sequence interactions may be simultaneously determined on a matrix.

A. Labeling Techniques

The target polynucleotide may be labeled by any of a number of convenient detectable markers. A fluorescent label is preferred because it provides a very strong signal with low

background. It is also optically detectable at high resolution and sensitivity through a quick scanning procedure. Other potential labeling moieties include, radioisotopes, chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, magnetic labels, and linked enzymes.

Another method for labeling may bypass any label of the target sequence. The target may be exposed to the probes, and a double strand hybrid is formed at those positions only. Addition of a double strand specific reagent will detect where hybridization takes place. An intercalative dye such as ethidium bromide may be used as long as the probes themselves do not fold back on themselves to a significant extent forming hairpin loops. See, e.g., Sheldon et al. (1986) U.S. Pat. No. 4,582,789. However, the length of the hairpin loops in short oligonucleotide probes would typically be insufficient to form a stable duplex.

In another embodiment, different targets may be simultaneously sequenced where each target has a different label. For instance, one target could have a green fluorescent label and a second target could have a red fluorescent label. The scanning step will distinguish sites of binding of the red label from those binding the green fluorescent label. Each sequence can be analyzed independently from one another.

Suitable chromogens will include molecules and compounds which absorb light in a distinctive range of wavelengths so that a color may be observed, or emit light when irradiated with radiation of a particular wave length or wave length range, e.g., fluorescers. Biliproteins, e.g., ficolerythrin, may also serve as labels.

A wide variety of suitable dyes are available, being primary chosen to provide an intense color with minimal absorption by their surroundings. Illustrative dye types include quinoline dyes, triarylmethane dyes, acridine dyes, alizarine dyes, phthaleins, insect dyes, azo dyes, anthraquinoid dyes, cyanine dyes, phenazathionium dyes, and phenazoxonium dyes.

A wide variety of fluorescers may be employed either by themselves or in conjunction with quencher molecules.

Fluorescers of interest fall into a variety of categories having certain primary functionalities. These primary

- 5 functionalities include 1- and 2-aminonaphthalene, p,p'-diaminostilbenes, pyrenes, quaternary phenanthridine salts, 9-aminoacridines, p,p'-diaminobenzophenone imines, anthracenes, oxacarbocyanine, merocyanine, 3-aminoequilenin, perylene, bis-benzoxazole, bis-p-oxazolyl benzene, 1,2-benzophenazin,
- 10 retinol, bis-3-aminopyridinium salts, hellebrigenin, tetracycline, sterophenol, benzimidazolylphenylamine, 2-oxo-3-chromen, indole, xanthen, 7-hydroxycoumarin, phenoxazine, salicylate, strophanthidin, porphyrins, triarylmethanes and flavin. Individual fluorescent compounds which have
- 15 functionalities for linking or which can be modified to incorporate such functionalities include, e.g., dansyl chloride; fluoresceins such as 3,6-dihydroxy-9-phenylxanthhydrol; rhodamineisothiocyanate; N-phenyl 1-amino-8-sulfonatonaphthalene; N-phenyl 2-amino-6-
- 20 sulfonatonaphthalene; 4-acetamido-4-isothiocyanato-stilbene-2,2'-disulfonic acid; pyrene-3-sulfonic acid; 2-toluidinonaphthalene-6-sulfonate; N-phenyl, N-methyl 2-aminoaphthalene-6-sulfonate; ethidium bromide; stebrine; auromine-0,2-(9'-anthroyl)palmitate; dansyl
- 25 phosphatidylethanolamine; N,N'-dioctadecyl oxacarbocyanine; N,N'-dihexyl oxacarbocyanine; merocyanine, 4-(3'pyrenyl)butyrate; d-3-aminodesoxy-equilenin; 12-(9'-anthroyl)stearate; 2-methylanthracene; 9-vinylanthracene; 2,2'-(vinylene-p-phenylene)bisbenzoxazole; p-bis[2-(4'-methyl-5-
- 30 phenyl-oxazolyl)]benzene; 6-dimethylamino-1,2-benzophenazin; retinol; bis(3'-aminopyridinium) 1,10-decandiyl diiodide; sulfonaphthylhydrazone of hellibrienin; chlorotetracycline; N-(7-dimethylamino-4-methyl-2-oxo-3-chromenyl)maleimide; N-[p-(2-benzimidazolyl)-phenyl]maleimide; N-(4-
- 35 fluoranthyl)maleimide; bis(homovanillic acid); resazarin; 4-chloro-7-nitro-2,1,3-benzooxadiazole; merocyanine 540; resorufin; rose bengal; and 2,4-diphenyl-3(2H)-furanone.

Desirably, fluorescers should absorb light above about 300 nm, preferably about 350 nm, and more preferably above about 400 nm, usually emitting at wavelengths greater than about 10 nm higher than the wavelength of the light absorbed. It should be noted that the absorption and emission characteristics of the bound dye may differ from the unbound dye. Therefore, when referring to the various wavelength ranges and characteristics of the dyes, it is intended to indicate the dyes as employed and not the dye which is unconjugated and characterized in an arbitrary solvent.

Fluorescers are generally preferred because by irradiating a fluorescer with light, one can obtain a plurality of emissions. Thus, a single label can provide for a plurality of measurable events.

Detectable signal may also be provided by chemiluminescent and bioluminescent sources. Chemiluminescent sources include a compound which becomes electronically excited by a chemical reaction and may then emit light which serves as the detectible signal or donates energy to a fluorescent acceptor. A diverse number of families of compounds have been found to provide chemiluminescence under a variety of conditions. One family of compounds is 2,3-dihydro-1,4-phthalazinedione. The most popular compound is luminol, which is the 5-amino compound. Other members of the family include the 5-amino-6,7,8-trimethoxy- and the dimethylamino[ca]benz analog. These compounds can be made to luminesce with alkaline hydrogen peroxide or calcium hypochlorite and base. Another family of compounds is the 2,4,5-triphenylimidazoles, with lophine as the common name for the parent product. Chemiluminescent analogs include para-dimethylamino and -methoxy substituents. Chemiluminescence may also be obtained with oxalates, usually oxalyl active esters, e.g., p-nitrophenyl and a peroxide, e.g., hydrogen peroxide, under basic conditions. Alternatively, luciferins may be used in conjunction with luciferase or lucigenins to provide bioluminescence.

Spin labels are provided by reporter molecules with an unpaired electron spin which can be detected by electron

spin resonance (ESR) spectroscopy. Exemplary spin labels include organic free radicals, transitional metal complexes, particularly vanadium, copper, iron, and manganese, and the like. Exemplary spin labels include nitroxide free radicals.

5

B. Scanning System

With the automated detection apparatus, the correlation of specific positional labeling is converted to the presence on the target of sequences for which the reagents have specificity of interaction. Thus, the positional information is directly converted to a database indicating what sequence interactions have occurred. For example, in a nucleic acid hybridization application, the sequences which have interacted between the substrate matrix and the target molecule can be directly listed from the positional information. The detection system used is described in U.S.S.N. 07/649,642 (VLSIPS CIP); and U.S.S.N. __/__, attorney docket number 11509-28 (automated VLSIPS). Although the detection described therein is a fluorescence detector, the detector may be replaced by a spectroscopic or other detector. The scanning system may make use of a moving detector relative to a fixed substrate, a fixed detector with a moving substrate, or a combination. Alternatively, mirrors or other apparatus can be used to transfer the signal directly to the detector. See, e.g., U.S.S.N. __/__, attorney docket number 11509-28 (automated VLSIPS), which is hereby incorporated herein by reference.

The detection method will typically also incorporate some signal processing to determine whether the signal at a particular matrix position is a true positive or may be a spurious signal. For example, a signal from a region which has actual positive signal may tend to spread over and provide a positive signal in an adjacent region which actually should not have one. This may occur, e.g., where the scanning system is not properly discriminating with sufficiently high resolution in its pixel density to separate the two regions. Thus, the signal over the spatial region may be evaluated pixel by pixel to determine the locations and the actual extent of positive signal. A true positive signal should, in theory, show a

uniform signal at each pixel location. Thus, processing by plotting number of pixels with actual signal intensity should have a clearly uniform signal intensity. Regions where the signal intensities show a fairly wide dispersion, may be particularly suspect and the scanning system may be programmed to more carefully scan those positions.

In another embodiment, as the sequence of a target is determined at a particular location, the overlap for the sequence would necessarily have a known sequence. Thus, the system can compare the possibilities for the next adjacent position and look at these in comparison with each other. Typically, only one of the possible adjacent sequences should give a positive signal and the system might be programmed to compare each of these possibilities and select that one which gives a strong positive. In this way, the system can also simultaneously provide some means of measuring the reliability of the determination by indicating what the average signal to background ratio actually is.

More sophisticated signal processing techniques can be applied to the initial determination of whether a positive signal exists or not. See, e.g., U.S.S.N. __/__, __, attorney docket number 11509-28 (automated VLSIPS).

From a listing of those sequences which interact, data analysis may be performed on a series of sequences. For example, in a nucleic acid sequence application, each of the sequences may be analyzed for their overlap regions and the original target sequence may be reconstructed from the collection of specific subsequences obtained therein. Other sorts of analyses for different applications may also be performed, and because the scanning system directly interfaces with a computer the information need not be transferred manually. This provides for the ability to handle large amounts of data with very little human intervention. This, of course, provides significant advantages over manual manipulations. Increased throughput and reproducibility is thereby provided by the automation of vast majority of steps in any of these applications.

XI. DATA ANALYSIS

A. General

Data analysis will typically involve aligning the proper sequences with their overlaps to determine the target sequence. Although the target "sequence" may not specifically correspond to any specific molecule, especially where the target sequence is broken and fragmented up in the sequencing process, the sequence corresponds to a contiguous sequence of the subfragments.

The data analysis can be performed by a computer using an appropriate program. See, e.g., Drmanac, R. et al. (1989) Genomics 4:114-128; and a commercially available analysis program available from the Genetic Engineering Center, P.O. Box 794, 11000 Belgrade, Yugoslavia. Although the specific manipulations necessary to reassemble the target sequence from fragments may take many forms, one embodiment uses a sorting program to sort all of the subsequences using a defined hierarchy. The hierarchy need not necessarily correspond to any physical hierarchy, but provides a means to determine, in order, which subfragments have actually been found in the target sequence. In this manner, overlaps can be checked and found directly rather than having to search throughout the entire set after each selection process. For example, where the oligonucleotide probes are 10-mers, the first 9 positions can be sorted. A particular subsequence can be selected as in the examples, to determine where the process starts. As analogous to the theoretical example provided above, the sorting procedure provides the ability to immediately find the position of the subsequence which contains the first 9 positions and can compare whether there exists more than 1 subsequence during the first 9 positions. In fact, the computer can easily generate all of the possible target sequences which contain given combination of subsequences. Typically there will be only one, but in various situations, there will be more.

An exemplary flow chart for a sequencing program is provided in Figure 4. In general terms, the program provides for automated scanning of the substrate to determine the

positions of probe and target interaction. Simple processing of the intensity of the signal may be incorporated to filter out clearly spurious signals. The positions with positive interaction are correlated with the sequence specificity of specific matrix positions, to generate the set of matching subsequences. This information is further correlated with other target sequence information, e.g., restriction fragment analysis. The sequences are then aligned using overlap data, thereby leading to possible corresponding target sequences which will, optimally, correspond to a single target sequence.

B. Hardware

A variety of computer systems may be used to run a sequencing program. The program may be written to provide both the detecting and scanning steps together and will typically be dedicated to a particular scanning apparatus. However, the components and functional steps may be separated and the scanning system may provide an output, e.g., through tape or an electronic connection into a separate computer which separately runs the sequencing analysis program. The computer may be any of a number of machines provided by standard computer manufacturers, e.g., IBM compatible machines, AppleTM machines, VAX machines, and others, which may often use a UNIXTM operating system. Of course, the hardware used to run the analysis program will typically determine what programming language would be used.

C. Software

Software would be easily developed by a person of ordinary skill in the programming art, following the flow chart provided, or based upon the input provided and the desired result.

Of course, an exemplary embodiment is a polynucleotide sequence system. However, the theoretical and mathematical manipulations necessary for data analysis of other linear molecules, such as polypeptides, carbohydrates, and various other polymers are conceptually similar. Simple branching polymers will usually also be sequencable using

similar technology. However, where there is branching, it may be desired that additional recognition reagents be used to determine the nature and location of branches. This can easily be provided by use of appropriate specific reagents which would be generated by methods similar to those used to produce specific reagents for linear polymers.

XII. SUBSTRATE REUSE

Where a substrate is made with specific reagents that are relatively insensitive to the handling and processing steps involved in a single cycle of use, the substrate may often be reused. The target molecules are usually stripped off of the solid phase specific recognition molecules. Of course, it is preferred that the manipulations and conditions be selected as to be mild and to not affect the substrate. For example, if a substrate is acid labile, a neutral pH would be preferred in all handling steps. Similar sensitivities would be carefully respected where recycling is desired.

A. Removal of Label

Typically for a recycling, the previously attached specific interaction would be disrupted and removed. This will typically involve exposing the substrate to conditions under which the interaction between probe and target is disrupted. Alternatively, it may be exposed to conditions where the target is destroyed. For example, where the probes are oligonucleotides and the target is a polynucleotide, a heating and low salt wash will often be sufficient to disrupt the interactions. Additional reagents may be added such as detergents, and organic or inorganic solvents which disrupt the interaction between the specific reagents and target. In an embodiment where the specific reagents are antibodies, the substrate may be exposed to a gentle detergent which will denature the specific binding between the antibody and its target. The conditions are selected to avoid severe disruption or destruction of the structure of the antibody and to maintain the specificity of the antibody binding site. Conditions with specific pH, detergent concentration, salt concentration, ionic

concentration, and other parameters may be selected which disrupt the specific interactions.

B. Storage and Preservation

5 As indicated above, the matrix will typically be maintained under conditions where the matrix itself and the linkages and specific reagents are preserved. Various specific preservatives may be added which prevent degradation. For example, if the reagents are acid or base labile, a neutral pH
10 buffer will typically be added. It is also desired to avoid destruction of the matrix by growth of organisms which may destroy organic reagents attached thereto. For this reason, a preservative such as cyanide or azide may be added. However, the chemical preservative should also be selected to preserve
15 the chemical nature of the linkages and other components of the substrate. Typically, a detergent may also be included.

C. Processes to Avoid Degradation of Oligomers

In particular, a substrate comprising a large number
20 of oligomers will be treated in a fashion which is known to maintain the quality and integrity of oligonucleotides. These include storing the substrate in a carefully controlled environment under conditions of lower temperature, cation depletion (EDTA and EGTA), sterile conditions, and inert argon
25 or nitrogen atmosphere.

XIII. INTEGRATED SEQUENCING STRATEGY

A. Initial Mapping Strategy

As indicated above, although the VLSIPS may be
30 applied to sequencing embodiments, it is often useful to integrate other concepts to simplify the sequencing. For example, nucleic acids may be easily sequenced by careful selection of the vectors and hosts used for amplifying and generating the specific target sequences. For example, it may
35 be desired to use specific vectors which have been designed to interact most efficiently with the VLSIPS substrate. This is also important in fingerprinting and mapping strategies. For example, vectors may be carefully selected having particular

complementary sequences which are designed to attach to a genetic or specific oligomer on the substrate. This is also applicable to situations where it is desired to target particular sequences to specific locations on the matrix.

5 In one embodiment, unnatural oligomers may be used to target natural probes to specific locations on the VLSIPS substrate. In addition, particular probes may be generated for the mapping embodiment which are designed to have specific combinations of characteristics. For example, the construction
10 of a mapping substrate may depend upon use of another automated apparatus which takes clones isolated from a chromosome walk and attaches them individually or in bulk to the VLSIPS substrate.

In another embodiment, a variety of specific vectors
15 having known and particular "targeting" sequences adjacent the cloning sites may be individually used to clone a selected probe, and the isolated probe will then be targetable to a site on the VLSIPS substrate with a sequence complementary to the "target" sequence.

20 B. Selection of Smaller Clones

In the fingerprinting and mapping embodiments, the selection of probes may be very important. Significant mathematical analysis may be applied to determine which
25 specific sequences should be used as those probes. Of course, for fingerprinting use, these sequences would be most desired that show significant heterogeneity across the human population. Selection of the specific sequences which would most favorably be utilized will tend to be single copy
30 sequences within the genome.

Various hybridization selection procedures may be applied to select sequences which tend not to be repeated within a genome, and thus would tend to be conserved across individuals. For example, hybridization selections may be made
35 for non-repetitive and single copy sequences. See, e.g., Britten and Kohne (1968) "Repeated Sequences in DNA," Science 161:529-540. On the other hand, it may be desired under certain circumstances to use repeated sequences. For example,

where a fingerprint may be used to identify or distinguish different species, or where repetitive sequences may be diagnostic of specific species, repetitive sequences may be desired for inclusion in the fingerprinting probes. In either
5 case, the sequencing capability will greatly assist in the selection of appropriate sequences to be used as probes.

Also as indicated above, various means for constructing an appropriate substrate may involve either mechanical or automated procedures. The standard VLSIPS
10 automated procedure involves synthesizing oligonucleotides or short polymers directly on the substrate. In various other embodiments, it is possible to attach separately synthesized reagents onto the matrix in an ordered array. Other
15 circumstances may lend themselves to transfer a pattern from a petri plate onto a solid substrate. Also, there are methods for site specifically directing collections of reagents to specific locations using unnatural nucleotides or equivalent sorts of targeting molecules.

While a brute force manual transfer process may be
20 utilized sequentially attaching various samples to successive positions, instrumentation for automating such procedures may also be devised. The automated system for performing such would preferably be relatively easily designed and conceptually easily understood.

25 XIV. COMMERCIAL APPLICATIONS

A. Sequencing

As indicated above, sequencing may be performed either de novo or as a verification of another sequencing
30 method. The present hybridization technology provides the ability to sequence nucleic acids and polynucleotides de novo, or as a means to verify either the Maxam and Gilbert chemical sequencing technique or Sanger and Coulson dideoxy- sequencing techniques. The hybridization method is useful to verify
35 sequencing determined by any other sequencing technique and to closely compare two similar sequences, e.g., to identify and locate sequence differences.

Besides polynucleotide sequencing, the present invention also provides means for sequencing other polymers. This includes polypeptides, carbohydrates, synthetic organic polymers, and other polymers. Again, the sequencing may be
5 either verification or de novo.

Of course, sequencing of can be very important in many different sorts of environments. For example, it will be useful in determining the genetic sequence of particular markers in various individuals. In addition, polymers may be
10 used as markers or for information containing molecules to encode information. For example, a short polynucleotide sequence may be included in large bulk production samples indicating the manufacturer, date, and location of manufacture of a product. For example, various drugs may be encoded with
15 this information with a small number of molecules in a batch. For example, a pill may have somewhere from 10 to 100 to 1,000 or more very short and small molecules encoding this information. When necessary, this information may be decoded from a sample of the material using a polymerase chain reaction
20 (PCR) or other amplification method. This encoding system may be used to provide the origin of large bulky samples without significantly affecting the properties of those samples. For example, chemical samples may also be encoded by this method thereby providing means for identifying the source and
25 manufacturing details of lots. The origin of bulk hydrocarbon samples may be encoded. Production lots of organic compounds such as benzene or plastics may be encoded with a short molecule polymer. Food stuffs may also be encoded using similar marking molecules. Even toxic waste samples can be
30 encoded determining the source or origin. In this way, proper disposal can be traced or more easily enforced.

Similar sorts of encoding may be provided by fingerprinting-type analysis. Whether the resolution is absolute or less so, the concept of coding information on
35 molecules such as nucleic acids, which can be amplified and later decoded, may be a very useful and important application.

This technology also provides the ability to include markers for origins of biological materials. For example, a

patented animal line may be transformed with a particular unnatural sequence which can be traced back to its origin. With a selection of multiple markers, the likelihood could be negligible that a combination of markers would have independently arisen from a source other than the patented or specifically protected source. This technique may provide a means for tracing the actual origin of particular biological materials. Bacteria, plants, and animals will be subject to marking by such encoding sequences.

B. Fingerprinting

As indicated above, fingerprinting technology may also be used for data encryption. Moreover, fingerprinting allows for significant identification of particular individuals. Where the fingerprinting technology is standardized, and used for identification of large numbers of people, related equipment and peripheral processing will be developed to accompany the underlying technology. For example, specific equipment may be developed for automatically taking a biological sample and generating or amplifying the information molecules within the sample to be used in fingerprinting analysis. Moreover, the fingerprinting substrate may be mass produced using particular types of automatic equipment. Synthetic equipment may produce the entire matrix simultaneously by stepwise synthetic methods as provided by the VLSIPS technology. The attachment of specific probes onto a substrate may also be automated, e.g., making use of the caged biotin technology. See, e.g., U.S.S.N. 07/612,671 (caged biotin CIP). As indicated above, there are automated methods for actually generating the matrix and substrate with distinct sequence reagents positionally located at each of the matrix positions. Where such reagents are, e.g., unnatural amino acids, a targeting function may be utilized which does not interfere with aa natural nucleotide functionality.

In addition, peripheral processing may be important and may be dedicated to this specific application. Thus, automated equipment for producing the substrates may be designed, or particular systems which take in a biological

sample and output either a computer readout or an encoded instrument, e.g., a card or document which indicates the information and can provide that information to others. An identification having a short magnetic strip with a few million
5 bits may be used to provide individual identification and important medical information useful in a medical emergency.

In fact, data banks may be set up to correlate all of this information of fingerprinting with medical information. This may allow for the determination of correlations between
10 various medical problems and specific DNA sequences. By collating large populations of medical records with genetic information, genetic propensities and genetic susceptibilities to particular medical conditions may be developed. Moreover, with standardization of substrates, the micro encoding data may
15 be also standardized to reproduce the information from a centralized data bank or on an encoding device carried on an individual person. On the other hand, if the fingerprinting procedure is sufficiently quick and routine, every hospital may routinely perform a fingerprinting operation and from that
20 determine many important medical parameters for an individual.

In particular industries, the VLSIPS sequencing, fingerprinting, or mapping technology will be particularly appropriate. As mentioned above, agricultural livestock suppliers may be able to encode and determine whether their
25 particular strains are being used by others. By incorporating particular markers into their genetic stocks, the markers will indicate origin of genetic material. This is applicable to seed producers, livestock producers, and other suppliers of medical or agricultural biological materials.

This may also be useful in identifying individual animals or plants. For example, these markers may be useful in determining whether certain fish return to their original breeding grounds, whether sea turtles always return to their original birthplaces, or to determine the migration patterns
30 and viability of populations of particular endangered species. It would also provide means for tracking the sources of particular animal products. For example, it might be useful for determining the origins of controlled animal substances
35

such as elephant ivory or particular bird populations whose importation or exportation is controlled.

As indicated above, polymers may be used to encode important information on source and batch and supplier. This is described in greater detail, e.g., "Applications of PCR to industrial problems," (1990) in Chemical and Engineering News 68:145, which is hereby incorporated herein by reference. In fact, the synthetic method can be applied to the storage of enormous amounts of information. Small substrates may encode enormous amounts of information, and its recovery will make use of the inherent replication capacity. For example, on regions of $10\text{ }\mu\text{m} \times 10\text{ }\mu\text{m}$, 1 cm^2 has 10^6 regions. An theory, the entire human genome could be attached in 1000 nucleotide segments on a 3 cm^2 surface. Genomes of endangered species may be stored on these substrates.

Fingerprinting may also be used for genetic tracing or for identifying individuals for forensic science purposes. See, e.g., Morris, J. et al. (1989) "Biostatistical Evaluation of Evidence From Continuous Allele Frequency Distribution DNA Probes in Reference to Disputed Paternity and Identity," J. Forensic Science 34:1311-1317, and references provided therein; each of which is hereby incorporated herein by reference.

In addition, the high resolution fingerprinting allows the distinguishability to high resolution of particular samples. As indicated above, new cell classifications may be defined based on combinations of a large number of properties. Similar applications will be found in distinguishing different species of animals or plants. In fact, microbial identification may become dependent on characterization of the genetic content. Tumors or other cells exhibiting abnormal physiology will be detectable by use of the present invention. Also, knowing the genetic fingerprint of a microorganism may provide very useful information on how to treat an infection by such organism.

Modifications of the fingerprint embodiments may be used to diagnose the condition of the organism. For example, a blood sample is presently used for diagnosing any of a number of different physiological conditions. A multi-dimensional

carbohydrates, and other sorts of molecules can be advantageously utilized using the present technology.

The present invention will be better understood by reference to the following illustrative examples. The

5 following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

- I. Sequencing
 - A. polynucleotide
 - B. polypeptide
 - C. short peptide
 - 1. Herz antibody identification
- II. Fingerprinting
 - A. polynucleotide fingerprint
 - B. peptide fingerprint
 - C. cell classification scheme
 - D. temporal development scheme
 - 1. developmental antigens
 - 2. developmental mRNA expression
 - E. diagnostic test
 - 1. viral identification
 - 2. bacterial identification
 - 3. other microbiological identifications
 - 4. allergy test (immobilized antigens)
 - F. individual (animal/plant) identification
 - 1. genetic
 - 2. immunological
 - G. genetic screen
 - 1. test alleles with markers
 - 2. amniocentesis
- III. Mapping
 - A. positionally located clones (caged biotin)
 - 1. short probes, long targets
 - 2. long targets, short probes
 - B. positionally defined clones
- IV. Conclusion

* * * * *

Relevant applications whose techniques are incorporated herein by reference are Pirrung, et al., U.S.S.N. 07/362,901 (VLSIPS parent), filed June 7, 1989; Pirrung et al, U.S.S.N. 07/492,462 (VLSIPS CIP), filed March 7, 1990; Barrett, et al., U.S.S.N. 07/435,316 (caged biotin) filed November 13, 1989; Barrett, et al., U.S.S.N. 07/612,671 (caged biotin CIP), filed November 13, 1990; and commonly assigned and simultaneously filed applications U.S.S.N. __/__, attorney docket number 11509-28 (automated VLSIPS) and U.S.S.N. __/__, attorney docket number 11509-26 (sequencing by synthesis).

Also, additional relevant techniques are described, e.g., in Sambrook, J., et al. (1989) Molecular Cloning: a Laboratory Manual, 2d Ed., vols 1-3, Cold Spring Harbor Press,

- New York; Greenstein and Winitz (1961) Chemistry of the Amino Acids, Wiley and Sons, New York; Bodzansky, M. (1988) Peptide Chemistry: a Practical Textbook, Springer-Verlag, New York; Harlow and Lane (1988) Antibodies: A Laboratory Manual, Cold Spring Harbor Press, New York; Glover, D. (ed.) (1987) DNA Cloning: A Practical Approach, vols 1-3, IRL Press, Oxford; Bishop and Rawlings (1987) Nucleic Acid and Protein Sequence Analysis: A Practical Approach, IRL Press, Oxford; Hames and Higgins (1985) Nucleic Acid Hybridisation: A Practical Approach, IRL Press, Oxford; Wu et al. (1989) Recombinant DNA Methodology, Academic Press, San Diego; Goding (1986) Monoclonal Antibodies: Principles and Practice, (2d ed.), Academic Press, San Diego; Finegold and Barron (1986) Bailey and Scott's Diagnostic Microbiology, (7th ed.), Mosby Co., St. Louis; Collins et al. (1989) Microbiological Methods, (6th ed.), Butterworth, London; Chaplin and Kennedy (1986) Carbohydrate Analysis: A Practical Approach, IRL Press, Oxford; Van Dyke (ed.) (1985) Bioluminescence and Chemiluminescence: Instruments and Applications, vol 1, CRC Press, Boca Rotan; and Ausubel et al. (ed.) (1990) Current Protocols in Molecular Biology, Greene Publishing and Wiley-Interscience, New York; each of which is hereby incorporated herein by reference.

The following examples are provided to illustrate the efficacy of the inventions herein. All operations were conducted at about ambient temperatures and pressures unless indicated to the contrary.

I. SEQUENCING

A. Polynucleotide

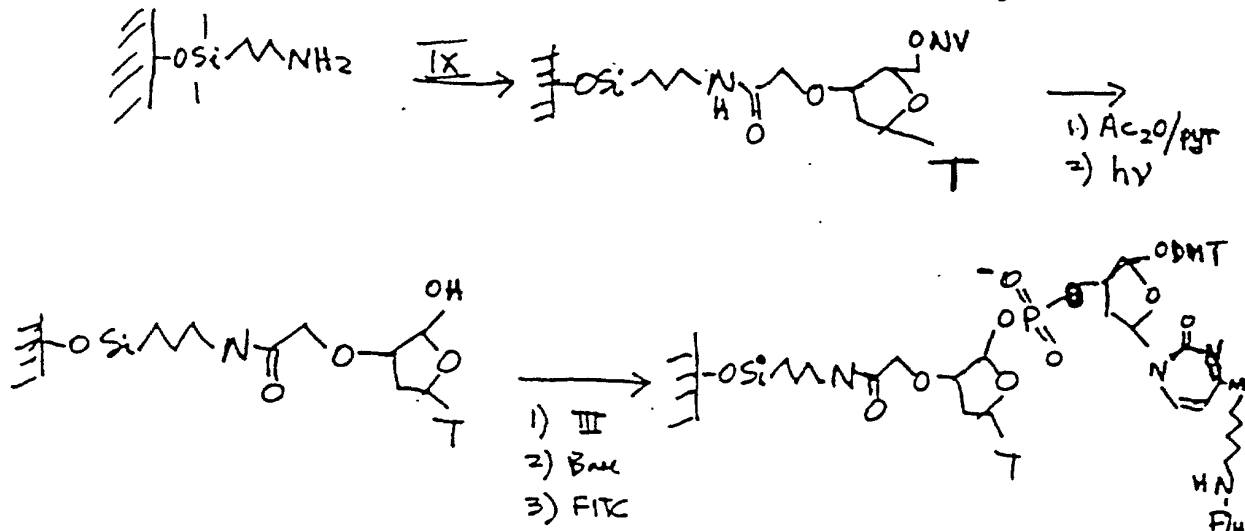
1. HPLC of the photolysis of 5'-O-nitroveratryl-thymidine.

In order to determine the time for photolysis of 5'-O-nitrovertryl thymidine to thymidine a 100 μ M solution of NV-Thym-OH (5'-O-nitrovertryl thymidine) in dioxane was made and ~200 μ l aliquots were irradiated (in a quartz cuvette 1 cm x 2 mm) at 362.3 nm for 20 sec, 40 sec, 60 sec, 2 min, 5 min, 10 min, 15 min, and 20 min. The resulting irradiated mixtures were then analyzed by HPLC using a Varian MicroPak SP column

(C₁₈ analytical) at a flow rate of 1 ml/min and a solvent system of 40% CH₃CN and 60% water. Thymidine has a retention time of 1.2 min and NVO-Thym-OH has a retention time of 2.1 min. It was seen that after 10 min of exposure the
5 deprotection was complete.

2. Preparation and Detection of Thymidine-Cytidine dimer (FITC)

The reaction is illustrated:



To an aminopropylated glass slide (standard VLSIPS) was added a mixture of the following:

- 12.2 mg of NVO-Thym-CO₂H (IX)
- 3.4 mg of HOBT (N-hydroxybenztriazal)
- 8.8 μl DIEA (Diisopropylethylamine)
- 11.1 mg BOP reagent
- 2.5 ml DMF

After 2 h coupling time (standard VLSIPS) the plate was washed, acetylated with acetic anhydride/pyridine, washed,

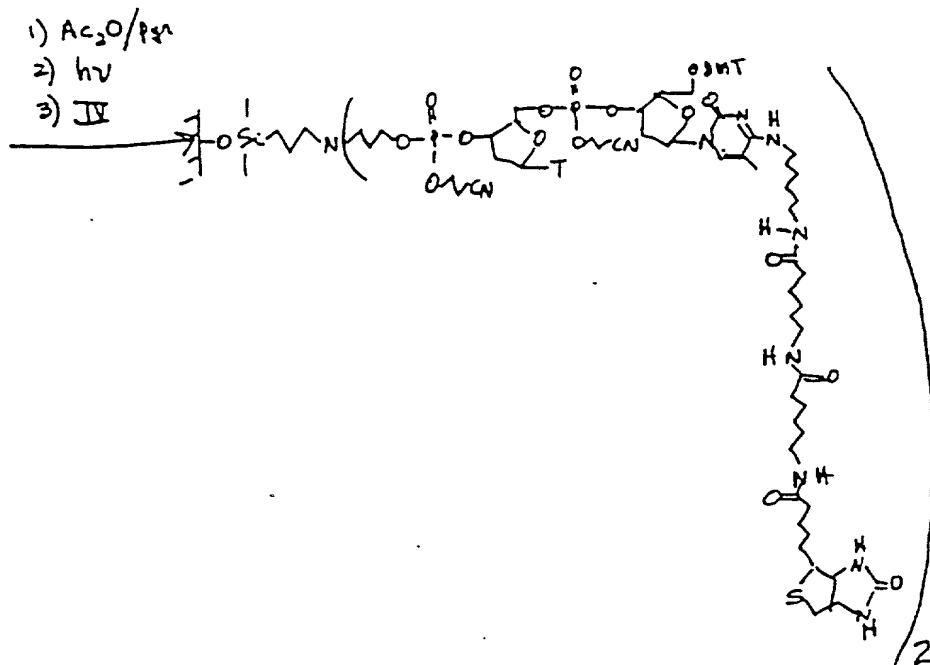
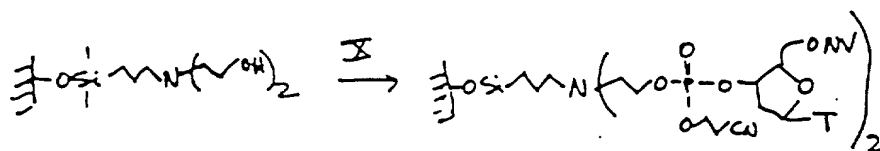
dried, and photolyzed in dioxane at 362 nm at 14 mW/cm² for 10 min using a 500 μm checkerboard mask. The slide was then taken and treated with a mixture of the following:

107 mg of Fmoc-amine modified C (III)

21 mg of tetrazole

1 ml anhydrous CH₃CN

After being treated for approximately 8 min, the slide was washed off with CH₃CN, dried, and oxidized with I₂/H₂O/THF/lutidine for 1 min. The slide was again washed, dried, and treated for 30 min with a 20% solution of DBU in DMF. After thorough rinsing of the slide, it was next exposed to a FITC solution (1mM fluorescein isothiocyanate [FITC] in DMF) for 50 min, then washed, dried, and examined by fluorescence microscopy. This reaction is illustrated:



3. Preparation and Detection of Thymidine-Cytidine dimer (Biotin)

An aminopropyl glass slide, was soaked in a solution of ethylene oxide (20% in DMF) to generate a hydroxylated surface. The slide was added a mixture of the following:

- 32 mg of NVO-T-OCED (X)
- 11 mg of tetrazole
- 0.5 ml of anhydrous CH_3CN

After 8 min the plate was then rinsed with acetonitrile, then oxidized with $\text{I}_2/\text{H}_2\text{O}/\text{THF}/\text{lutidine}$ for 1 min, washed and dried. The slide was then exposed to a 1:3 mixture of acetic anhydride:pyridine for 1 h, then washed and dried. The substrate was then photolyzed in dioxane at 362 nm at 14 mW/cm² for 10 min using a 500 μm checkerboard mask, dried, and then treated with a mixture of the following:

- 65 mg of biotin modified C (IV)
- 11 mg of tetrazole
- 0.5 ml anhydrous CH_3CN

After 8 min the slide was washed with CH_3CN then oxidized with $\text{I}_2/\text{H}_2\text{O}/\text{THF}/\text{lutidine}$ for 1 min, washed, and then dried. The slide was then soaked for 30 min in a PBS/0.05% Tween 20 buffer and the solution then shaken off. The slide was next treated with FITC-labeled streptavidin at 10 $\mu\text{g}/\text{ml}$ in the same buffer system for 30 min. After this time the streptavidin-buffer system was rinsed off with fresh PBS/0.05% Tween 20 buffer and then the slide was finally agitated in distilled water for about 1/2 h. After drying, the slide was examined by fluorescence microscopy (see Fig. 2 and Fig. 3).

4. substrate preparation

Before attachment of reactive groups it is preferred to clean the substrate which is, in a preferred embodiment, a glass substrate such as a microscope slide or cover slip. A roughened surface will be useable but a plastic or other solid substrate is also appropriate. According to one embodiment the slide is soaked in an alkaline bath consisting of, e.g., 1 liter of 95% ethanol with 120 ml of water and 120 grams of sodium hydroxide for 12 hours. The slides are washed with a

buffer and under running water, allowed to air dry, and rinsed with a solution of 95% ethanol.

The slides are then aminated with, e.g., aminopropyltriethoxysilane for the purpose of attaching amino groups to the glass surface on linker molecules, although other omega functionalized silanes could also be used for this purpose. In one embodiment 0.1% aminopropyltriethoxysilane is utilized, although solutions with concentrations from $10^{-7}\%$ to 10% may be used, with about $10^{-3}\%$ to 2% preferred. A 0.1% mixture is prepared by adding to 100 ml of a 95% ethanol/5% water mixture, 100 microliters (μ l) of aminopropyltriethoxysilane. The mixture is agitated at about ambient temperature on a rotary shaker for an appropriate amount of time, e.g., about 5 minutes. 500 μ l of this mixture is then applied to the surface of one side of each cleaned slide. After 4 minutes or more, the slides are decanted of this solution and thoroughly rinsed three times or more by dipping in 100% ethanol.

After the slides dry, they are heated in a 110-120°C vacuum oven for about 20 minutes, and then allowed to cure at room temperature for about 12 hours in an argon environment. The slides are then dipped into DMF (dimethylformamide) solution, followed by a thorough washing with methylene chloride.

25

5. linker attachment, blocking of free sites

The aminated surface of the slide is then exposed to about 500 μ l of, for example, a 30 millimolar (mM) solution of NVOC-nucleotide- NHS (N-hydroxysuccinimide) in DMF for attachment of a NVOC-nucleotide to each of the amino groups. See, e.g., SIGMA Chemical Company for various nucleotide derivatives. The surface is washed with, for example, DMF, methylene chloride, and ethanol.

Any unreacted aminopropyl silane on the surface, i.e., those amino groups which have not had the NVOC-nucleotide attached, are now capped with acetyl groups (to prevent further reaction) by exposure to a 1:3 mixture of acetic anhydride in pyridine for 1 hour. Other materials which may perform this

Of course, greater diversity is obtained by using masking strategies which will also include the synthesis of polymers having a length of less than ℓ . If, in the extreme case, all polymers having a length less than or equal to ℓ are synthesized, the number of polymers synthesized will be:

$$n^{\ell} + n^{\ell-1} + \dots + n^1. \quad (3)$$

The maximum number of lithographic steps needed will generally be n^{ℓ} for each "layer" of monomers, i.e., the total number of masks (and, therefore, the number of lithographic steps) needed will be $n^{\ell} \times \ell$. The size of the transparent mask regions will vary in accordance with the area of the substrate available for synthesis and the number of sequences to be formed. In general, the size of the synthesis areas will be:

$$\text{size of synthesis areas} = (A)/(S)$$

where:

A is the total area available for synthesis; and
S is the number of sequences desired in the area.

It will be appreciated by those of skill in the art that the above method could readily be used to simultaneously produce thousands or millions of oligomers on a substrate using the photolithographic techniques disclosed herein. Consequently, the method results in the ability to practically test large numbers of, for example, di, tri, tetra, penta, hexa, hepta, octa, nona, deca, even dodecanucleotides, or larger polynucleotides (or correspondingly, polypeptides).

The above example has illustrated the method by way of a manual example. It will of course be appreciated that automated or semi-automated methods could be used. The substrate would be mounted in a flow cell for automated addition and removal of reagents, to minimize the volume of reagents needed, and to more carefully control reaction conditions. Successive masks will be applicable manually or automatically. See, e.g., U.S.S.N. 07/492,462 (VLSIPS CIP) and U.S.S.N. ____/____, attorney docket number 11509-28 (automated VLSIPS).

7. labeling of target

The target oligonucleotide can be labeled using standard procedures referred to above. As discussed, for certain situations, a reagent which recognizes interaction, e.g., ethidium bromide, may be provided in the detection step.

5 Alternatively, fluorescence labeling techniques may be applied, see, e.g., Smith, et al. (1986) Nature, 321: 674-679; and Prober, et al. (1987) Science, 238:336-341. The techniques described therein will be followed with minimal modifications as appropriate for the label selected.

10

8. dimers of A, C, G, and T

The described technique may be applied, with photosensitive blocked nucleotides corresponding to adenine, cytosine, guanine, and thymine, to make combinations of
15 polynucleotides consisting of each of the four different nucleotides. All 16 possible dimers would be made using a minor modification of the described method.

9. 10-mers of A, C, G, and T

20 The described technique for making dimers of A, C, G, and T may be further extended to make longer oligonucleotides. The automated system described, e.g., in U.S.S.N. 07/492,462 (VLSIPS CIP), and U.S.S.N. __/__, attorney docket number 11509-28 (automated VLSIPS), can be adapted to make all
25 possible 10-mers composed of the 4 nucleotides A, C, G, and T. The photosensitive, blocked nucleotide analogues have been described above, and would be readily adaptable to longer oligonucleotides.

30

10. specific recognition hybridization to 10-mers

The described hybridization conditions are directly applicable to the sequence specific recognition reagents attached to the substrate, produced as described immediately
35 above. The 10-mers have an inherent property of hybridizing to a complementary sequence. For optimum discrimination between full matching and some mismatch, the conditions of hybridization should be carefully selected, as described above. Careful control of the conditions, and titration of parameters

should be performed to determine the optimum collective conditions.

11. hybridization

5 Hybridization conditions are described in detail, e.g., in Hames and Higgins (1985) Nucleic Acid Hybridisation: A Practical Approach; and the considerations for selecting particular conditions are described, e.g., in Wetmur and Davidson, (1988) J. Mol. Biol. 31:349-370, and Wood et al. 10 (1985) Proc. Natl. Acad. Sci. USA 82:1585-1588. As described above, conditions are desired which can distinguish matching along the entire length of the probe from where there is one or more mismatched bases. The length of incubation and conditions will be similar, in many respects, to the hybridization 15 conditions used in Southern blot transfers. Typically, the GC bias may be minimized by the introduction of appropriate concentrations of the alkylammonium buffers, as described above.

20 Titration of the temperature and other parameters is desired to determine the optimum conditions for specificity and distinguishability of absolutely matched hybridization from mismatched hybridization.

25 A fluorescently labeled target or set of targets are generated, as described in Prober, et al. (1987) Science 238:336-341, or Smith, et al. (1986) Nature 321:674-679. Preferably, the target or targets are of the same length as, or slightly longer, than the oligonucleotide probes attached to the substrate and they will have known sequences. Thus, only a few of the probes hybridize perfectly with the target, and 30 which particular ones did would be known.

 The substrate and probes are incubated under appropriate conditions for a sufficient period of time to allow hybridization to completion. The time is measured to determine when the probe-target hybridizations have reached completion.

35 A salt buffer which minimizes GC bias is preferred, incorporating, e.g., buffer, such as tetramethyl ammonium or tetraethyl ammonium ion at between about 2.4 and 3.0 M. See Wood, et al. (1985) Proc. Nat'l Acad. Sci. USA 82:1585-1588.

This time is typically at least about 30 min, and may be as long as about 1-5 days. Typically very long matches will hybridize more quickly, very short matches will hybridize less quickly, depending upon relative target and probe concentrations. The hybridization will be performed under conditions where the reagents are stable for that time duration.

Upon maximal hybridization, the conditions for washing are titrated. Three parameters initially titrated are time, temperature, and cation concentration of the wash step. The matrix is scanned at various times to determine the conditions at which the distinguishability between true perfect hybrid and mismatched hybrid is optimized. These conditions will be preferred in the sequencing embodiments.

12. positional detection of specific interaction

As indicated above, the detection of specific interactions may be performed by detecting the positions where the labeled target sequences are attached. Where the label is a fluorescent label, the apparatus described, e.g., in U.S.S.N. 07/492,462 (VLSIPS CIP); and U.S.S.N. __/__, attorney docket number 11509-28, may be advantageously applied. In particular, the synthetic processes described above will result in a matrix pattern of specific sequences attached to the substrate, and a known pattern of interactions can be converted to corresponding sequences.

In an alternative embodiment, a separate reagent which differentially interacts with the probe and interacted probe/targets can indicate where interaction occurs or does not occur. A single-strand specific reagent will indicate where no interaction has taken place, while a double-strand specific reagent will indicate where interaction has taken place. An intercalating dye, e.g., ethidium bromide, may be used to indicate the positions of specific interaction.

13. analysis

Conversion of the positional data into sequence specificity will provide the set of subsequences whose analysis

by overlap segments, may be performed, as described above. Analysis is provided by the methodology described above, or using, e.g., software available from the Genetic Engineering Center, P.O. Box 794, 11000 Belgrade, Yugoslavia (Yugoslav group). See, also, Macevycz, PCT publication no. WO 90/04652, which is hereby incorporated herein by reference.

B. Polypeptide

The description of the preparation of short peptides on a substrate incorporates by reference sections in U.S.S.N. 07/492,462 (VLSIPS CIP), and described below.

1. slide preparation

Preparation of the substrate follows that described above for nucleotides.

2. linker attachment, blocking of free sites

The aminated surface of the slide is exposed to about 500 μ l of, e.g., a 30 millimolar (mM) solution of NVOC-GABA (gamma amino butyric acid) NHS (N-hydroxysuccinimide) in DMF for attachment of a NVOC-GABA to each of the amino groups. The surface is washed with, for example, DMF, methylene chloride, and ethanol. See U.S.S.N. __/__, attorney docket number 11509-28, for details on amino acid chemistry.

Any unreacted aminopropyl silane on the surface, i.e., those amino groups which have not had the NVOC-GABA attached, are now capped with acetyl groups (to prevent further reaction) by exposure to a 1:3 mixture of acetic anhydride in pyridine for 1 hour. Other materials which may perform this residual capping function include trifluoroacetic anhydride, formicacetic anhydride, or other reactive acylating agents. Finally, the slides are washed again with DMF, methylene chloride, and ethanol.

3. synthesis of 8 trimers of "A" and "B"

See U.S.S.N. 07/492,462 (VLSIPS CIP) which describes the preparation of glycine and phenylalanine trimers. The technique is similar to the method described above for making

triners of C and T, but substituting photosensitive blocked glycine for the C derivative and photosensitive blocked phenylalamine for the T derivative.

5 4. synthesis of a dimer of an aminopropyl group and a fluorescent group

 In synthesizing the dimer of an aminopropyl group and a fluorescent group, a functionalized durapore membrane was used as a substrate. The Durapore membrane was a
10 polyvinylidene difluoride with aminopropyl groups. The aminopropyl groups were protected with the DDZ group by reaction of the carbonyl chloride with the amino groups, a reaction readily known to those of skill in the art.² The surface bearing these groups was placed in a solution of THF and
15 contacted with a mask bearing a checkerboard pattern of 1 mm opaque and transparent regions. The mask was exposed to ultraviolet light having a wavelength down to at least about 280 nm for about 5 minutes at ambient temperature, although a wide range of exposure times and temperatures may be
20 appropriate in various embodiments of the invention. For example, in one embodiment, an exposure time of between about 1 and 5000 seconds may be used at process temperatures of between -70 and +50°C.

 In one preferred embodiment, exposure times of
25 between about 1 and 500 seconds at about ambient pressure are used. In some preferred embodiments, pressure above ambient is used to prevent evaporation.

 The surface of the membrane was then washed for about 1 hour with a fluorescent label which included an active ester
30 bound to a chelate of a lanthanide. Wash times will vary over a wide range of values from about a few minutes to a few hours. These materials fluoresce in the red and the green visible region. After the reaction with the active ester in the fluorophore was complete, the locations in which the
35 fluorophore was bound could be visualized by exposing them to ultraviolet light and observing the red and the green fluorescence. It was observed that the derivatized regions of the substrate closely corresponded to the original pattern of the mask.

5. demonstration of signal capability

Signal detection capability was demonstrated using a low-level standard fluorescent bead kit manufactured by Flow Cytometry Standards and having model no. 824. This kit includes 5.8 μm diameter beads, each impregnated with a known number of fluorescein molecules.

One of the beads was placed in the illumination field on the scan stage in a field of a laser spot which was initially shuttered. After being positioned in the illumination field, the photon detection equipment was turned on. The laser beam was unblocked and it interacted with the particle bead, which then fluoresced. Fluorescence curves of beads impregnated with 7,000 and 29,000 fluorescein molecules, are shown in Figs. 11A and 11B, respectively of U.S.S.N. 07/492,462 (VLSIPS CIP). On each curve, traces for beads without fluorescein molecules are also shown. These experiments were performed with 488 nm excitation, with 100 μW of laser power. The light was focused through a 40 power 0.75 NA objective.

The fluorescence intensity in all cases started off at a high value and then decreased exponentially. The fall-off in intensity is due to photobleaching of the fluorescein molecules. The traces of beads without fluorescein molecules are used for background subtraction. The difference in the initial exponential decay between labeled and nonlabeled beads is integrated to give the total number of photon counts, and this number is related to the number of molecules per bead. Therefore, it is possible to deduce the number of photons per fluorescein molecule that can be detected. This calculation indicates the radiation of about 40 to 50 photons per fluorescein molecule are detected.

6. determination of the number of molecules per unit area

Aminopropylated glass microscope slides prepared according to the methods discussed above were utilized in order to establish the density of labeling of the slides. The free amino termini of the slides were reacted with FITC (fluorescein

isothiocyanate) which forms a covalent linkage with the amino group. The slide is then scanned to count the number of fluorescent photons generated in a region which, using the estimated 40-50 photons per fluorescent molecule, enables the calculation of the number of molecules which are on the surface per unit area.

A slide with aminopropyl silane on its surface was immersed in a 1 mM solution of FITC in DMF for 1 hour at about ambient temperature. After reaction, the slide was washed twice with DMF and then washed with ethanol, water, and then ethanol again. It was then dried and stored in the dark until it was ready to be examined.

Through the use of curves similar to those shown in Fig. 11 of U.S.S.N. 07/492,462 (VLSIPS CIP), and by integrating the fluorescent counts under the exponentially decaying signal, the number of free amino groups on the surface after derivitization was determined. It was determined that slides with labeling densities of 1 fluorescein per $10^3 \times 10^3$ to $\sim 2 \times 2$ nm could be reproducibly made as the concentration of aminopropyltriethoxysilane varied from $10^{-5}\%$ to $10^{-1}\%$.

7. removal of NOVC and attachment of a fluorescent marker

NVOC-GABA groups were attached as described above.

The entire surface of one slide was exposed to light so as to expose a free amino group at the end of the gamma amino butyric acid. This slide, and a duplicate which was not exposed, were then exposed to fluorescein isothiocyanate (FITC).

Fig. 12A of U.S.S.N. 07/492,462 (VLSIPS CIP) illustrates the slide which was not exposed to light, but which was exposed to FITC. The units of the x axis are time and the units of the y axis are counts. The trace contains a certain amount of background fluorescence. The duplicate slide was exposed to 350 nm broadband illumination for about 1 minute (12 mW/cm^2 , $\sim 350 \text{ nm}$ illumination), washed and reacted with FITC. A large increase in the level of fluorescence is observed, which indicates photolysis has exposed a number of amino groups on the surface of the slides for attachment of a fluorescent marker.

8. use of a mask in removal of NVOC

The next experiment was performed with a 0.1% aminopropylated slide. Light from a Hg-Xe arc lamp was imaged
5 onto the substrate through a laser-ablated chrome-on-glass mask in direct contact with the substrate.

This slide was illuminated for approximately 5 minutes, with 12 mW of 350 nm broadband light and then reacted with the 1 mM FITC solution. It was put on the laser detection
10 scanning stage and a graph was plotted as a two-dimensional representation of position color-coded for fluorescence intensity. The experiment was repeated a number of times through various masks. The fluorescence patterns for a 100x100 μm mask, a 50 μm mask, a 20 μm mask, and a 10 μm mask indicate
15 that the mask pattern is distinct down to at least about 10 μm squares using this lithographic technique.

9. attachment of YGGFL and subsequent exposure to herz antibody and goat anti-mouse antibody

20

In order to establish that receptors to a particular polypeptide sequence would bind to a surface-bound peptide and be detected, Leu enkephalin was coupled to the surface and recognized by an antibody. A slide was derivatized with 0.1% amino propyl-triethoxysilane and protected with NVOC. A 500 μm
25 checkerboard mask was used to expose the slide in a flow cell using backside contact printing. The Leu enkephalin sequence (H_2N -tyrosine, glycine, glycine, phenylalanine, leucine-COOH, otherwise referred to herein as YGGFL) was attached via its
30 carboxy end to the exposed amino groups on the surface of the slide. The peptide was added in DMF solution with the BOP/HOBT/DIEA coupling reagents and recirculated through the flow cell for 2 hours at room temperature.

A first antibody, known as the Herz antibody, was
35 applied to the surface of the slide for 45 minutes at 2 $\mu\text{g}/\text{ml}$ in a supercocktail (containing 1% BSA and 1% ovalbumin also in this case). A second antibody, goat anti-mouse fluorescein conjugate, was then added at 2 $\mu\text{g}/\text{ml}$ in the supercocktail buffer, and allowed to incubate for 2 hours.

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The results of this experiment were plotted as fluorescence intensity as a function of position. This image was taken at 10 μm steps and showed that not only can deprotection be carried out in a well defined pattern, but also that (1) the method provided for successful coupling of peptides to the surface of the substrate, (2) the surface of a bound peptide was available for binding with an antibody, and (3) that the detection apparatus capabilities were sufficient to detect binding of a receptor. Moreover, the Herz antibody is a sequence specific reagent which may be used advantageously as a sequence specific recognition reagent. It may be used, if specificity is high, for sequencing purposes, and, at least, for fingerprinting and mapping uses.

10. monomer-by-monomer formation of YGGFL and subsequent exposure to labeled antibody

Monomer-by-monomer synthesis of YGGFL and GGFL in alternate squares was performed on a slide in a checkerboard pattern and the resulting slide was exposed to the Herz antibody.

A slide is derivatized with the aminopropyl group, protected in this case with t-BOC (t-butoxycarbonyl). The slide was treated with TFA to remove the t-BOC protecting group. E-aminocaproic acid, which was t-BOC protected at its amino group, was then coupled onto the aminopropyl groups. The aminocaproic acid serves as a spacer between the aminopropyl group and the peptide to be synthesized. The amino end of the spacer was deprotected and coupled to NVOC-leucine. The entire slide was then illuminated with 12 mW of 325 nm broadband illumination. The slide was then coupled with NVOC-phenylalanine and washed. The entire slide was again illuminated, then coupled to NVOC-glycine and washed. The slide was again illuminated and coupled to NVOC-glycine to form the sequence shown in the last portion of Fig. 13A of U.S.S.N. 07/492,462 (VLSIPS CIP).

Alternating regions of the slide were then illuminated using a projection print using a 500x500 μm checkerboard mask; thus, the amino group of glycine was exposed only in the lighted areas. When the next coupling chemistry

step was carried out, NVOC-tyrosine was added, and it coupled only at those spots which had received illumination. The entire slide was then illuminated to remove all the NVOC groups, leaving a checkerboard of YGGFL in the lighted areas and in the other areas, GGFL. The Herz antibody (which recognizes the YGGFL, but not GGFL) was then added, followed by goat anti-mouse fluorescein conjugate.

The resulting fluorescence scan showed dark areas containing the tetrapeptide GGFL, which is not recognized by the Herz antibody (and thus there is no binding of the goat anti-mouse antibody with fluorescein conjugate), and red areas in which YGGFL was present. The YGGFL pentapeptide is recognized by the Herz antibody and, therefore, there is antibody in the lighted regions for the fluorescein-conjugated goat anti-mouse to recognize.

Similar patterns for a 50 μ m mask used in direct contact ("proximity print") with the substrate provided a pattern which was more distinct and the corners of the checkerboard pattern were touching as a result of the mask being placed in direct contact with the substrate (which reflects the increase in resolution using this technique).

11. monomer-by-monomer synthesis of YGGFL and PGGFL

A synthesis using a 50 μ m checkerboard mask was conducted. However, P was added to the GGFL sites on the substrate through an additional coupling step. P was added by exposing protected GGFL to light through a mask, and subsequent exposure to P in the manner set forth above. Therefore, half of the regions on the substrate contained YGGFL and the remaining half contained PGGFL.

The fluorescence plot for this experiment showed the regions are again readily discernable between those in which binding did and did not occur. This experiment demonstrated that antibodies are able to recognize a specific sequence and that the recognition is not length-dependent.

12. monomer-by-monomer synthesis
of YGGFL and YPGGFL

In order to further demonstrate the operability of the invention, a 50 μ m checkerboard pattern of alternating YGGFL and YPGGFL was synthesized on a substrate using techniques like those set forth above. The resulting fluorescence plot showed that the antibody was clearly able to recognize the YGGFL sequence and did not bind significantly at the YPGGFL regions.

13. synthesis of an array of sixteen different amino acid sequences and estimation of relative binding affinity to herz antibody

Using techniques similar to those set forth above, an array of 16 different amino acid sequences (replicated four times) was synthesized on each of two glass substrates. The sequences were synthesized by attaching the sequence NVOC-GFL across the entire surface of the slides. Using a series of masks, two layers of amino acids were then selectively applied to the substrate. Each region had dimensions of 0.25 cm x 0.0625 cm. The first slide contained amino acid sequences containing only L- amino acids while the second slide contained selected D- amino acids. Various regions on the first and second slides, were duplicated four times on each slide. The slides were then exposed to the Herz antibody and fluorescein-labeled goat anti-mouse antibodies.

A fluorescence plot of the first slide, which contained only L- amino acids showed red areas (indicating strong binding, i.e., 149,000 counts or more) and black areas (indicating little or no binding of the Herz antibody, i.e., 20,000 counts or less). The sequence YGGFL was clearly most strongly recognized. The sequences YAGFL and YSGFL also exhibited strong recognition of the antibody. By contrast, most of the remaining sequences showed little or no binding. The four duplicate portions of the slide were extremely consistent in the amount of binding shown therein.

A fluorescence plot of the D- amino acid slide indicated that strongest binding was exhibited by the YGGFL sequence. Significant binding was also detected to YaGFL,

YsGFL, and YpGFL. The remaining sequences showed less binding with the antibody. Low binding efficiency of the sequence yGGFL was observed.

5 Table 6 lists the various sequences tested in order of relative fluorescence, which provides information regarding relative binding affinity.

Table 6.
Apparent Binding to Herz Ab

	<u>L- a.a. Set</u>	<u>D- a.a. Set</u>
5	YGGFL	YGGFL
	YAGFL	YaGFL
	YSGFL	YsGFL
	LGGFL	YpGFL
	FGGFL	fGGFL
10	YPGFL	yGGFL
	LAGFL	faGFL
	FAGFL	wGGFL
	WGGFL	yaGFL
		fpGFL
15		waGFL

14. illustrative alternative embodiment

According to an alternative embodiment of the invention, the methods provide for attaching to the surface a caged binding member which, in its caged form, has a relatively low affinity for other potentially binding species, such as receptors and specific binding substances. Such techniques are more fully described in copending application Serial No. 404,920, filed September 8, 1989, and incorporated herein by reference for all purposes. See also U.S.S.N. 07/435,316 (caged biotin parent) and U.S.S.N. 07/612,671 (caged biotin CIP), each of which is hereby incorporated herein by reference.

According to this alternative embodiment, the invention provides methods for forming predefined regions on a surface of a solid support, wherein the predefined regions are capable of immobilizing receptors. The methods make use of caged binding members attached to the surface to enable selective activation of the predefined regions. The caged binding members are liberated to act as binding members ultimately capable of binding receptors upon selective activation of the predefined regions. The activated binding members are then used to immobilize specific molecules such as

receptors on the predefined region of the surface. The above procedure is repeated at the same or different sites on the surface so as to provide a surface prepared with a plurality of regions on the surface containing, for example, the same or different receptors. When receptors immobilized in this way have a differential affinity for one or more ligands, screenings and assays for the ligands can be conducted in the regions of the surface containing the receptors.

The alternative embodiment may make use of novel caged binding members attached to the substrate. Caged (unactivated) members have a relatively low affinity for receptors of substances that specifically bind to uncaged binding members when compared with the corresponding affinities of activated binding members. Thus, the binding members are protected from reaction until a suitable source of energy is applied to the regions of the surface desired to be activated. Upon application of a suitable energy source, the caging groups labilize, thereby presenting the activated binding member. A typical energy source will be light.

Once the binding members on the surface are activated they may be attached to a receptor. The receptor chosen may be a monoclonal antibody, a nucleic acid sequence, a drug receptor, etc. The receptor will usually, though not always, be prepared so as to permit attaching it, directly or indirectly, to a binding member. For example, a specific binding substance having a strong binding affinity for the binding member and a strong affinity for the receptor or a conjugate of the receptor may be used to act as a bridge between binding members and receptors if desired. The method uses a receptor prepared such that the receptor retains its activity toward a particular ligand.

Preferably, the caged binding member attached to the solid substrate will be a photoactivatable biotin complex, i.e., a biotin molecule that has been chemically modified with photoactivatable protecting groups so that it has a significantly reduced binding affinity for avidin or avidin analogs than does natural biotin. In a preferred embodiment, the protecting groups localized in a predefined region of the

surface will be removed upon application of a suitable source of radiation to give binding members, that are biotin or a functionally analogous compound having substantially the same binding affinity for avidin or avidin analogs as does biotin.

5 In another preferred embodiment, avidin or an avidin analog is incubated with activated binding members on the surface until the avidin binds strongly to the binding members. The avidin so immobilized on predefined regions of the surface can then be incubated with a desired receptor or conjugate of a
10 desired receptor. The receptor will preferably be biotinylated, e.g., a biotinylated antibody, when avidin is immobilized on the predefined regions of the surface. Alternatively, a preferred embodiment will present an avidin/biotinylated receptor complex, which has been previously
15 prepared, to activated binding members on the surface.

II. FINGERPRINTING

The above section on generation of reagents for sequencing provides specific reagents useful for fingerprinting
20 applications. Fingerprinting embodiments may be applied towards polynucleotide fingerprinting, polypeptide fingerprinting, cell and tissue classification, cell and tissue temporal development stage classification, diagnostic tests, forensic uses for individual identification, classification of
25 organisms, and genetic screening of individuals. Mapping applications are also described below.

A. Polynucleotide Fingerprint

Polynucleotide fingerprinting may use reagents
30 similar to those described above for probing a sequence for the presence of specific subsequences found therein. Typically, the subsequences used for fingerprinting will be longer than the sequences used in oligonucleotide sequencing. In particular, specific long segments may be used to determine the
35 similarity of different samples of nucleic acids. They may also be used to fingerprint whether specific combinations of information are provided therein. Particular probe sequences are selected and attached in a positional manner to a

the different cell types. Once a cell classification scheme is developed and the structural features which define it are identified using the present invention, homogeneous cell population expressing these features may be separated from others. Standard cell sorters may be coupled with recognition reagents and labels which can distinguish various cell types.

a. T-Cell Classes

T-cell classes are defined on the basis of expression of particular antigens characteristic of each class. For example, mouse T-cell differentiation markers include the LY antigens. With the plurality of different antigens which may be tested using antibody or other recognition reagents, new populations and classes of cells may be defined. For example, different neural cell types may be defined on the basis of cell surface antigens. Different tissue types will be defined on the basis of tissue specific antigens. Developmental cell classes will be similarly defined. All of these screenings can make use of the VLSIPS substrates with specific recognition molecules attached thereto. The substrates are exposed to the cell types directly, assaying for attachment of cells to specific regions, or are exposed to products of a population of cells, e.g., a supernatant, or a cell lysate.

Once a cell classification scheme has been correlated with specific structural markers therein, reagents which recognize those features may be developed and used in a fluorescence activated cell sorter as described, e.g., in Dangl, J. and Herzenberg (1982) J. Immunological Methods 52: 1-14; and Becton Dickinson, Fluorescence Activated Cell Sorters Division, San Jose, California. This will provide a homogeneous population of cells whose function has been defined by structure.

b. B-Cell Classes

The present cell classification scheme may also be used to determine specific B-cell classes. For example, B-cells specific for producing IgM, IgG, IgD, IgE, and IgA may be defined by the internal expression of specific mRNA sequences

encoding each type of immunoglobulin. The classification scheme may depend on either extracellularly expressed markers which are correlated as being diagnostic of specific stages in development, or intracellular mRNA sequences which indicate particular functions.

D. Temporal Development Scheme

1. Developmental Antigens

The present fingerprinting invention also allows cell classification by expression of developmental antigens. For example, a lymphocyte stem cell expresses a particular combination of antigens. As the lymphocyte develops through a program developmental scheme, at various stages it expresses particular antigens which are diagnostic of particular stages in development. Again, the fingerprinting methodology allows for the definition of specific structural features which are diagnostic of developmental or functional features which will allow classification of cells into temporal developmental classes. Cells, products of those cells, or lysates of those cells will be assayed to determine the developmental stage of the source cells. In this manner, once a developmental stage is defined, specific synchronized populations of cells will be selected out of another population. These synchronized populations may be very important in determining the biological mechanisms of development.

2. Developmental mRNA Expression

Besides expressed antigens, the present invention also allows for fingerprinting of the mRNA population of a cell. In this fashion, the mRNA population, which should be a good determinant of developmental stage, will be correlated with other structural features of the cell. In this manner, cells at specific developmental stages will be characterized by the intracellular environment, as well as the extracellular environment. The present invention also allows the combination of definitions based, in part, upon antigens and, in part, upon mRNA expression.

In one embodiment, the two may be combined in a single incubation step. A particular incubation condition may be found which is compatible with both hybridization recognition non-hybridization recognition molecules. Thus, 5 e.g., an incubation condition may be selected which allows both specificity of antibody binding and specificity of nucleic acid hybridization. This allows simultaneous performance of both types of interactions on a single matrix. Again, where developmental mRNA patterns are correlated with structural 10 features, or with probes which are able to hybridize to intracellular mRNA populations, a cell sorter may be used to sort specifically those cells having desired mRNA population patterns.

15 E. Diagnostic Tests

The present invention also provides the ability to perform diagnostic tests. Diagnostic tests typically are based upon a fingerprint type assay, which tests for the presence of specific diagnostic structural features. Thus, the present 20 invention provides means for viral strain identification, bacterial strain identification, and other diagnostic tests using positionally defined specific reagents. The present invention also allows for determining a spectrum of allergies, diagnosing a biological sample for any or all of the above, and 25 testing for many other conditions.

1. Viral Identification

The present invention provides reagents and methodology for identifying viral strains. The specific 30 reagents may be either antibodies or recognition proteins which bind to specific viral epitopes preferably surface exposed, but may make use of internal epitopes, e.g., in a denatured viral sample. In an alternative embodiment, the viral genome may be probed for specific sequences which are characteristic of 35 particular viral strains. As above, a combination of the two may be performed simultaneously in a single interaction step, or in separate tests, e.g., for both genetic characteristics and epitope characteristics.

2. Bacterial Identification

Similar techniques will be applicable to identifying a bacterial source. This may be useful in diagnosing bacterial infections, or in classifying sources of particular bacterial species. For example, the bacterial assay may be useful in determining the natural range of survivability of particular strains of bacteria across regions of the country or in different ecological niches.

3. Other Microbiological Identifications

The present invention provides means for diagnosis of other microbiological and other species, e.g., protozoal species and parasitic species in a biological sample, but also provides the means for assaying a combination of different infections. For example, a biological specimen may be assayed for the presence of any or all of these microbiological species. In human diagnostic uses, typical samples will be blood, sputum, stool, urine, or other samples.

4. Allergy Tests

An immobilized set of antigens may be attached to a solid substrate and, instead of the standard skin reaction tests, a blood sample may be assayed on such a substrate to determine the presence of antibodies, e.g., IgE or other type antibodies, which may be diagnostic of an allergic or immunological susceptibility. A standard radioallergosorbent test (RAST) may be used to check a much larger population of antigens.

In addition, an allergy like test may be used to diagnose the immunological history of a particular individual. For example, by testing the circulating antibodies in a blood sample, which reflects the immunological history and memory of an individual, it may be determined what infections may not have been historically presented to the immune system. In this manner, it may be possible to specifically supplement an immune system for a short period of time with IgG fractions made up of specific types of gamma globulins. Thus, hepatitis gamma

globulin injections may be better designed for a particular environment which a person is expected to be exposed. This also provides the ability to identify genetically equivalent individuals who have immunologically different experiences.

5 Thus, a blood sample from an individual who has a particular combination of circulating antibodies will likely be different from the combination of circulating antibodies found in a genetically similar or identical individual. This could allow for the distinction between clones of particular animals, e.g.,
10 mice, rats, or other animals.

F. Individual Identification

The present invention provides the ability to fingerprint and identify a genetic individual. This individual
15 may be a bacterial or lower microorganism, as described above in diagnostic tests, or of a plant or animal. An individual may be identified genetically or immunologically, as described.

1. Genetic

20 Genetic fingerprinting has been utilized in comparing different related species in Southern hybridization blots. Genetic fingerprinting has also been used in forensic studies, see, e.g., Morris et al. (1989) J. Forensic Science 34: 1311-1317, and references cited therein. As described above, an
25 individual may be identified genetically by a sufficiently large number of probes. The likelihood that another individual would have an identical pattern over a sufficiently large number of probes may be statistically negligible. However, it is often quite important that a large number of probes be used
30 where the statistical probability of matching is desired to be particularly low. In fact, the probes will optimally be selected for having high heterogeneity among the population. In addition, the fingerprint method may make use of the pattern of homologies indicated by a series of more and more stringent
35 washes. Then, each position has both a sequence specificity and a homology measurement, the combination of which greatly increases the number of dimensions and the statistical

likelihood of a perfect pattern match with another genetic individual.

2. Immunological

5 As indicated above in the diagnostic tests, it is possible to identify a particular immune system within a genetically homogeneous class of organisms by virtue of her immunological history. For example, a large colony of cloned mice may be distinguishable by virtue of each immunological
10 history. For example, one mouse may have had an immunological response to exposure to antigen A to which her genetically identical sibling may have not been exposed. By virtue of this differential history, the first of the pair will likely have a high antibody titer against the antigen A whereas her
15 genetically identical sibling will have not had a response to that antigen by virtue of never having been exposed to it. For this reason, immune systems may be identified by their immunological memories. Thus, immunological experience may also be a means for identifying a particular individual at a
20 particular moment in her lifetime.

This same immunological screening may be used for ~~other~~ sorts of identifiable biological products. For example, an individual may be identified by her combination of expressed proteins. These proteins may reflect a physiological state of
25 the individual, and would thus be useful in certain circumstances where diagnostic tests may be performed. For example, an individual may be identified, in part, by the presence of particular metabolic products.

In fact, a plant origin may be determined by virtue
30 of having within its genome an unnatural sequence introduced to it by genetic breeders. Thus, a marker nucleic acid sequence may be introduced as a means to determine whether a genetic strain of a plant or animal originated from another particular source.

35

G. Genetic Screening

1. test alleles with markers

The present invention provides for the ability to screen for genetic variations of individuals. For example, a number of genetic diseases are linked with specific alleles. See, e.g., Scriber, C. et al. (eds.) (1989) The Metabolic Bases of Inherited Disease, McGraw-Hill, New York. In one embodiment, cystic fibrosis has been correlated with a specific gene, see, Gregory et al. (1990) Nature 347: 382-386. A number of alleles are correlated with specific genetic deficiencies. See, e.g., McKusick, V. (1990) Genetic Inheritance in Man: Catalogs of Autosomal Dominant, Autosomal Recessive, and X-linked Phenotypes, Johns Hopkins University Press, Baltimore; Ott, J. (1985) Analysis of Human Genetic Linkage, Johns Hopkins University Press, Baltimore; Track, R. et al. (1989) Banbury Report 32: DNA Technology and Forensic Science, Cold Spring Harbor Press, New York; each of which is hereby incorporated herein by reference.

2. Amniocentesis

Typically, amniocentesis is used to determine whether chromosome translocations have occurred. The mapping procedure may provide the means for determining whether these translocations have occurred, and for detecting particular alleles of various markers.

III. MAPPING

A. Positionally Located Clones

The present invention allows for the positional location of specific clones useful for mapping. For example, caged biotin may be used for specifically positioning a probe to a location on a matrix pattern.

In addition, the specific probes may be positionally directed to specific locations on a substrate by targeting. For example, polypeptide specific recognition reagents may be attached to oligonucleotide sequences which can be complementarily targeted to specific locations on a VLSIPS substrate. Hybridization conditions, as applied for

oligonucleotide probes, will be used to target the reagents to locations on a substrate having complementary oligonucleotides synthesized thereon. In another embodiment, oligonucleotide probes may be attached to specific polypeptide targeting reagents such as an antigen or antibody. These reagents can be directed towards a complementary antigen or antibody already attached to a VLSIPS substrate.

In another embodiment, an unnatural nucleotide which does not interfere with natural nucleotide complementary hybridization may be used to target oligonucleotides to particular positions on a substrate. Unnatural optical isomers of natural nucleotides should be ideal candidates.

In this way, short probes may be used to determine the mapping of long targets or long targets may be used to map the position of shorter probes. See, e.g., Craig et al. 1990 Nuc. Acids Res. 18: 2653-2660.

B. Positionally Defined Clones

Positionally defined clones may be transferred to a new substrate by either physical transfer or by synthetic means. Synthetic means may involve either a production of the probe on the substrate using the VLSIPS synthetic methods, or may involve the attachment of a targeting sequence made by VLSIPS synthetic methods which will target that positionally defined clone to a position on a new substrate. Both methods will provide a substrate having a number of positionally defined probes useful in mapping.

IX. Conclusion

The present inventions provide greatly improved methods and apparatus for synthesis of polymers on substrates. It is to be understood that the above description is intended to be illustrative and not restrictive. Many embodiments will be apparent to those of skill in the art upon reviewing the above description. By way of example, the invention has been described primarily with reference to the use of photoremovable protective groups, but it will be readily recognized by those of skill in the art that sources of radiation other than light

could also be used. For example, in some embodiments it may be desirable to use protective groups which are sensitive to electron beam irradiation, x-ray irradiation, in combination with electron beam lithograph, or x-ray lithography techniques.

5 Alternatively, the group could be removed by exposure to an electric current. The scope of the invention should, therefore, be determined not with reference to the above description, but should instead be determined with reference to the appended claims, along with the full scope of equivalents
10 to which such claims are entitled.

All publications and patent applications referred to herein are incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually incorporated by reference. The
15 present invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the appended claims.

WHAT IS CLAIMED IS:

1. A composition comprising a plurality of positionally distinguishable sequence specific reagents attached to a solid substrate, which reagents are capable of specifically binding to a predetermined subunit sequence of a preselected multi-subunit length having at least three subunits, said reagents representing substantially all possible sequences of said preselected length.
2. A composition of Claim 1, wherein said subunit sequence is a polynucleotide or a polypeptide.
3. A composition of Claim 1, wherein said preselected multi-subunit length is five subunits and said subunit sequence is a polynucleotide sequence.
4. A composition of Claim 1, wherein said specific reagent is an oligonucleotide of at least about five nucleotides.
5. A composition of Claim 1, wherein said specific reagent is a monoclonal antibody.
6. A composition of Claim 1, wherein said specific reagents are all attached to a single solid substrate.
7. A composition of Claim 1, wherein said reagents comprise about 3000 different sequences.

8. A composition of Claim 1, wherein said reagents represents at least about 25% of the possible subsequences of said preselected length.

5 9. A composition of Claim 1, wherein said reagents are localized in regions of the substrate having a density of at least 25 regions per square centimeter.

10 10. A composition of Claim 6, wherein said substrate has a surface area of less than about 4 square centimeters.

11. A method of analyzing a sequence of a polynucleotide or a polypeptide, said method comprising the step of:

15 a) exposing said polynucleotide or polypeptide to a composition of Claim 1.

12. A method of identifying or comparing a target sequence with a reference, said method comprising the step of:

20 a) exposing said target sequence to a composition of Claim 1;

b) determining the pattern of positions of said reagents which specifically interact with said target sequence; and

25 c) comparing said pattern with the pattern exhibited by said reference when exposed to said composition.

13. A method for sequencing a segment of a polynucleotide comprising the steps of:

a) combining:

i) a substrate comprising a plurality of chemically synthesized and positionally distinguishable oligonucleotides capable of recognizing defined oligonucleotide sequences; and

ii) a target polynucleotide; thereby forming high fidelity matched duplex structures of complementary subsequences of known sequence; and

b) determining which of said reagents have specifically interacted with subsequences in said target polynucleotide.

14. A method of Claim 13, wherein said segment is substantially the entire length of said polynucleotide.

15. A method for sequencing a polymer, said method comprising the steps of:

a) preparing a plurality of reagents which each specifically bind to a subsequence of preselected length;

b) positionally attaching each of said reagents to one or more solid phase substrates, thereby producing substrates of

positionally definable sequence specific probes;

- c) combining said substrates with a target polymer whose sequence is to be determined; and
- d) determining which of said reagents have specifically interacted with subsequences in said target polymer.

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10 16. A method of Claim 15, wherein said substrates are beads.

15 17. A method of Claim 15, wherein said plurality of reagents comprise substantially all possible subsequences of said preselected length found in said target.

20 18. A method of Claim 15, wherein said solid phase substrates are a single substrate having attached thereto reagents recognizing substantially all possible subsequences of preselected length found in said target.

25 19. A method of Claim 15, further comprising the step of analyzing a plurality of said recognized subsequences to assemble a sequence of said target polymer.

20. A method of Claim 16, wherein at least some of said plurality of substrates have one subsequence specific reagent attached thereto, and said substrates are coded to indicate the specificity of said reagent.

21. A method of using a fluorescent nucleotide to detect interactions with oligonucleotide probes of known sequence, said method comprising:

- a) attaching said nucleotide to a target
unknown polynucleotide sequence, and
- b) exposing said target polynucleotide sequence to a collection of positionally defined oligonucleotide probes of known sequences to determine the sequences of said probes which interact with said target.

22. A method of Claim 21, further comprising the

step of:

- a) collating said known sequences to determine the overlaps of said known sequences to determine the sequence of said target sequence.

23. A method of mapping a plurality of sequences relative to one another, said method comprising:

- a) preparing a substrate having a plurality of positionally attached sequence specific probes are attached;
- b) exposing each of said sequences to said substrate, thereby determining the patterns of interaction between said sequence specific probes and said sequences; and

- c) determining the relative locations of said sequence specific probe interactions on said sequences to determine the overlaps and order of said sequences.

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24. A method of claim 23, wherein said sequence specific probes are oligonucleotides.

10

25. A method of claim 23, wherein said sequences are nucleic acid sequences.

26. A method of preparing sequences on a substrate comprising the steps of:

15

- a) exposing a first region of said substrate to an activator to remove a protective group;
- b) exposing at least said first region to a first monomer;
- c) exposing a second region to an activator to remove a protective group; and
- d) exposing at least said second region to a second monomer.

20

27. The method as recited in claim 26 wherein said steps of exposing to an activator use an activator selected from the group consisting of ion beams, electron beams, gamma rays, x-rays, ultra-violet radiation, light, infra-red radiation, microwaves, electric currents, radiowaves, and combinations thereof.

25

28. The method as recited in claim 26 wherein said protective groups are photosensitive protective groups.

29. The method as recited in claim 26 wherein said steps of exposing to an activator are steps of applying light to selected regions of said substrate.

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30. The method as recited in claim 26 wherein said first and the second monomers are amino acids.

31. The method as recited in claim 26 further comprising a step of screening sequences on said substrate for affinity with a receptor, said step of screening further comprising the step of exposing said substrate to said receptor and testing for the presence of said receptor in said first and said second region.

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32. The method as recited in claim 31 wherein said receptor is an antibody.

33. The method as recited in claim 26 wherein said substrate is selected from the group consisting of polymerized Langmuir Blodgett film, functionalized glass, germanium, silicon, polymers, (poly)tetrafluoro-ethylene, polystyrene, gallium arsenide, and combinations thereof.

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34. The method as recited in claim 26 wherein said protective group is selected from the group consisting of ortho-nitrobenzyl derivatives, 6-nitroveratryloxy-carbonyl, 2-nitrobenzyloxy-carbonyl, cinnamoyl derivatives, and mixtures thereof.

15

35. The method as recited in claim 26 wherein said first and second regions each have total areas of less than 1 cm².

36. The method as recited in claim 26 wherein said first and second regions each have total areas of between about 1 μm^2 and 10,000 μm^2 .

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37. The method as recited in claim 29 wherein said light is monochromatic coherent light.

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38. The method as recited in claim 26 wherein said steps of exposing to an activator are carried out with a solution in contact with said substrate.

39. The method as recited in claim 38 wherein said solution further comprises said first or said second monomer.

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40. The method as recited in claim 31 wherein said receptor further comprises a marker selected from the group consisting of radioactive markers and fluorescent

markers and wherein said step of testing for the presence of the receptor is a step of detecting said marker.

41. The method as recited in claim 26 wherein the steps of exposing to an
5 activator further comprise steps of:

a) placing a mask adjacent to said substrate, said mask having substantially transparent regions and sub-stantially opaque regions at a wavelength of light; and

b) illuminating said mask with a light source, said light source producing at least said wavelength of light.

10 42. The method as recited in claim 26 wherein said steps are repeated so as to synthesize 10^3 or more different sequences on said substrate.

15 43. The method as recited in claim 26 wherein said steps are repeated so as to synthesize 10^6 or more dif-fer-ent sequences on said substrate.

44. A method of synthesizing a plurality of chemical sequences, said chemical sequences comprising at least a first and a second monomer, comprising the steps of:

20 a) at a first region on a substrate having at least a first and a second region, said first and said second region comprising a substrate protective group, activating said first region to remove said substrate protective group in said first region;

b) exposing said first monomer to said sub-strate, said first monomer further comprising a first monomer protective group, said first monomer binding at said first region;

25 c) activating said second region to remove said substrate protective group in said second region;

d) exposing said second monomer to said sub-strate, said second monomer further comprising a second monomer protective group, said second monomer binding at said second region;

30 e) activating said first region to remove said first monomer protective group;

f) exposing a third monomer to said sub-strate, said third monomer binding at said first region to produce a first sequence;

g) activating said second region to remove said second monomer protective group; and

h) exposing a fourth monomer to said sub-strate, said fourth monomer binding at said second region to produce a second sequence, said second sequence different from said first sequence.

5 45. A method of synthesizing a plurality of chemical sequences, said chemical sequences comprising at least a first and a second monomer, comprising the steps of:

a) on a substrate having at least a first and a second region deactivating said first region to provide a first protective group in said first region;

10 b) exposing said first monomer to said substrate, said first monomer binding at said second region;

c) removing said protective group in said first region;

d) deactivating said second region to provide a second protective group in said second region;

15 e) exposing said second monomer to said substrate, said second monomer binding at said first region;

f) removing said protective group in said second region;

g) deactivating said first region to provide a protective group in said first region;

20 h) exposing a third monomer to said sub-strate, said third monomer binding at said second region to produce a first sequence;

i) removing said protective group in said first region; and

25 j) exposing a fourth monomer to said sub-strate, said fourth monomer binding at said first region to produce a second sequence, said second sequence different than said first sequence.

46. A method of synthesizing at least a first polymer sequence and a second polymer sequence on a substrate, said first polymer sequence having a different monomer sequence from said second polymer sequence, comprising the steps of:

30 a) inserting a first mask between said sub-strate and an energy source, said mask having first regions and second regions, said first regions permitting passage of energy from said source, said second regions blocking energy from said source;

b) directing energy from said source at said substrate, said energy removing a protective group from first portions of said first polymer under said first regions of said first mask;

c) exposing a second portion of said first polymer to said substrate to create a first polymer sequence;

d) inserting a second mask between said sub-strate and said energy source, said second mask having first regions and second regions;

e) directing energy from said source at said substrate, said energy removing said protective group under said first regions of said second mask from first portions of said second polymer; and

f) exposing a second portion of said second polymer to said substrate, said second portion of said second polymer binding with said first portion of said second polymer to create a polymer second sequence.

47. The method as recited in claim 46 wherein said energy is selected from the group consisting of ion beams, electron beams, gamma rays, x-rays, ultra-violet radiation, light, infra-red radiation, microwaves, electric fields, radio-waves, and combinations thereof.

48. The method as recited in claim 44 wherein said protective groups are photosensitive protective groups.

49. The method as recited in claims 44 or 45 wherein said steps of activating and deactivating are steps of applying light to selected regions of said substrate.

50. The method as recited in claims 44 or 45 wherein said first and said second monomers are amino acids.

51. The method as recited in claims 44, 45 or 46 further comprising a step of screening said first and said second sequences for affinity with a first receptor, said step of screening further comprising a step of exposing said substrate to said first receptor and testing for the presence of said first receptor.

52. The method as recited in claim 51 wherein said step of screening is a step of screening with antibodies.

53. The method as recited in claims 44, 45 or 46 wherein said substrate is selected from the group consisting of a polymerized Langmuir Blodgett film, functionalized glass, germanium, silicon, polymers, (poly)tetrafluoro-ethylene, gallium arsenide, gallium phosphide, silicon oxide, silicon nitride and combinations thereof.

54. The method as recited in claim 44 wherein said protective group, said first monomer protective group, and said second monomer protective group are selected from the group consisting of ortho-nitrobenzyl derivatives, 6-nitroveratryloxycarbonyl, 2-nitrobenzyloxy--carbonyl, and mixtures thereof.

55. The method as recited in claim 45 wherein said protective group is a cinnamate group.

56. The method as recited in claims 44 or 45 wherein said first and second regions each have total areas of less than 1 cm^2 .

57. The method as recited in claims 44 or 45 wherein said first and second regions each have total areas of between about $1 \text{ }\mu\text{m}^2$ and $10,000 \text{ }\mu\text{m}^2$.

58. The method as recited in claim 49 wherein said light is monochromatic coherent light.

59. The method as recited in claim 44 wherein said steps of activating are carried out with a solution in contact with said substrate.

60. The method as recited in claim 59 wherein said solution further comprises a monomer.

61. The method as recited in claim 51 wherein said receptor further comprises a marker selected from the group consisting of radioactive markers and fluorescent markers and wherein said step of testing for the presence of the receptor is a step of detecting said marker.

62. The method as recited in claims 44 or 45 wherein two of said first, said second, said third, and said fourth monomers are the same monomers.

5 63. The method as recited in claim 46 wherein the step of inserting a second mask is a step of translating said first mask from a first position to a second position.

64. The method as recited in claim 66 wherein the step of inserting a second mask is a step of rotating said first mask.

10 65. The method as recited in claim 51 further comprising the step of exposing said substrate to a second, labeled receptor, said second, labeled receptor binding at multiple sites on said first receptor.

15 66. The method as recited in claim 65 wherein said first receptor is an antibody of a first animal species and said second receptor is an antibody derived from a second species and directed at said first species.

67. The method as recited in claim 44 wherein:

20 a) said first monomer protective group is removable upon exposure to a first wavelength of light;

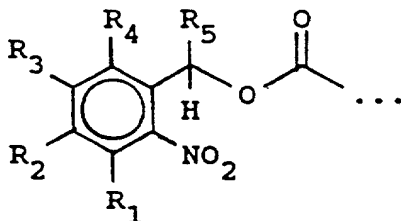
b) said second monomer protective group is removable upon exposure to a second wavelength of light;

25 c) said step of activating said first region to remove said first monomer protective group is a step of exposing substantially all of said substrate to said first wavelength of light; and

d) said step of activating said second region to remove said second monomer protective group is a step of exposing substantially all of said substrate to said second wavelength of light.

30 68. A method as recited in claims 44 or 46 wherein said protective groups are of the form:

5



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where R₁ is alkoxy, alkyl, halo, aryl, alkenyl, or hydrogen; R₂ is alkoxy, alkyl, halo, aryl,
 15 nitro, or hydrogen; R₃ is alkoxy, alkyl, halo, nitro, aryl, or hydrogen; R₄ is alkoxy, alkyl,
 hydrogen, aryl, halo, or nitro; and R₅ is alkyl, alkynyl, cyano, alkoxy, hydrogen, halo, aryl, or
 alkenyl.

69. A method of screening a plurality of amino acid sequences for binding
 20 with a receptor comprising the steps of:

a) on a glass plate having at least a first surface, said at least a first surface
 comprising a photoprotective material selected from the group consisting of nitroveratryloxy
 carbonyl and nitrobenzyloxy carbonyl, reacting said at least a first surface with t-
 butoxycarbonyl for storage, said glass plate substantially transparent to at least ultraviolet
 25 light;

b) exposing said at least a first surface to TFA to remove said t-
 butoxycarbonyl;

c) placing said glass plate on a reactor, said reactor comprising a reactor space,
 said at least a first surface exposed to said reactor space;

30 d) placing a mask at a first position on said glass plate, said mask comprising
 first locations and second locations, said first locations substantially transparent to at least
 ultraviolet light and said second locations substantially opaque to at least ultraviolet light,
 said second locations comprising a light blocking material on a first surface of said mask,
 said first surface of said mask placed in contact with said glass plate;

- e) filling said reactor space with a reaction solution;
- f) illuminating said mask with at least ultraviolet light, said ultraviolet light removing said photoprotective material from said at least a first surface of said glass plate under said first locations of said mask;
- 5 g) exposing said first surface to a first amino acid, said first amino acid binding to regions of said at least a first surface from which said photoprotective material was removed, said first amino acid comprising said photoprotective group at a terminus thereof;
- h) placing a mask in contact with said glass plate at a second position;
- i) illuminating said mask with at least ultraviolet light, said ultraviolet light removing said photoprotective material from said at least a first surface of said glass plate under said first locations of said mask;
- 10 j) exposing said at least a first surface to a second amino acid, said second amino acid binding to regions of said at least a first surface from which said photoprotective material was removed, said second amino acid comprising said photoprotective group at a terminus thereof;
- 15 k) placing a mask in contact with said glass plate at a third position;
- l) illuminating said mask with at least ultraviolet light, said ultraviolet light removing said photoprotective material from said at least a first surface of said glass plate under said first locations of said mask;
- 20 m) exposing said at least a first surface to a third amino acid, said third amino acid binding to regions of said at least a first surface from which said photoprotective material was removed;
- n) placing a mask in contact with said glass plate at a fourth position;
- o) illuminating said mask with at least ultraviolet light, said ultraviolet light removing said photoprotective material from said at least a first surface of said glass plate under said first locations of said mask;
- 25 p) exposing said at least a first surface to a fourth amino acid, said fourth amino acid binding to regions of said at least a first surface from which said photoprotective material was removed, said at least a first surface comprising at least first, second, third, and fourth amino acid sequences;
- 30 q) exposing said at least a first surface to an antibody of interest, said antibody of interest binding more strongly to at least one of said first, said second, said third, or said fourth amino acid sequences;

r) exposing said at least a first surface to a receptor, said receptor recognizing said antibody of interest and binding at multiple locations thereof, said receptor comprising fluorescein;

5 s) exposing said at least a first surface to light, said first surface fluorescing in at least a region where said more strongly bound amino acid sequence is located; and

t) detecting and recording fluoresced light intensity as a function of location across said at least a first surface.

10 70. A method of identifying at least one peptide sequence for binding with a receptor comprising the steps of:

a) on a substrate having a plurality of polypeptides, each having a photoremovable protective group, irradiating first selected polypeptides to remove said protective group;

15 b) contacting said polypeptides with a first amino acid to create a first sequence, second polypeptides on said substrate comprising a second sequence; and

c) identifying which of said first or said second sequence binds with said receptor.

20 71. The method as recited in claim 70 wherein said step of identifying further comprises a step of detecting the presence of a marker selected from the group consisting of radioactive markers and fluorescent markers in said receptor.

25 72. The method as recited in claim 70 wherein said step of irradiating is a step of masking a light source with a mask, said mask comprising first transparent regions and second opaque regions.

73. The method as recited in claim 72 wherein the step of identifying further comprises the steps of:

a) exposing a first receptor to said substrate; and

30 b) exposing a receptor to said first receptor to said substrate, said receptor to said first receptor comprising a marker.

74. The method as recited in claim 73 wherein said marker is selected from the group consisting of radioactive markers and fluorescent markers.

75. The method as recited in claim 73 wherein said first receptor is an antibody from a first species and said receptor to said first receptor is an antibody from a second species directed at said first species.

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76. A method for screening a plurality of polymers for biological activity comprising exposing a receptor to a substrate having said plurality of said polymers on a surface thereof, each of said polymers occupying an area of less than about 1 cm².

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77. A method for screening as recited in claim 73 wherein said area is less than about 0.1 cm².

78. A method as recited in claim 73 wherein said area is less than about 10,000 μm².

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79. A method as recited in claim 73 wherein said area is less than about 100 μm².

80. Apparatus for preparation of a plurality of polymers comprising:

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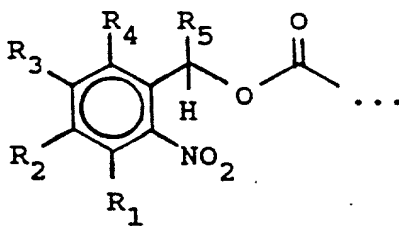
a) a substrate with a surface, said surface comprising a reactive portion, said reactive portion activated upon exposure to an energy source so as to react with a monomer; and

b) means for selectively protecting and exposing portions of said surface from said energy source.

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81. Apparatus as recited in claim 80 wherein said reactive portion further comprises a protective group, said protective group of the form:

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where R₁ is alkoxy, alkyl, halo, aryl, alkenyl, or hydrogen; R₂ is alkoxy, alkyl, halo, aryl, nitro, or hydrogen; R₃ is alkoxy, alkyl, halo, nitro, aryl, or hydrogen; R₄ is alkoxy, alkyl, hydrogen, aryl, halo, or nitro; and R₅ is alkyl, alkynyl, cyano, alkoxy, hydrogen, halo, aryl, or alkenyl.

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82. Apparatus as recited in claim 80 wherein said reactive portion further comprises linker molecules.

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83. Apparatus as recited in claim 82 wherein said linker molecules are selected from the group consisting of ethylene glycol oligomers, diamines, diacids, amino acids, and combinations thereof.

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84. Apparatus as recited in claim 80 wherein said means for selectively protecting further comprises a mask.

85. Apparatus as recited in claim 80 wherein said means for selectively protecting further comprises a light valve.

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86. Apparatus as recited in claim 80 wherein said energy source is a light source.

30

87. Apparatus as recited in claim 80 wherein said reactive portion further comprises a composition selected from the group consisting of nitroveratryloxy carbonyl, nitrobenzyloxy carbonyl, dimethyl-dimethoxybenzyloxy carbonyl, 5-bromo-7-nitroindoliny, hydroxy-2-methyl cinnamoyl, and 2-oxymethylene anthraquinone.

88. Apparatus for preparation of a substrate having a plurality of amino acid sequences thereon, said apparatus comprising:

a) a substrate with a surface;

b) a protective group on said surface, said protective group removable upon exposure to an energy source, said energy source selected from the group consisting of light, electron beams, and x-ray radiation;

c) means for directing said energy source at selected locations on said surface;

5 and

d) means for exposing amino acids to said surface for binding to said surface.

89. Apparatus for screening polymers comprising a substrate with a surface, said surface comprising at least two predefined regions, said predefined regions
10 containing different monomer sequences thereon, said predefined regions each occupying an area of less than about 0.1 cm^2 .

90. Apparatus as recited in claim 89 wherein said area is less than about
15 0.01 cm^2 .

91. Apparatus as recited in claim 89 wherein said area is less than 10000
 μm^2 .

92. Apparatus as recited in claim 89 wherein said area is less than about
20 $100 \mu\text{m}^2$.

93. Apparatus as recited in claims 89, 90, 91, or 92 wherein said monomer sequences are substantially pure within said predefined regions.

25 94. A substrate for screening for biological activity, said substrate comprising 10^3 or more different ligands on a surface thereof in predefined regions.

95. A substrate as recited in claim 94 wherein said substrate comprises 104
or more different ligands in predefined regions.

30

96. A substrate as recited in claim 94 wherein said substrate comprises 105
or more different ligands in predefined regions.

97. A substrate as recited in claim 94 wherein said substrate comprises 106 or more different ligands in predefined regions.

98. A substrate as recited in claims 94, 95, 96, or 97 wherein the ligands are peptides.

99. A substrate as recited in claim 89 wherein said ligands are substantially pure within said predefined regions.

100. Apparatus for screening for biological activity comprising:

a) a substrate comprising a plurality of polymer sequences, said polymer sequences attached to a surface of said substrate at known locations on said substrate, each of said sequences occupying an area of less than about 0.1 cm²;

b) means for exposing said substrate to a receptor, said receptor marked with a fluorescent marker, said receptor binding with at least one of said sequences; and

c) means for detecting a location of said fluorescent marker on said substrate.

101. Apparatus for forming a plurality of polymer sequences comprising:

a) a substrate, said substrate having at least a first surface and a second surface, said second surface comprising a photoremovable protective material, said substrate substantially transparent to at least light of a first wavelength;

b) a reactor body, said reactor body having a mounting surface with a reaction fluid cavity therein, said second surface maintained in a sealed relationship with said mounting surface; and

c) a light source for producing light of at least said first wavelength and directed at a surface of said substrate.

102. Apparatus as recited in claim 101 wherein said light source is directed at said first surface.

103. Apparatus as recited in claim 101 further comprising a mask, said mask placed between said light source and said first surface, said mask having first regions substantially transparent to said first wavelength of light and second regions substantially opaque to said first wavelength of light.

104. Apparatus as recited in claim 101 wherein said cavity comprises a fluid inlet and a fluid outlet, said fluid inlet connected to a pump for flowing reaction fluids through said cavity.

5

105. Apparatus as recited in claim 101 wherein said cavity further comprises a plurality of raised sections.

106. Apparatus as recited in claim 103 wherein said mask further comprises a glass plate.

10

107. Apparatus as recited in claim 106 wherein said opaque regions on said mask comprise chrome.

15

108. Apparatus as recited in claim 101 wherein at least a portion of said second surface comprises a second photoremovable protective group, said second photoremovable protective group activatable upon exposure to light of a second wavelength.

20

109. Apparatus as recited in claim 101 further comprising first and second gaskets on said mounting surface and means for maintaining a vacuum between said first and second gaskets.

25

110. Apparatus as recited in claim 101 wherein said substrate has a thickness of less than 1 mm.

111. Apparatus as recited in claim 101 wherein said substrate has a thickness of less than 0.5 mm.

30

112. Apparatus as recited in claim 101 wherein said substrate has a thickness of less than 0.05 mm.

113. Apparatus as recited in claim 103 wherein said mask is in direct contact with said substrate.

114. Apparatus as recited in claim 113 wherein opaque regions of said mask are placed in direct contact with said substrate.

115. Apparatus as recited in claim 101 further comprising a liquid crystal
5 light valve for selectively controlling exposure of light to said substrate.

116. Apparatus as recited in claim 101 further comprising a fiber optic faceplate between said light source and said substrate.

117. Apparatus as recited in claim 101 further comprising a molecular
10 microcrystal between said light source and said substrate.

118. Apparatus as recited in claim 101 wherein said cavity comprises light
15 absorptive materials.

119. Apparatus as recited in claim 118 wherein said light absorptive material is N,N-diethylamino 2,4-dinitrobenzene.

120. Apparatus as recited in claim 101 wherein said cavity is filled with a
20 carrier solution.

121. Apparatus as recited in claim 120 wherein said carrier material comprises a material selected from the group of 1-hydroxybenzotriazole, dimethylformamide, diisopropylethylamine, and benzotriazolyl-n-oxy-
25 tris(dimethylamino)phosphoniumhexafluorophosphate.

122. Apparatus as recited in claim 101 wherein said substrate is a fiber optic faceplate.

123. Apparatus for detection of fluorescent marked regions on a substrate
30 comprising:

- a) a light source for directing light at a surface of said substrate;
- b) a means for detecting light fluoresced from said surface in response to said light source;

c) means for translating said substrate from a first position to a second position; and

d) means for storing fluoresced light intensity as a function of location on said substrate, said means for storing connected to said means for translating and said means for
5 detecting.

124. Apparatus as recited in claim 123 further comprising video display means for displaying light intensity as a function of location on said substrate.

10 125. Apparatus as recited in claim 123 wherein said means for detecting comprises a photomultiplier tube and a photon counter.

126. Apparatus as recited in claim 124 wherein said means for directing light further comprises a dichroic mirror, said mirror reflecting light at a wavelength of said
15 light source and passing said fluoresced light.

127. Apparatus as recited in claim 125 wherein said light source is a laser light source.

20 128. Apparatus as recited in claim 126 wherein said means for storing is a programmed digital computer.

129. Apparatus as recited in claim 127 further comprising a microscope, said light source directed at said substrate through said microscope, said means for detecting
25 receiving light from said microscope.

130. A method of identifying at least one polymer for binding with a receptor comprising the steps of:

a) on a substrate, said substrate comprising polymers immobilized on a surface
30 of said substrate, said polymers comprising a photoremovable protective group, irradiating a first region of said substrate without irradiating a second region of said substrate to remove said protecting group from said polymers in said first region; and

b) contacting said substrate with a first monomer to couple said monomer to said polymer in said first region, forming a first polymer on said substrate in said first region that is different from said polymer in said second region.

5 131. The method as recited in claim 130 wherein said step of irradiating is a step of masking a light source with a mask, said mask comprising first transparent regions and second opaque regions, said transparent regions transmitting light from said source to said first regions, and said opaque regions blocking light from said source to said second regions.

10 132. The method as recited in claim 130 wherein said first and second regions each have total areas less than about 1 cm².

15 133. The method as recited in claim 130 wherein said steps of irradiating are conducted with a monochromatic light.

20 134. The method as recited in claim 130 wherein said step of irradiating a first region is a step of masking a light source with a mask located in a first position, and wherein said step of irradiating a second region is a step of masking a light source with said mask located in a second position.

 135. The method as recited in claim 130 wherein the step of irradiating further comprises the steps of:

25 a) placing a mask adjacent to said substrate, said mask having substantially transparent regions and substantially opaque regions at a wavelength of light; and

 b) illuminating said mask with a light source, said light source producing at least said wavelength of light.

136. An array of oligonucleotides, the array comprising:
a planar solid support having at least a first surface; and
at least 1000 different oligonucleotides attached to the first surface of the solid
support in an areas of less than 1 cm², wherein each of the different oligonucleotides is
5 attached to the surface of the solid support in a different known location, and has a different
sequence.

137. The array of claim 136, wherein each different oligonucleotides is
from about 4 to about 20 nucleotides in length.

138. The array of claim 136, wherein each different oligonucleotide is at
least 12 nucleotides in length.

139. The array of claim 136, wherein each different oligonucleotide is 2-
15 100 nucleotides in length.

140. The array of claim 136, wherein the array comprises at least 1,000
different oligonucleotides attached to the first surface of the solid support.

141. The array of claim 136, wherein the array comprises at least 10,000
different oligonucleotides attached to the first surface of the solid support.

142. The array of claim 136, wherein each of the different known locations
is physically separated from each other of the known locations.

143. The array of claim 136, wherein said planar solid support is glass.

144. The array of claim 136, wherein said oligonucleotides are attached to
the first surface of the solid support through a linker group.

145. The array of claim 136, wherein the oligonucleotide in the different
known locations are at least 20% pure.

146. The array of claim 136, wherein the oligonucleotides in the different known locations are at least 50% pure.

147. The array of claim 136, wherein the oligonucleotide in the different
5 known locations are at least 80% pure.

148. The array of claim 136, wherein the oligonucleotide in the different known locations are at least 90% pure.

10 149. The array of claim 136, wherein the oligonucleotides in the different known locations are of known sequences.

150. The array of claim 136, wherein said array is produced by a binary synthesis process, said process comprising the steps of:

15 providing a planar solid support, said solid support having a plurality of compounds immobilized on a surface thereof, said compounds having protecting groups coupled thereto;
deprotecting a first portion of said plurality of compounds on said surface and not a second portion of said plurality of compounds;

reacting said first portion of said plurality of compounds with a first component of
20 said oligonucleotide;

deprotecting at least a third portion of said plurality of compounds on said surface, said third portion comprising a fraction of said first portion of said plurality of compounds; reacting said at least third portion of said plurality of compounds with a second component of said oligonucleotide; and

25 optionally repeating said binary synthesis steps to produce said oligonucleotide array.

151. An array of nucleic acids, the array comprising:

a planar support having at least a first surface; and

30 at least 1000 different nucleic acids attached to the first surface of the solid support within an area of 1 cm², wherein each of the different nucleic acids is attached to the surface of the solid support in a different known location, has a different determinable sequence.

152. The array of claim 151, wherein each different nucleic acid is at least 20 nucleotides in length.

153. The array of claim 151, wherein the array comprises at least 1,000 different nucleic acids attached to the first surface of the solid support.

5 154. The array of claim 151, wherein the array comprises at least 10,000 different nucleic acids attached to the first surface of the solid support.

155. The array of claim 151, wherein each of the different known locations is physically separated from each of the other known locations.

10

156. The array of claim 151, wherein said planar solid support is glass.

157. The array of claim 151, wherein said nucleic acids are attached to the first surface of the solid support through a linker group.

15

158. The array of claim 151, wherein the nucleic acids in the different known locations comprise nucleic acids that are at least 20% pure.

159. The array of claim 151, wherein the nucleic acid in the different known locations comprise nucleic acids that are at least 50% pure.

20

160. The array of claim 151, wherein the nucleic acids in the different known locations are at least 80% pure.

25 161. The array of claim 151, the nucleic acids in the different known locations are at least 90% pure.

162. The array of claim 151, wherein said array is produced by a binary synthesis process, said process comprising the steps of:
30 providing a planar, solid support, said solid support having a plurality of compounds immobilized on a surface thereof, said compounds having protecting groups coupled thereto; deprotecting a first portion of said plurality of compounds on said surface and not a second portion of said plurality of compounds;
reacting said first portion of said plurality of compounds with a first reactant;

deprotecting at least a third portion of said plurality of compounds on said surface, said third portion comprising a fraction of said first portion of said plurality of compounds; reacting said at least third portion of said plurality of compounds with a second reactant; and optionally repeating said binary synthesis steps to produce said nucleic acid array--.

5

163. The array of claim 151, wherein the nucleic acids are covalently attached to the support. ,

10 164. An array of nucleic acids, the array comprising:
a planar support having at least a first surface; and
a plurality of different nucleic acids attached to the first surface of the solid support at a density exceeding 10,000 different nucleic acids/cm², wherein each of the different nucleic acids is attached to the surface of the solid support in a different known location, and has a
15 different determinable sequence.

165. An array of nucleic acids, the array comprising:
a planar support having at least a first surface; and
a plurality of different nucleic acids attached to the first surface of the solid support at
20 a density exceeding 400 different nucleic acids/cm², wherein each of the different nucleic acids is attached to the surface of the solid support in a different known location, has a different determinable sequence, wherein the surface and the support are made from different materials.

25 166. The array of claim 151, wherein the different known locations are square in shape.

167. The array of claim 151, wherein the substrate is glass.

30 168. The array of claim 151, wherein the substrate is silicon dioxide.

169. The array of claim 151, wherein the substrate is (poly)tetrafluoroethylene, (poly)vinylidenedifluoride, polystyrene or polycarbonate.

170. The method of claim 151, wherein the substrate is optically transparent.

171. The array of claim 151, wherein the substrate is functionalized with groups that attach to the plurality of different nucleic acids.

5

231

	after 1990-1993	after 1994-1997	after 1998-2000	after 2001-2003	after 2004-2006	after 2007-2009	after 2010-2012	after 2013-2015	after 2016-2018	after 2019-2021	after 2022-2024	after 2025-2027	after 2028-2030	after 2031-2033	after 2034-2036	after 2037-2039	after 2040-2042	after 2043-2045	after 2046-2048	after 2049-2051	after 2052-2054	after 2055-2057	after 2058-2060	after 2061-2063	after 2064-2066	after 2067-2069	after 2070-2072	after 2073-2075	after 2076-2078	after 2079-2081	after 2082-2084	after 2085-2087	after 2088-2090	after 2091-2093	after 2094-2096	after 2097-2099	after 2100-2102	after 2103-2105	after 2106-2108	after 2109-2111	after 2112-2114	after 2115-2117	after 2118-2120	after 2121-2123	after 2124-2126	after 2127-2129	after 2130-2132	after 2133-2135	after 2136-2138	after 2139-2141	after 2142-2144	after 2145-2147	after 2148-2150	after 2151-2153	after 2154-2156	after 2157-2159	after 2160-2162	after 2163-2165	after 2166-2168	after 2169-2171	after 2172-2174	after 2175-2177	after 2178-2180	after 2181-2183	after 2184-2186	after 2187-2189	after 2190-2192	after 2193-2195	after 2196-2198	after 2199-2201	after 2202-2204	after 2205-2207	after 2208-2210	after 2211-2213	after 2214-2216	after 2217-2219	after 2220-2222	after 2223-2225	after 2226-2228	after 2229-2231	after 2232-2234	after 2235-2237	after 2238-2240	after 2241-2243	after 2244-2246	after 2247-2249	after 2250-2252	after 2253-2255	after 2256-2258	after 2259-2261	after 2262-2264	after 2265-2267	after 2268-2270	after 2271-2273	after 2274-2276	after 2277-2279	after 2280-2282	after 2283-2285	after 2286-2288	after 2289-2291	after 2292-2294	after 2295-2297	after 2298-2300	after 2301-2303	after 2304-2306	after 2307-2309	after 2310-2312	after 2313-2315	after 2316-2318	after 2319-2321	after 2322-2324	after 2325-2327	after 2328-2330	after 2331-2333	after 2334-2336	after 2337-2339	after 2340-2342	after 2343-2345	after 2346-2348	after 2349-2351	after 2352-2354	after 2355-2357	after 2358-2360	after 2361-2363	after 2364-2366	after 2367-2369	after 2370-2372	after 2373-2375	after 2376-2378	after 2379-2381	after 2382-2384	after 2385-2387	after 2388-2390	after 2391-2393	after 2394-2396	after 2397-2399	after 2400-2402	after 2403-2405	after 2406-2408	after 2409-2411	after 2412-2414	after 2415-2417	after 2418-2420	after 2421-2423	after 2424-2426	after 2427-2429	after 2430-2432	after 2433-2435	after 2436-2438	after 2439-2441	after 2442-2444	after 2445-2447	after 2448-2450	after 2451-2453	after 2454-2456	after 2457-2459	after 2460-2462	after 2463-2465	after 2466-2468	after 2469-2471	after 2472-2474	after 2475-2477	after 2478-2480	after 2481-2483	after 2484-2486	after 2487-2489	after 2490-2492	after 2493-2495	after 2496-2498	after 2499-2501	after 2502-2504	after 2505-2507	after 2508-2510	after 2511-2513	after 2514-2516	after 2517-2519	after 2520-2522	after 2523-2525	after 2526-2528	after 2529-2531	after 2532-2534	after 2535-2537	after 2538-2540	after 2541-2543	after 2544-2546	after 2547-2549	after 2550-2552	after 2553-2555	after 2556-2558	after 2559-2561	after 2562-2564	after 2565-2567	after 2568-2570	after 2571-2573	after 2574-2576	after 2577-2579	after 2580-2582	after 2583-2585	after 2586-2588	after 2589-2591	after 2592-2594	after 2595-2597	after 2598-2600	after 2601-2603	after 2604-2606	after 2607-2609	after 2610-2612	after 2613-2615	after 2616-2618	after 2619-2621	after 2622-2624	after 2625-2627	after 2628-2630	after 2631-2633	after 2634-2636	after 2637-2639	after 2640-2642	after 2643-2645	after 2646-2648	after 2649-2651	after 2652-2654	after 2655-2657	after 2658-2660	after 2661-2663	after 2664-2666	after 2667-2669	after 2670-2672	after 2673-2675	after 2676-2678	after 2679-2681	after 2682-2684	after 2685-2687	after 2688-2690	after 2691-2693	after 2694-2696	after 2697-2699	after 2700-2702	after 2703-2705	after 2706-2708	after 2709-2711	after 2712-2714	after 2715-2717	after 2718-2720	after 2721-2723	after 2724-2726	after 2727-2729	after 2730-2732	after 2733-2735	after 2736-2738	after 2739-2741	after 2742-2744	after 2745-2747	after 2748-2750	after 2751-2753	after 2754-2756
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5

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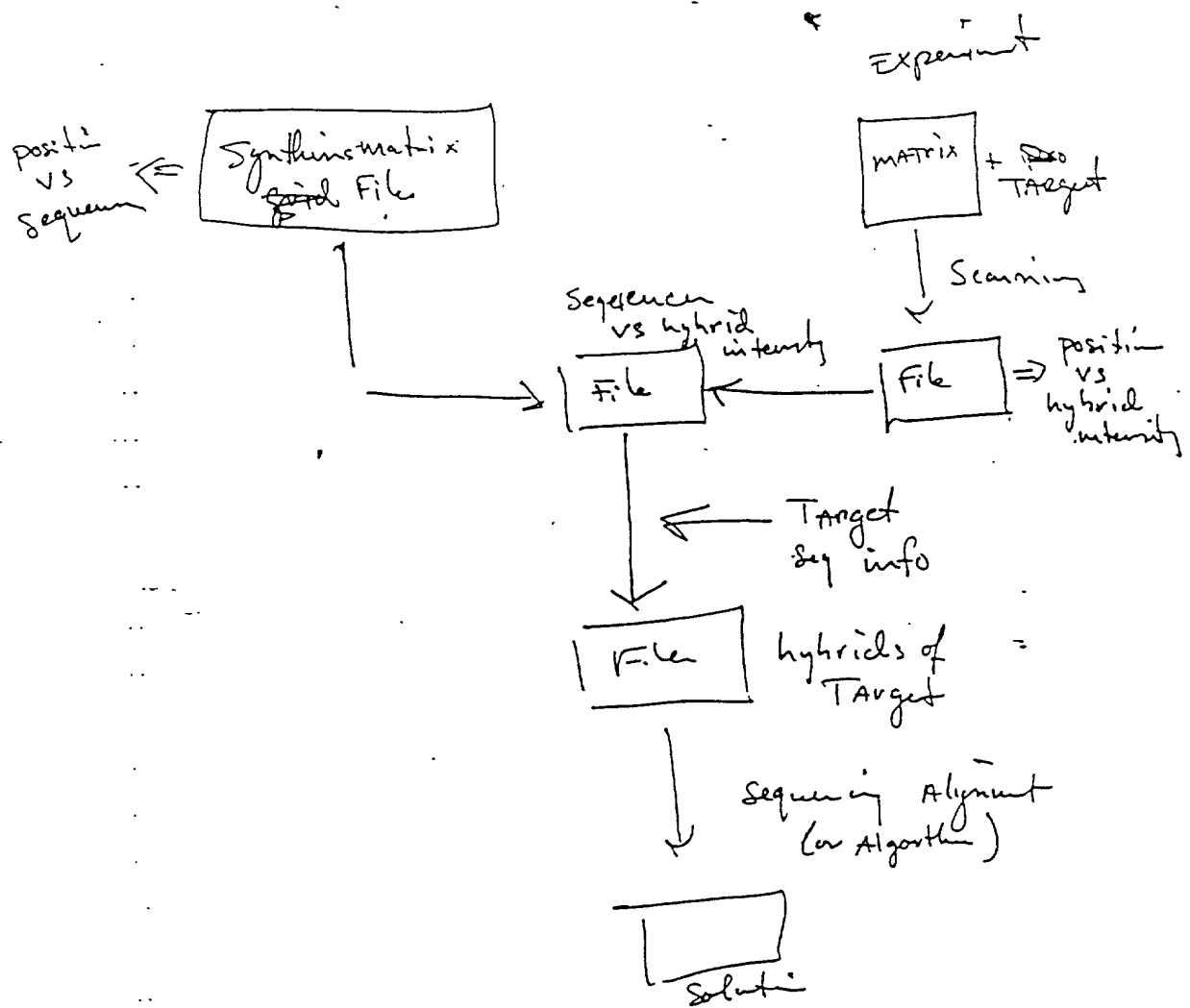


Figure 1

007000-81019900

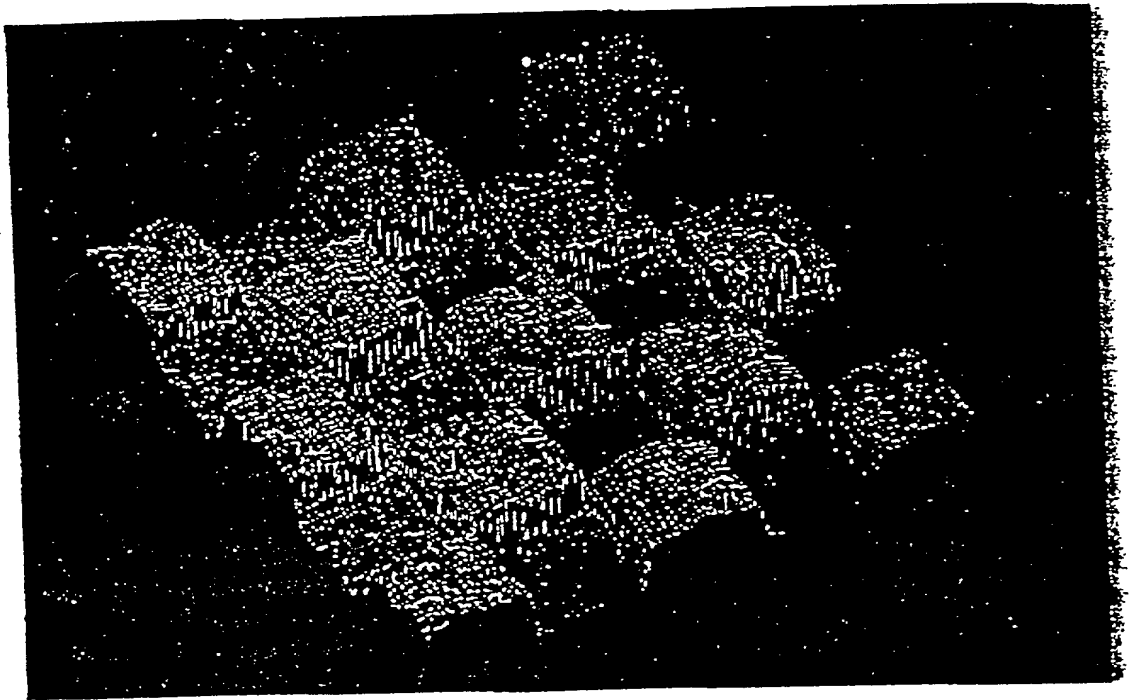


Figure 2

007000" 81019900

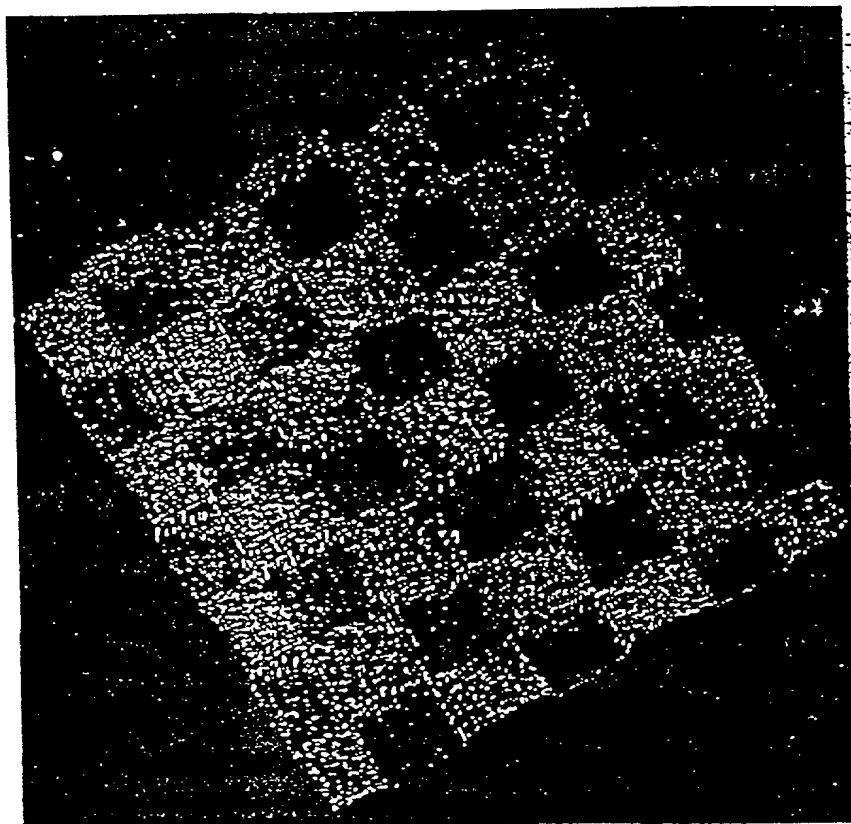


Figure 3

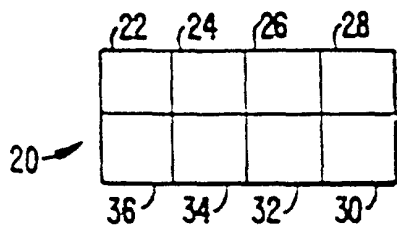


FIG. 4 A.

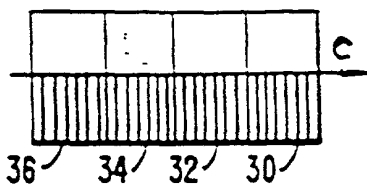


FIG. 4 B.

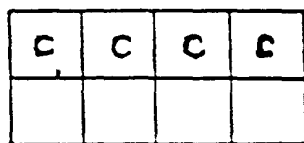


FIG. 4 C.

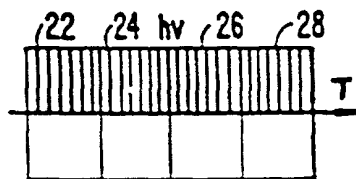


FIG. 4 D.

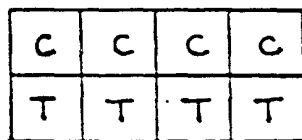


FIG. 4 E.

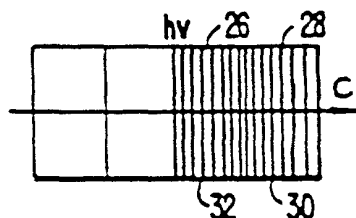


FIG. 4 F.

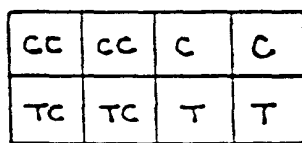


FIG. 4 G.

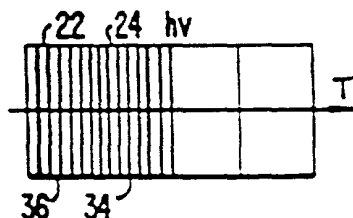


FIG. 4 H.

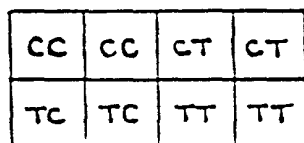


FIG. 4 I.

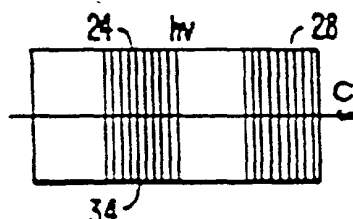


FIG. 4 J.

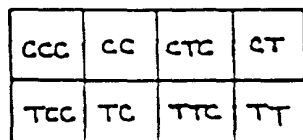


FIG. 4 K.

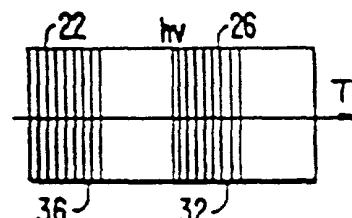


FIG. 4 L.

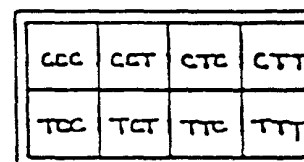


FIG. 4 M.

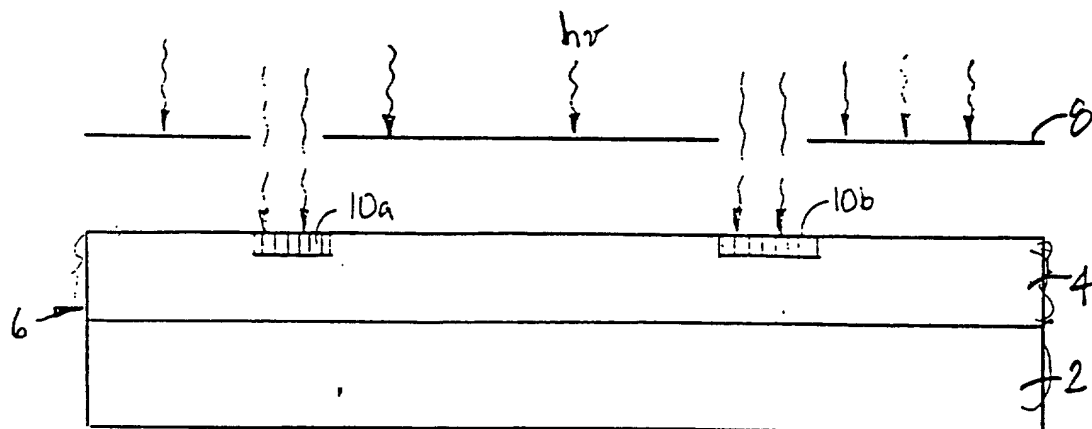


FIG. 5

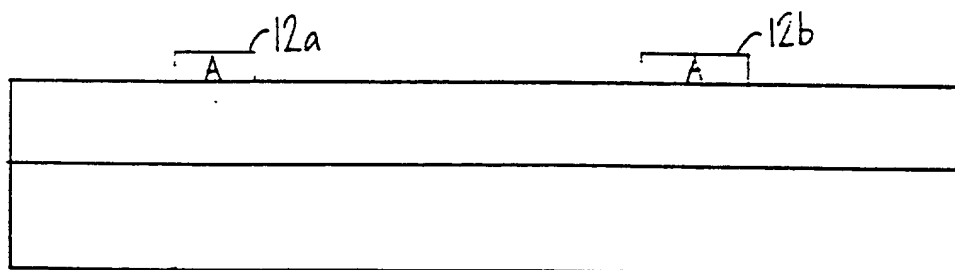


FIG. 6

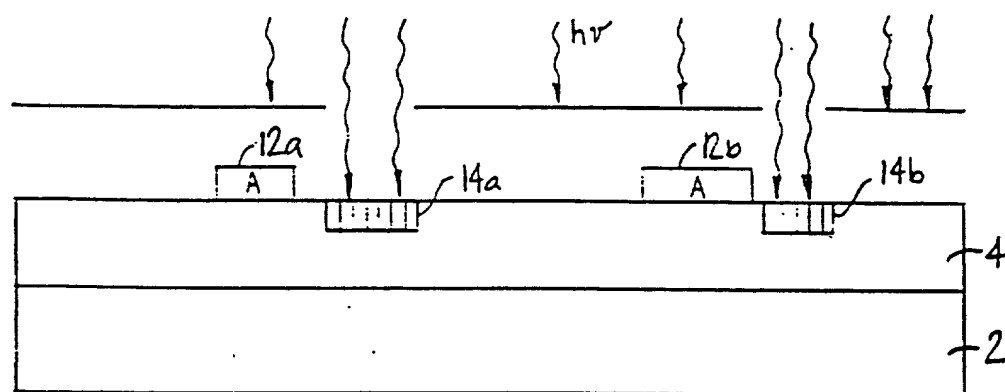


FIG. 7

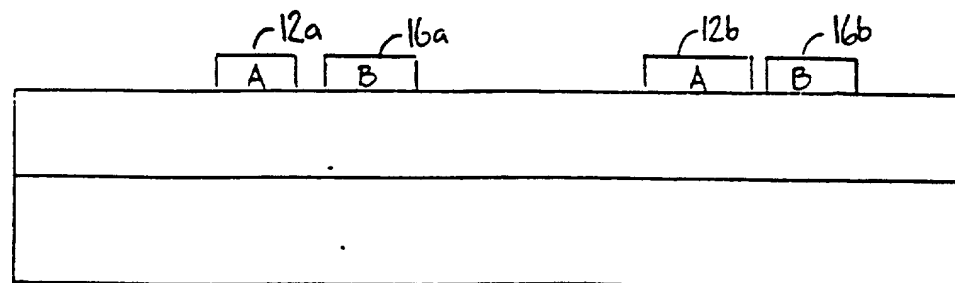


FIG. 8

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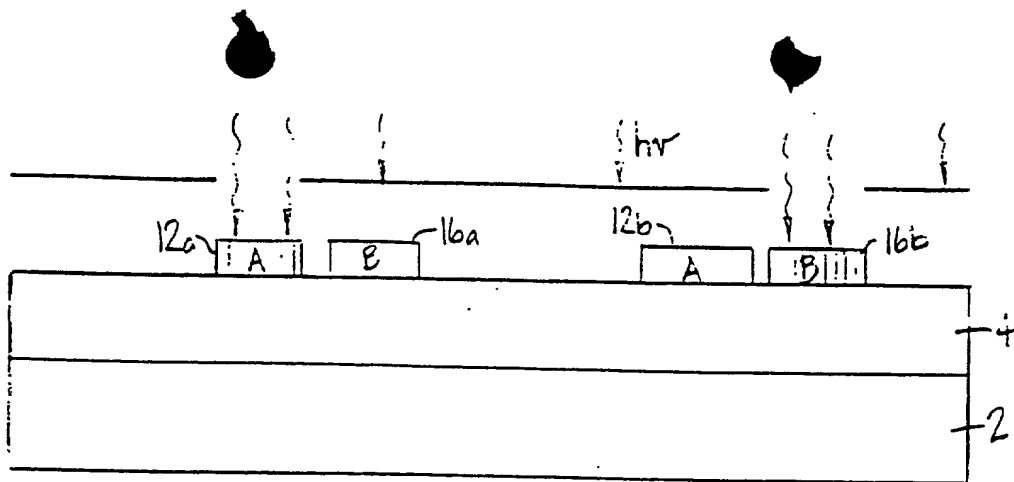


FIG. 9

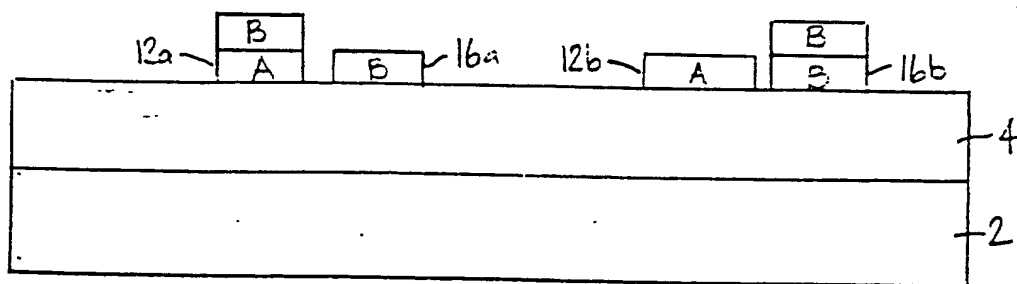


FIG. 10

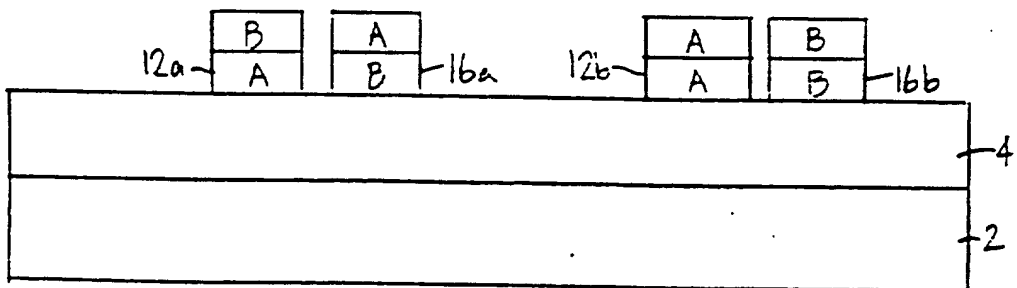


FIG. 11

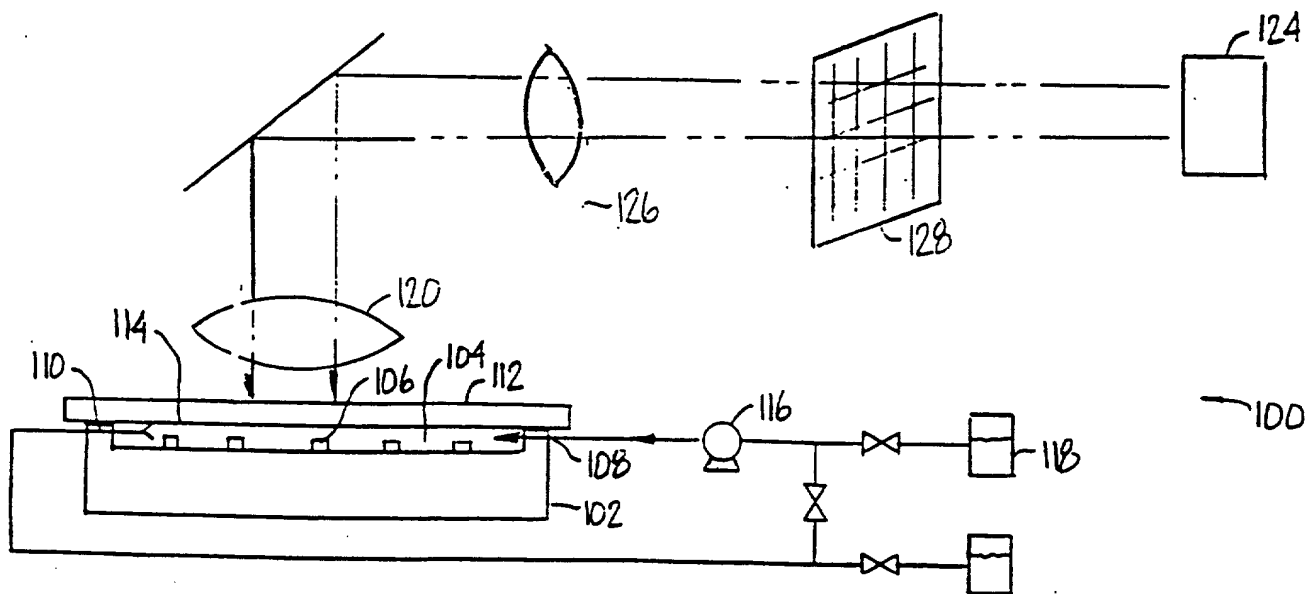


FIG. 12 A

007060-81619960

007000-8409960

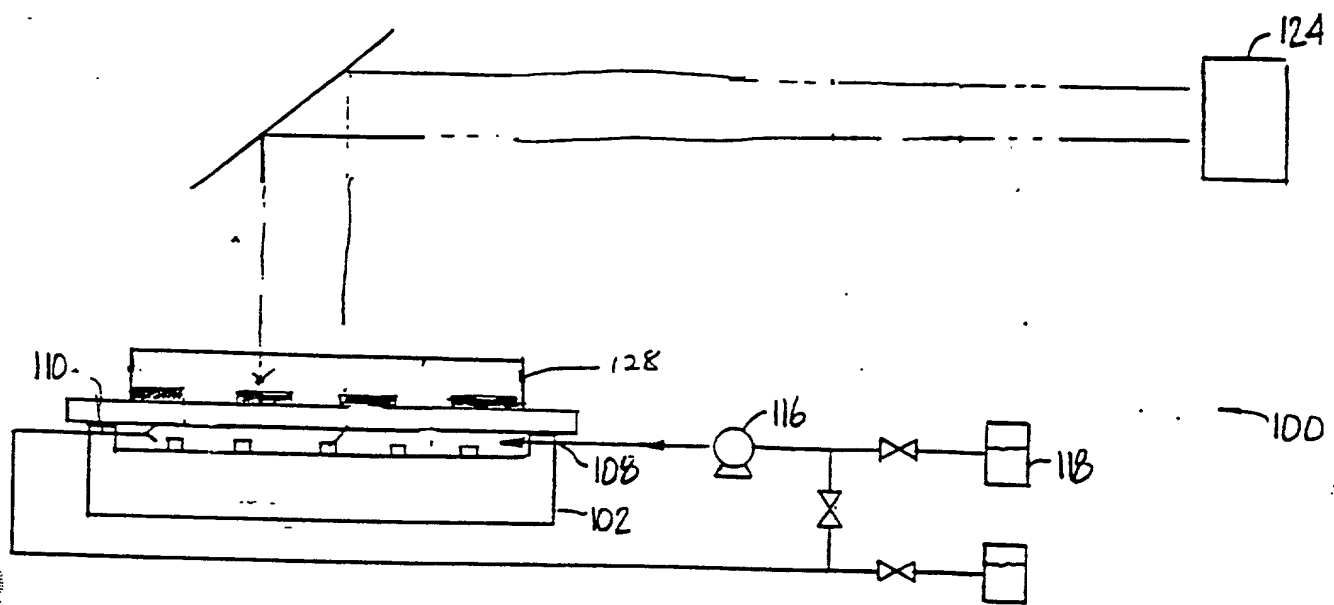


FIG. 12 B

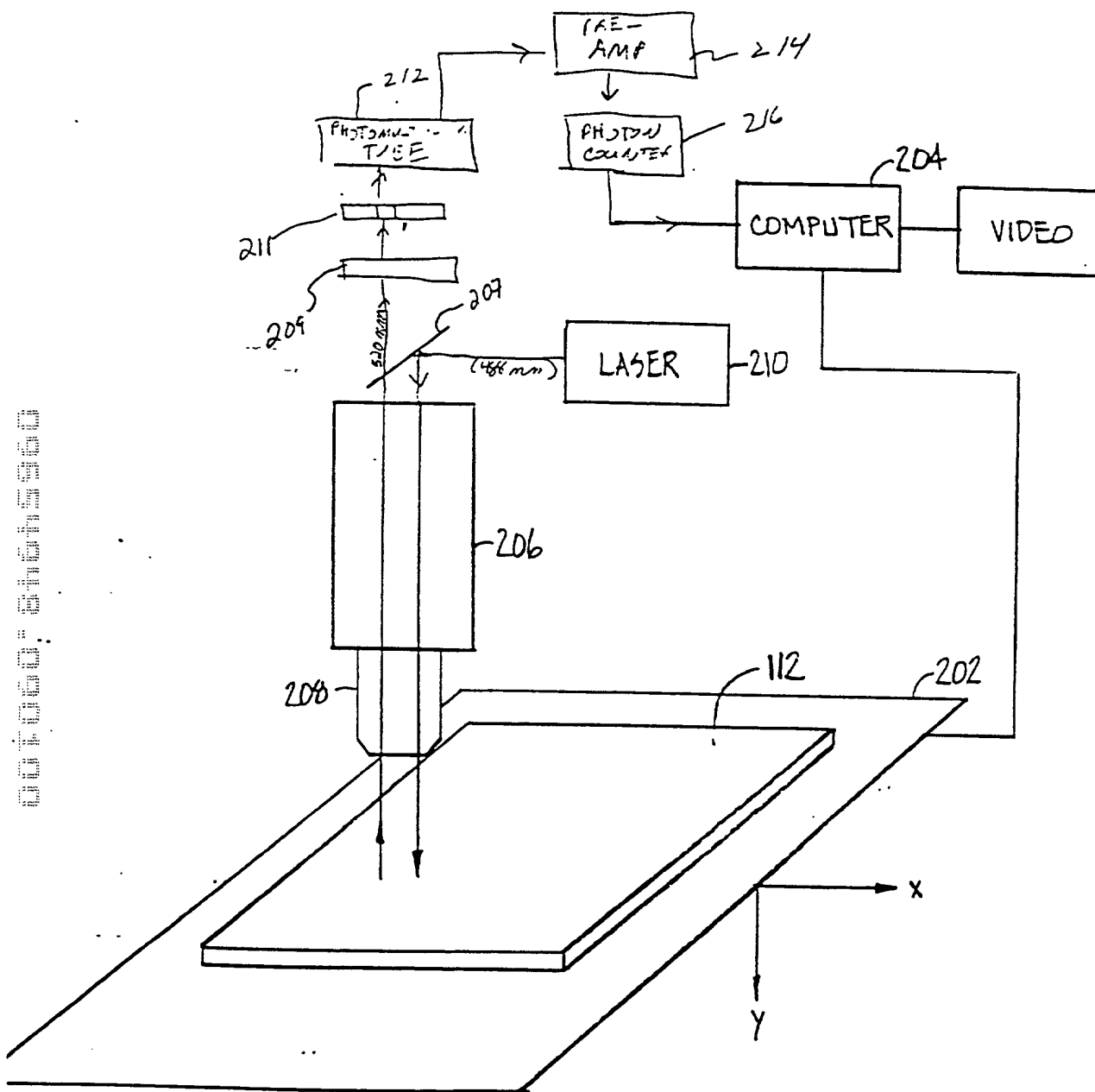


FIG. 13

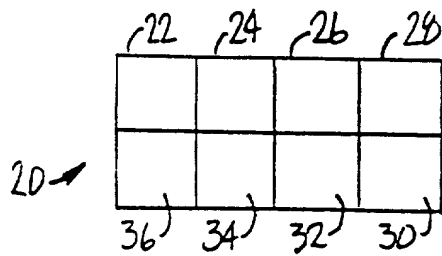


FIG. 14 A

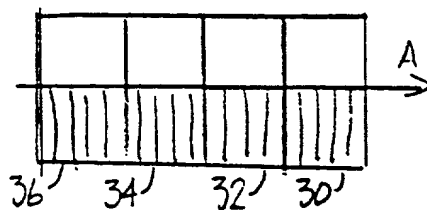


FIG. 14 B

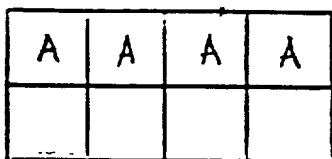


FIG. 14 C

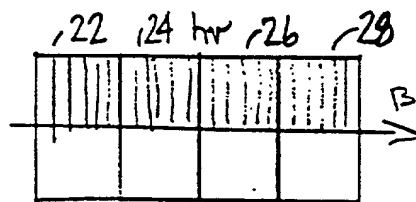


FIG. 14 D

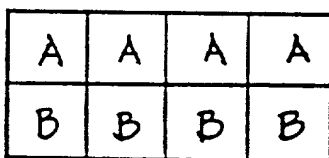


FIG. 14 E

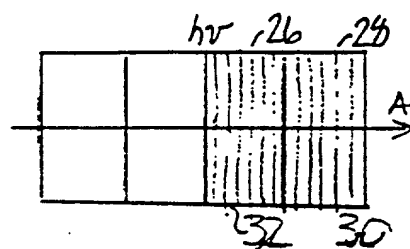


FIG. 14 F

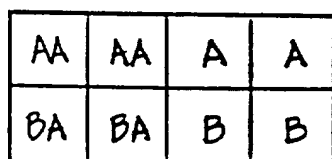


FIG. 14 G

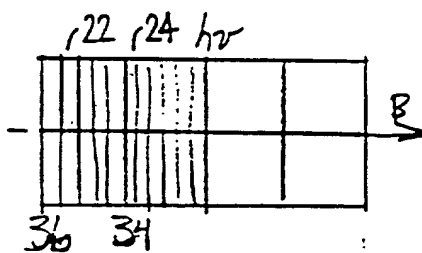


FIG. 14 H

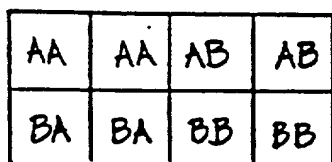


FIG. 14 I

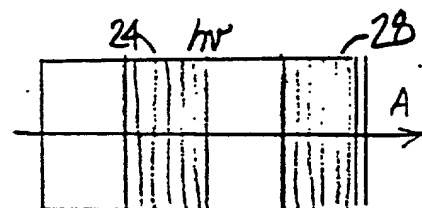
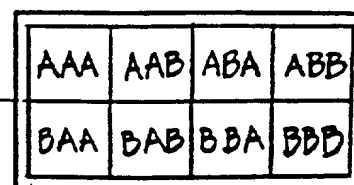
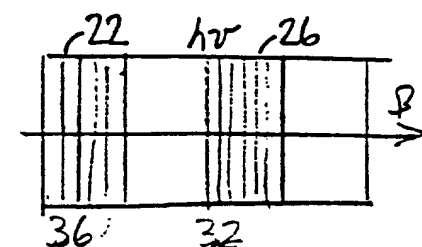
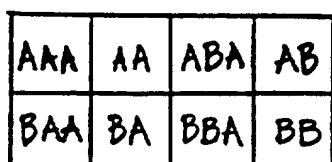


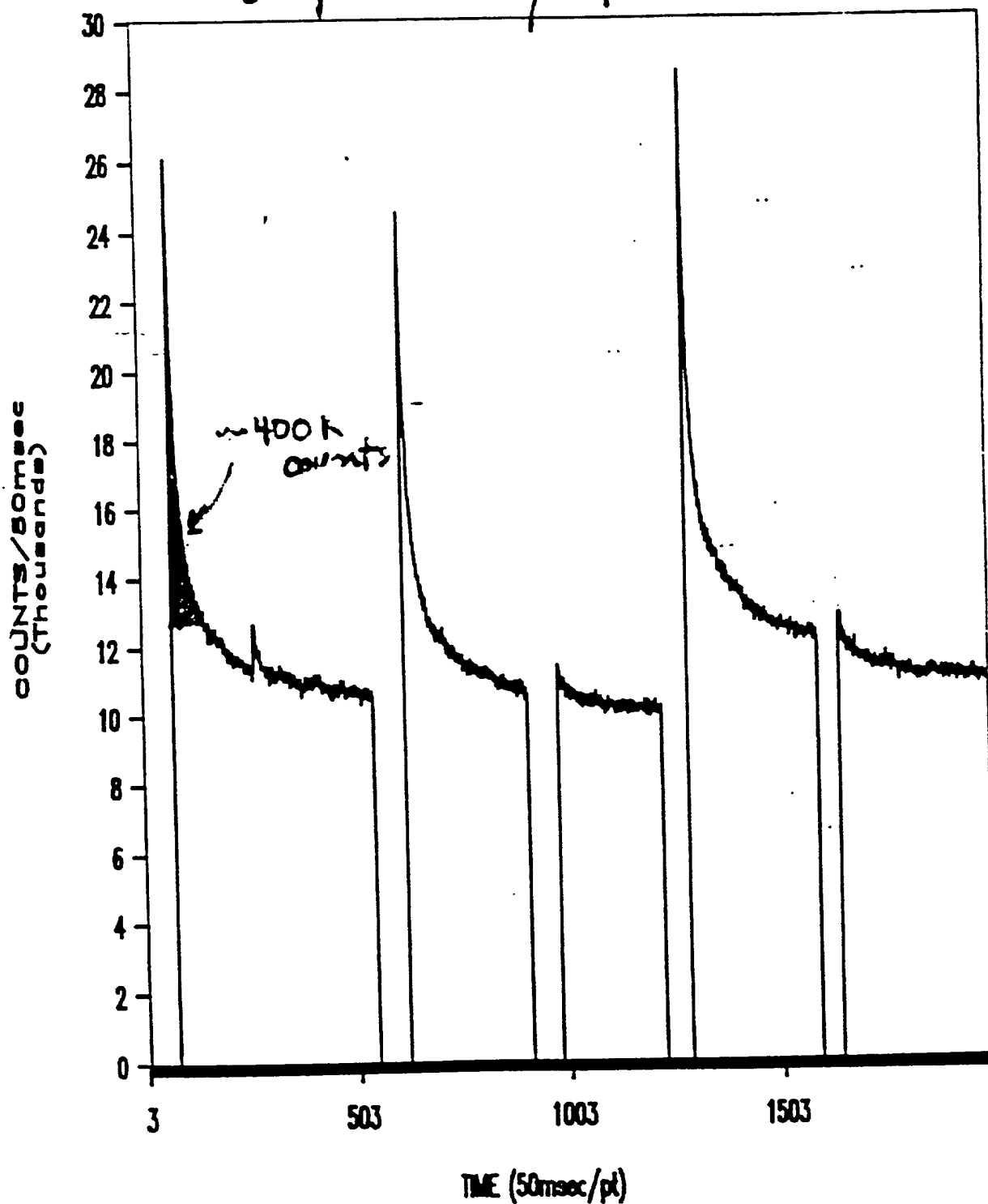
FIG. 14 J



00000-81019900

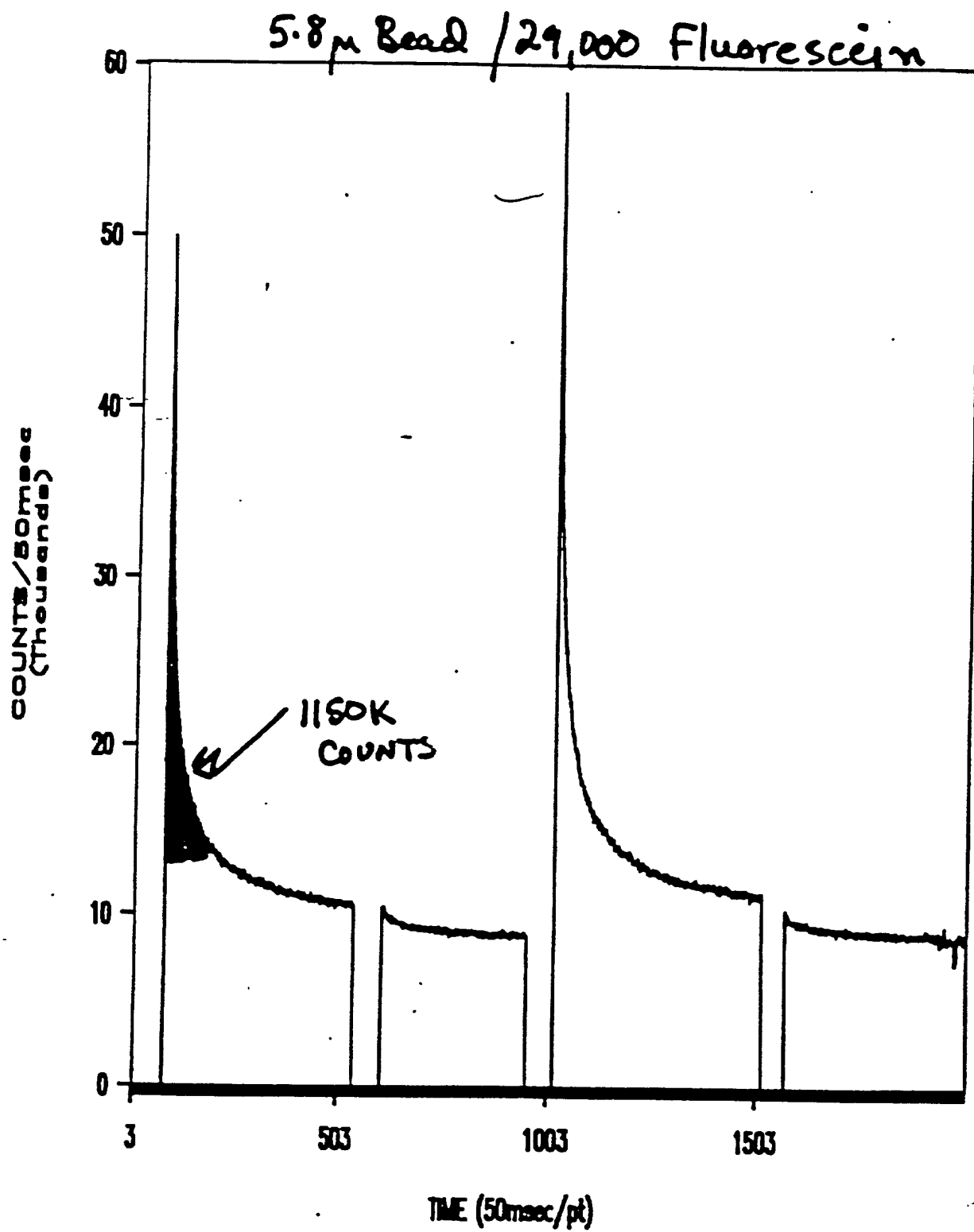
FIG. 15 A

5.9 μ beads / 7,000 Fluorescein



$\sim 2 \times 10^{-6}$ chromophore / A^2

FIG. 15 B



2.8×10^{-6} chromophore / \AA^2

007060-81019500

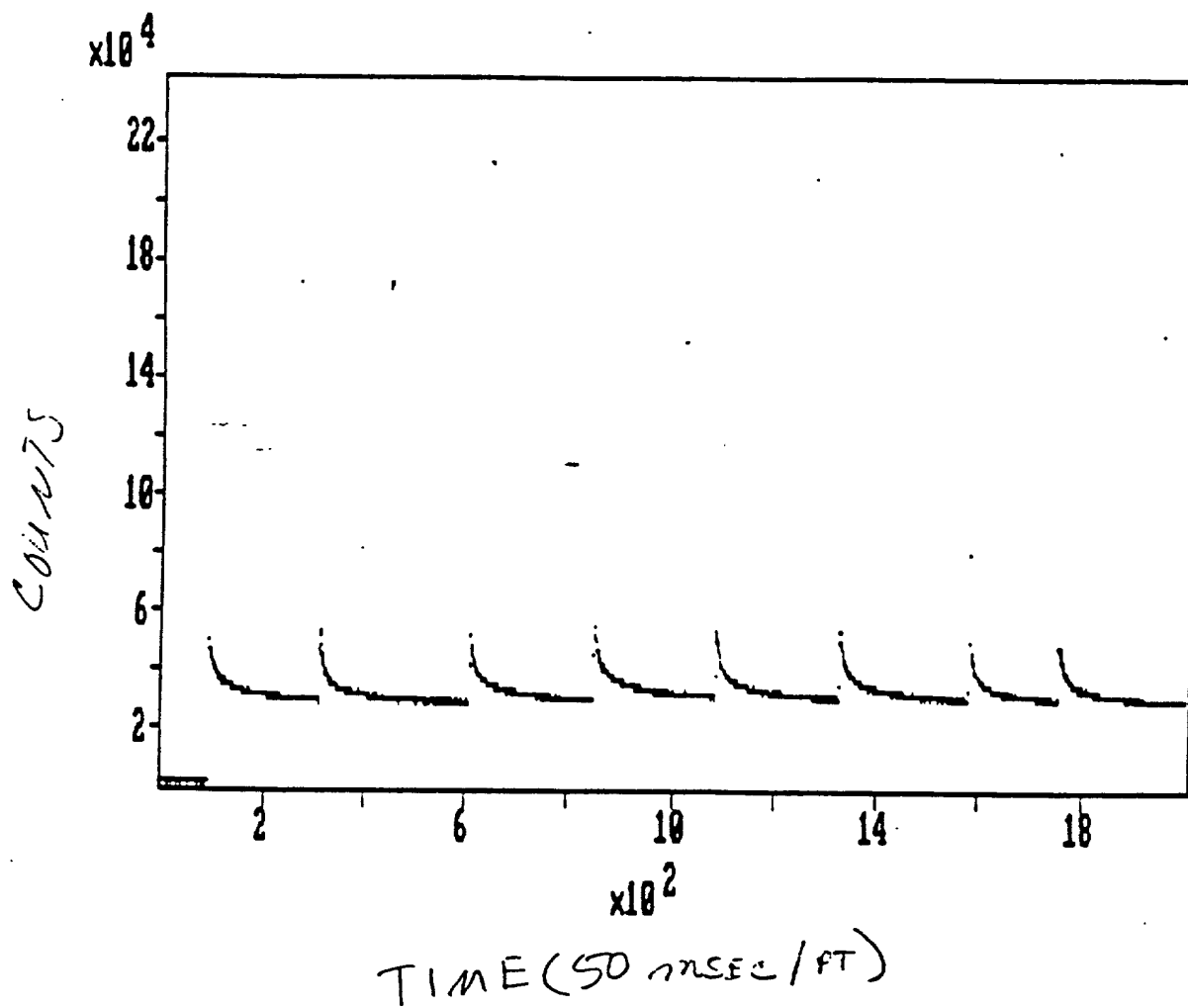
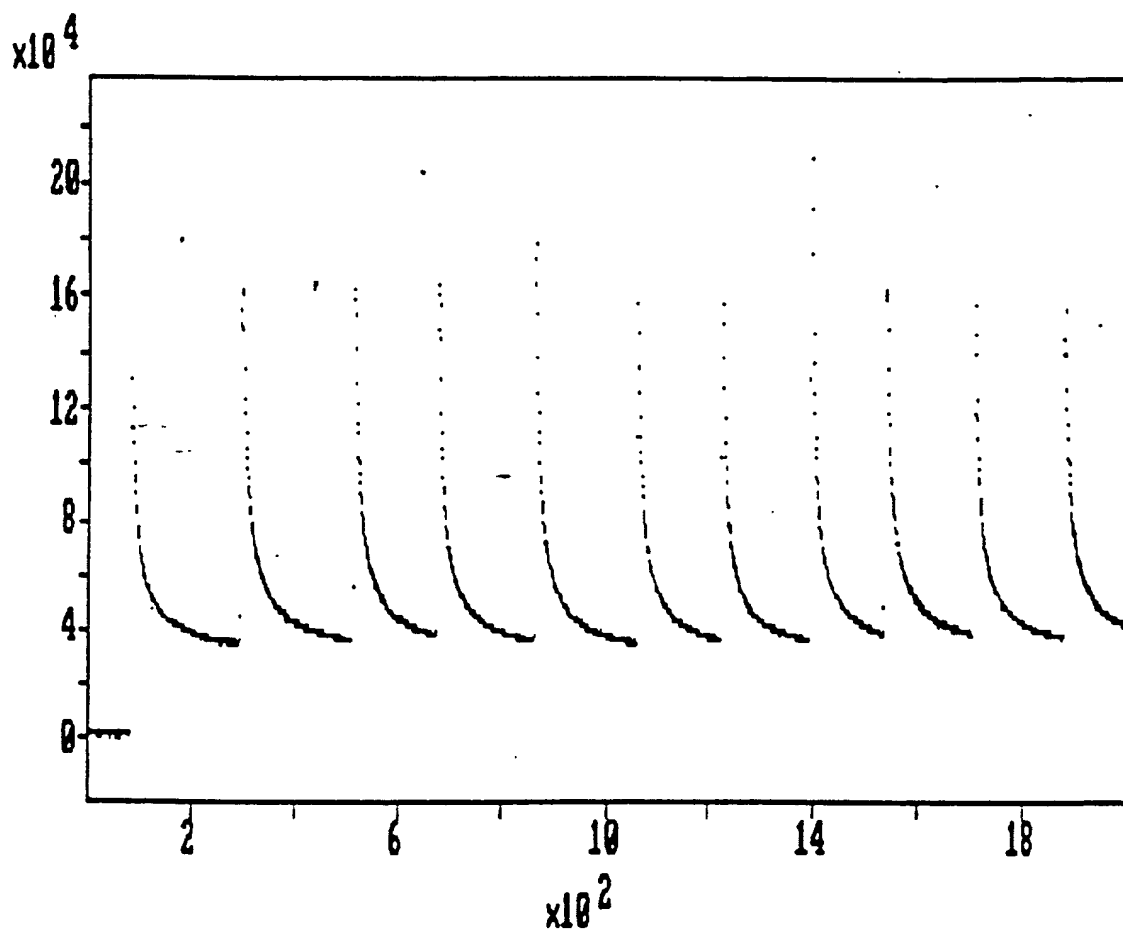


FIG. 16 A

21/10/00



TIME (50 msec/pt)

FIG. 16 B

FIG. 17 A

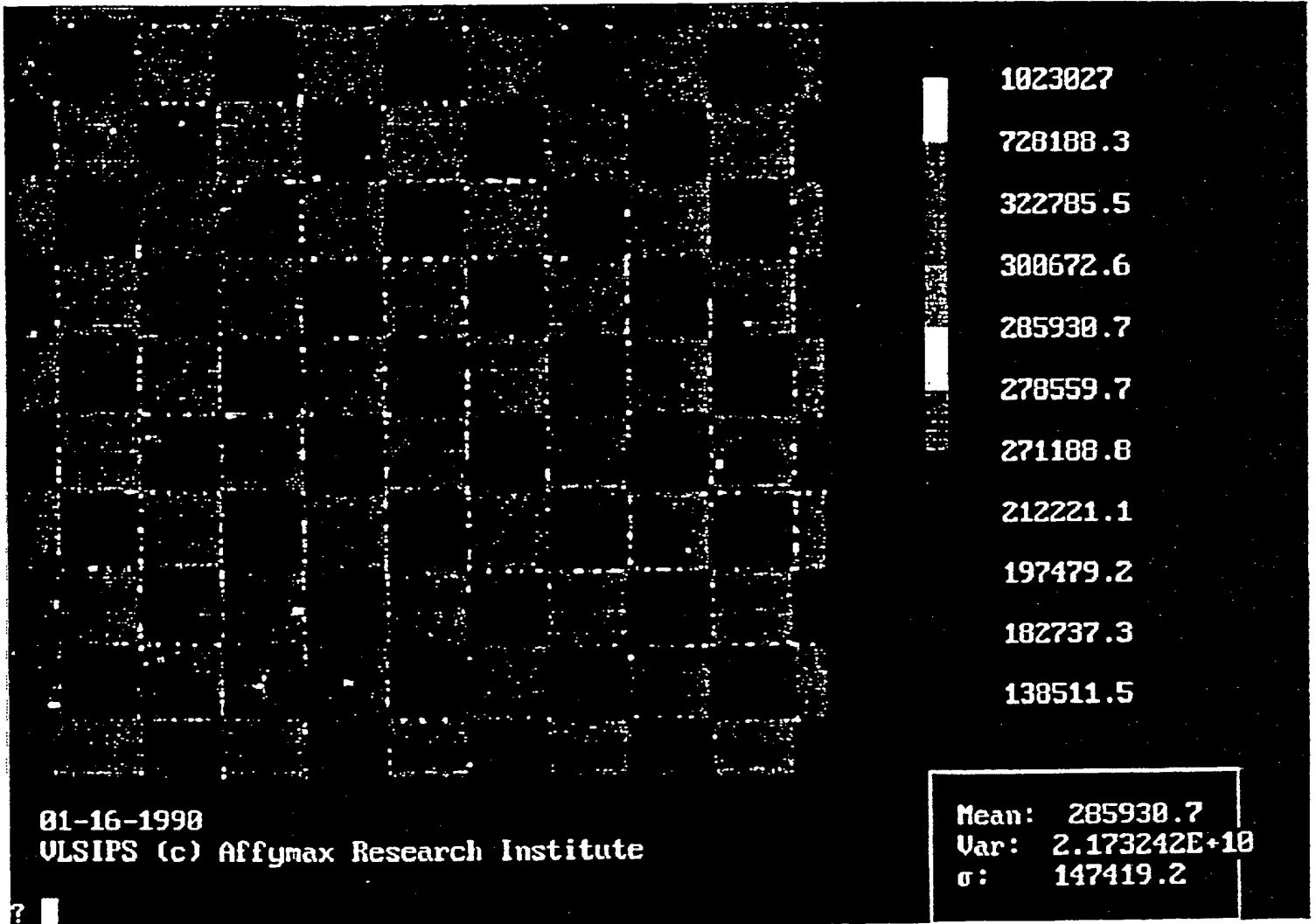


FIG. 17 B

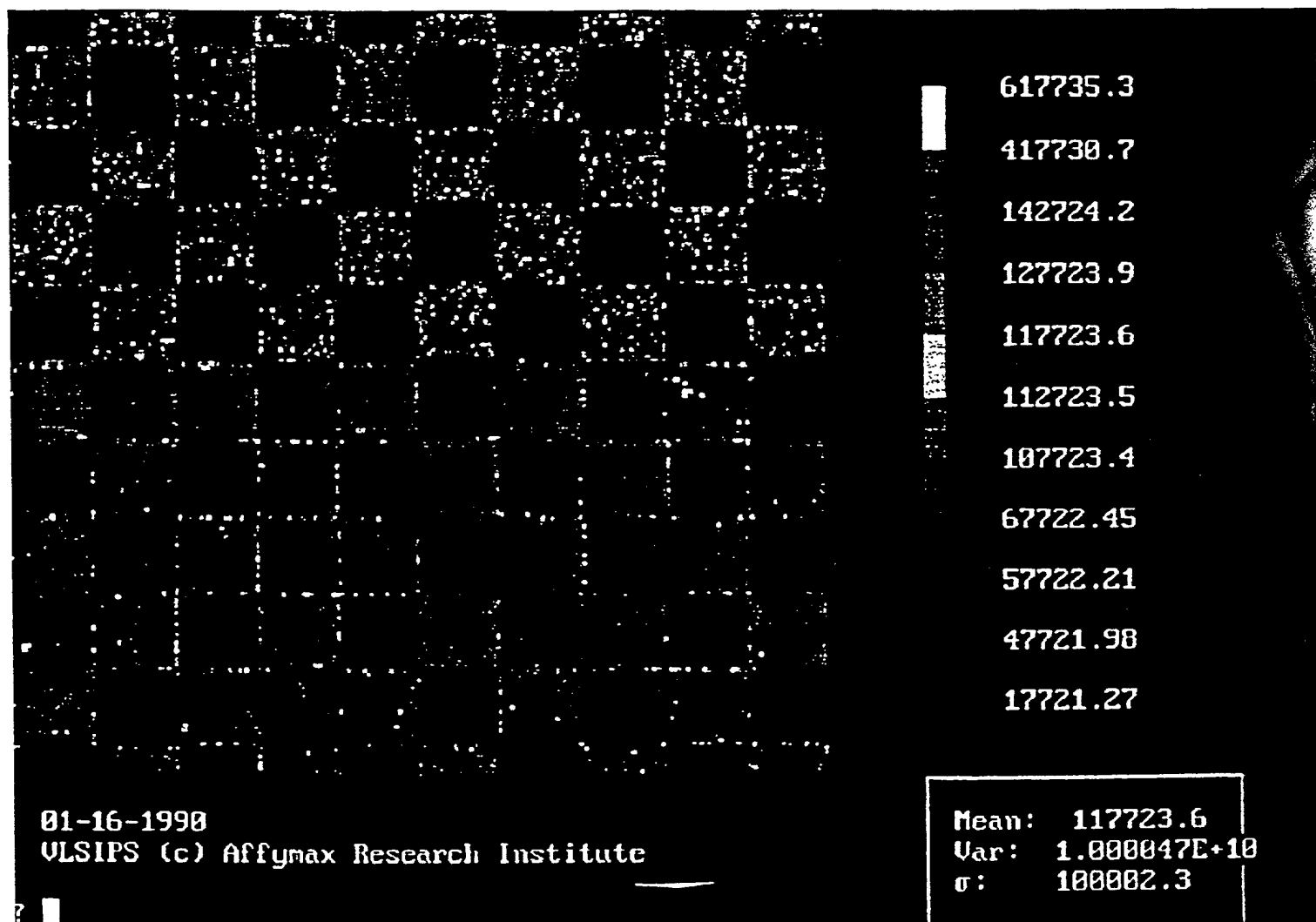


FIG. 17 C

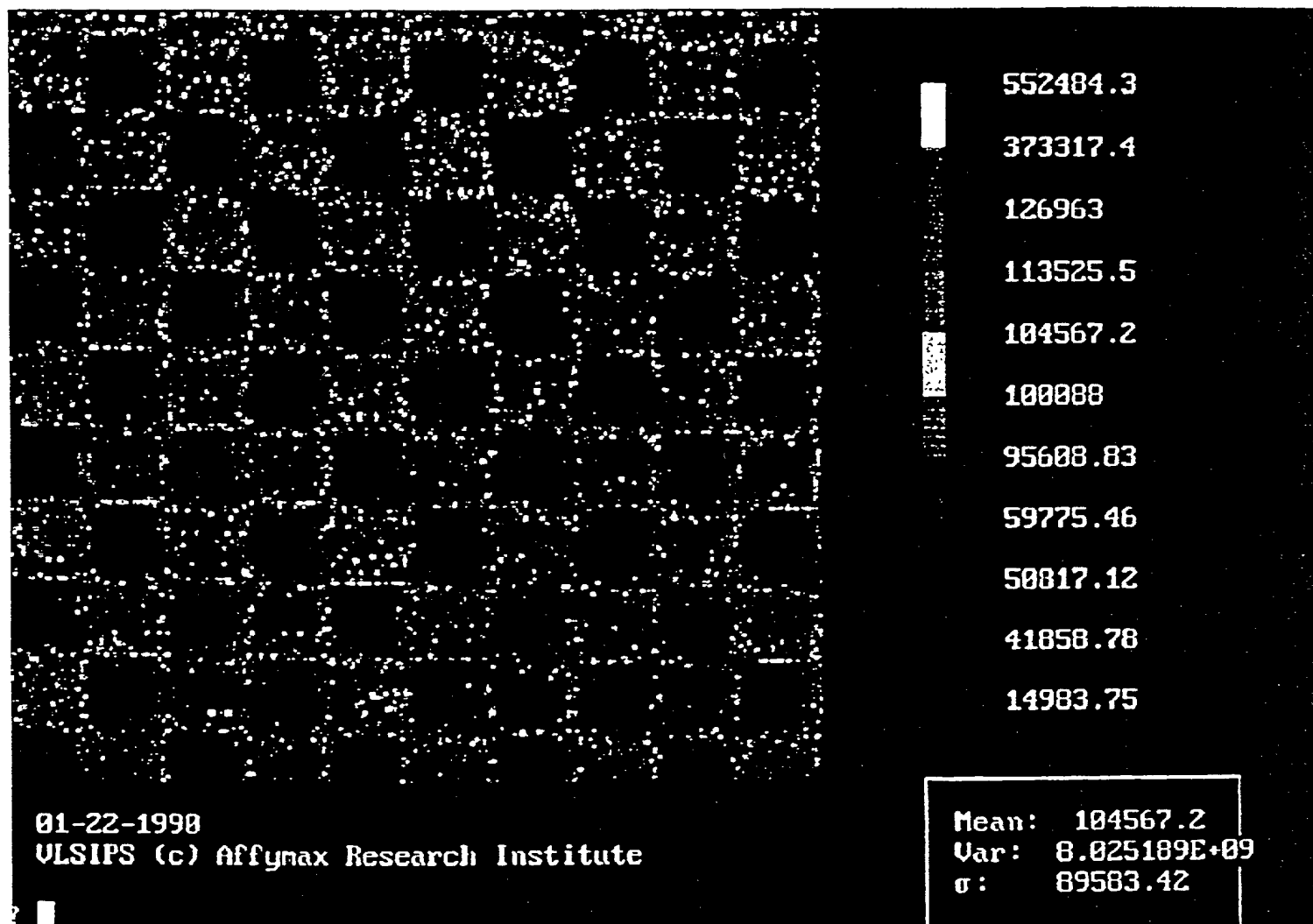


FIG. 17 D

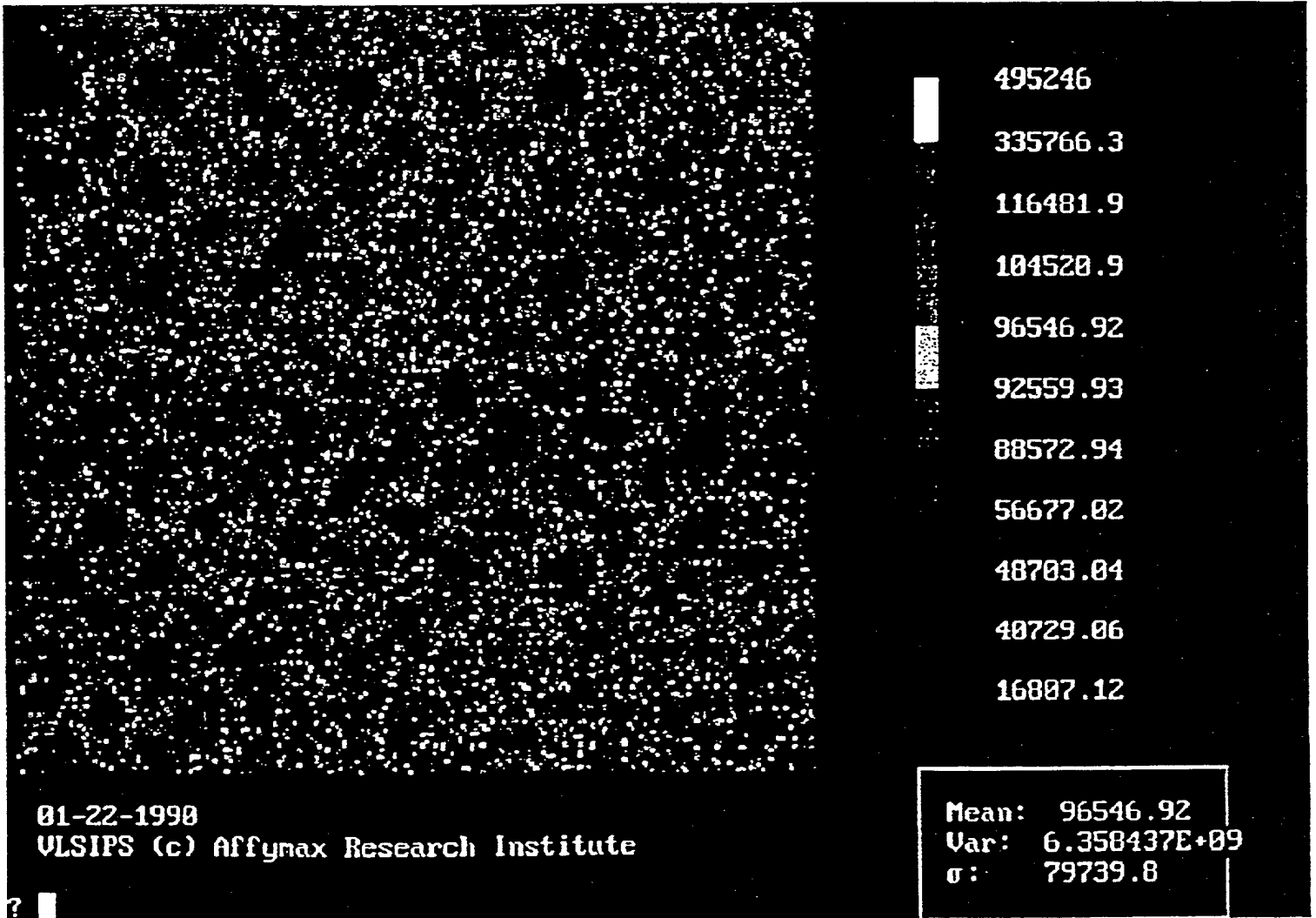


FIG. 18

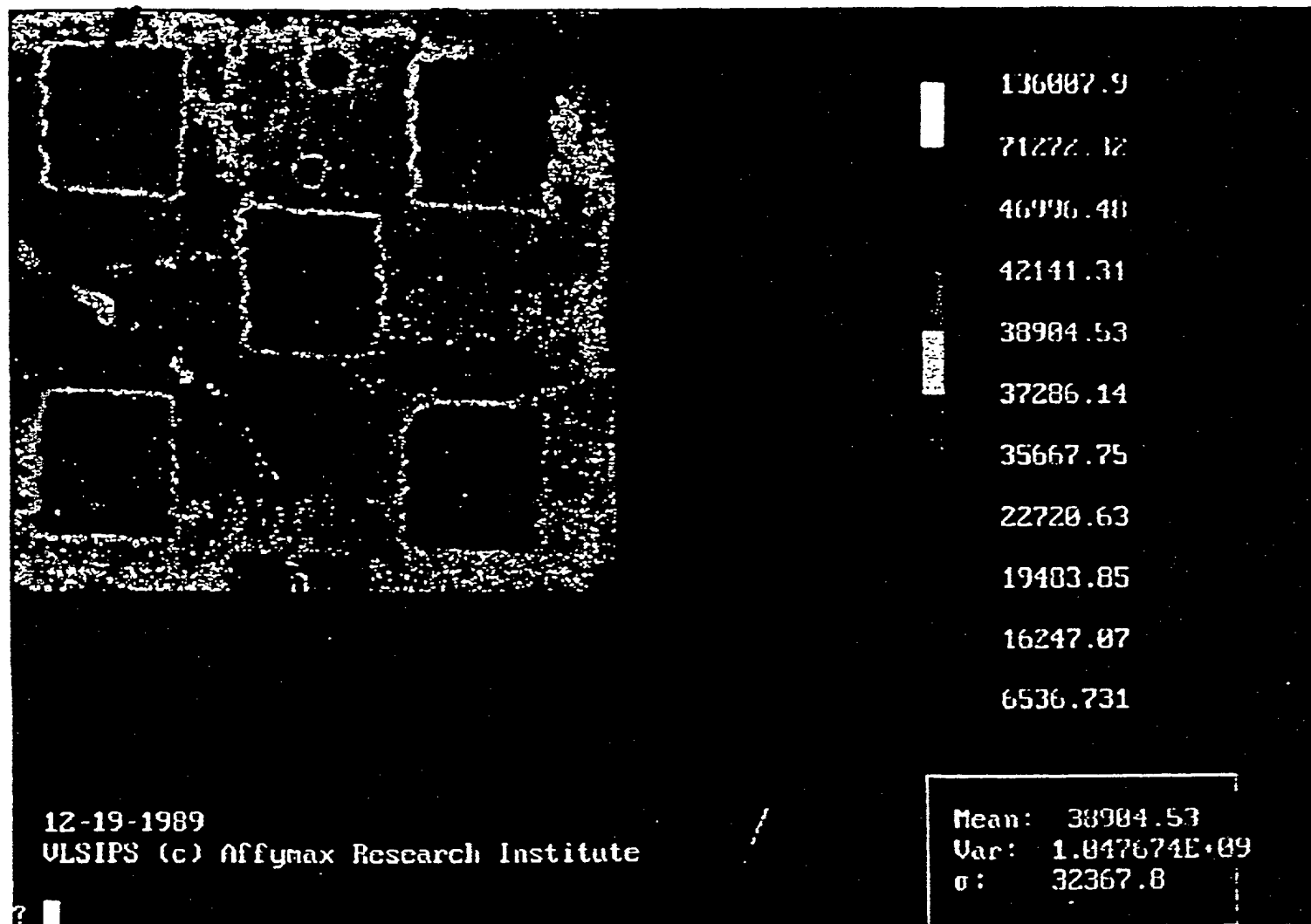
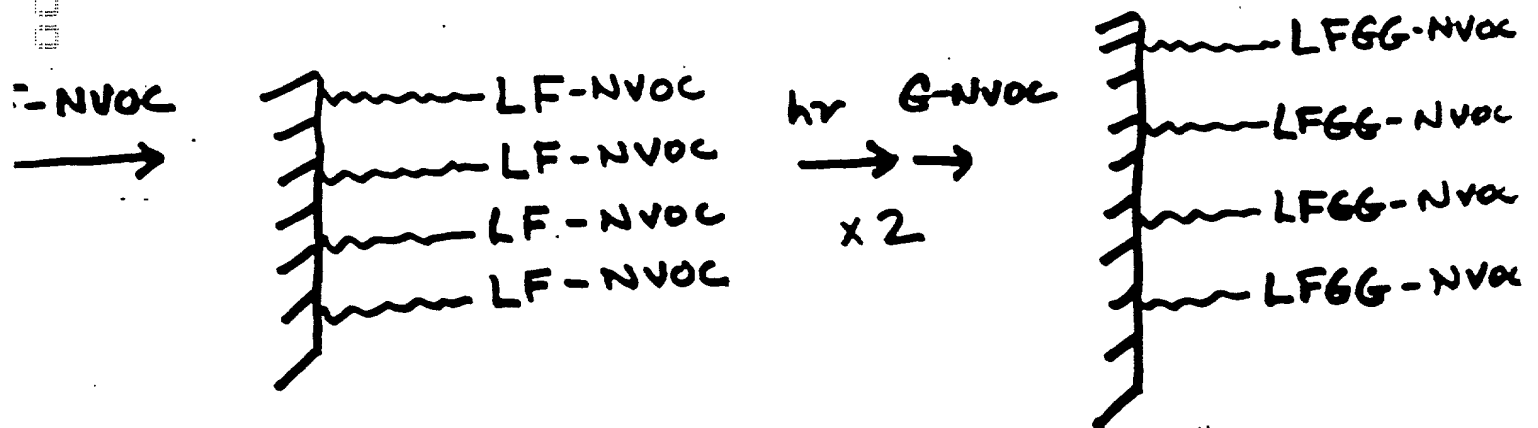
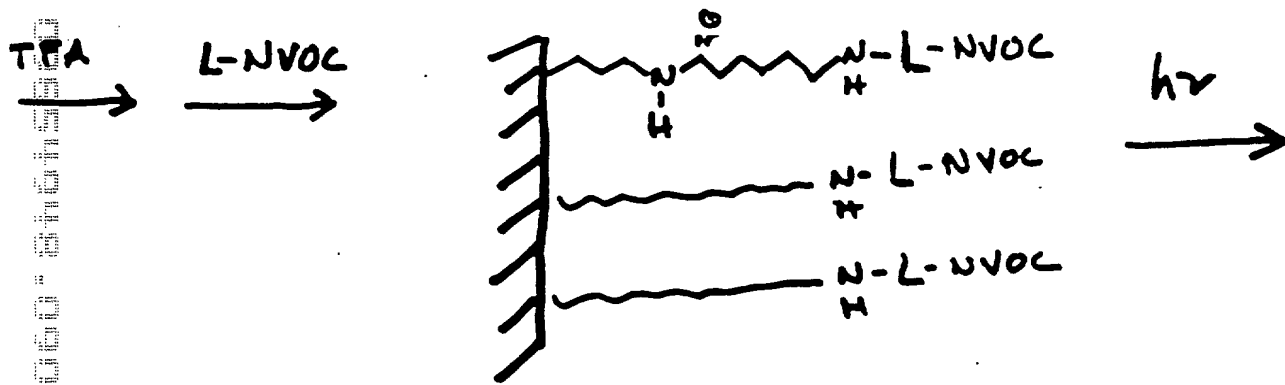
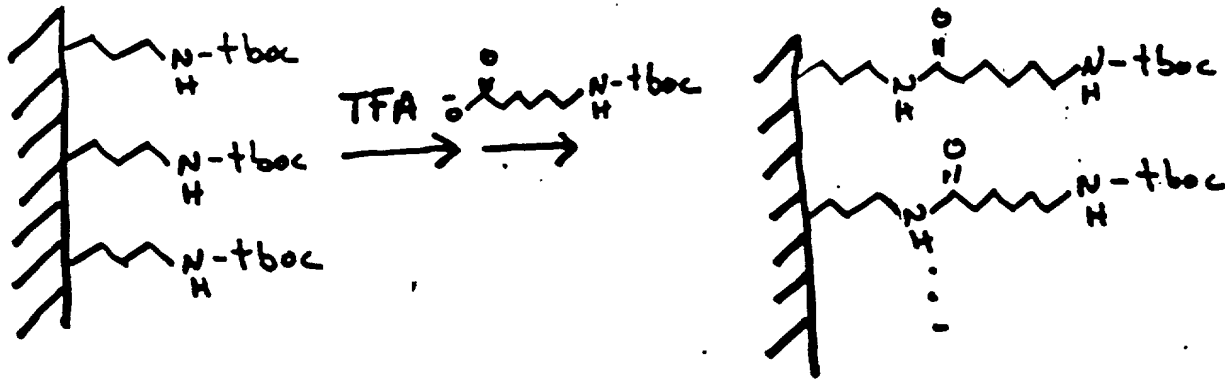


FIG. 19 A



NVOC GGFL

FIG. 19 B

$\downarrow \cdot h v$

A hand-drawn diagram of a 3x3 grid. The grid is formed by three horizontal and three vertical lines. The cells are shaded in a checkerboard pattern: the top-left, top-right, and bottom-middle cells are shaded with diagonal lines from top-left to bottom-right, while the other cells are unshaded.


$$H_2NGGFL$$

↓ NVOEY, hr

H₂N/GGFZ

↓ H E K Z

↓ GOAT ANTI-MOUSE-F1

FI  $\text{H}_2\text{NY}_6\text{GF}_2$

H2NY66FZ

FIG. 19 C

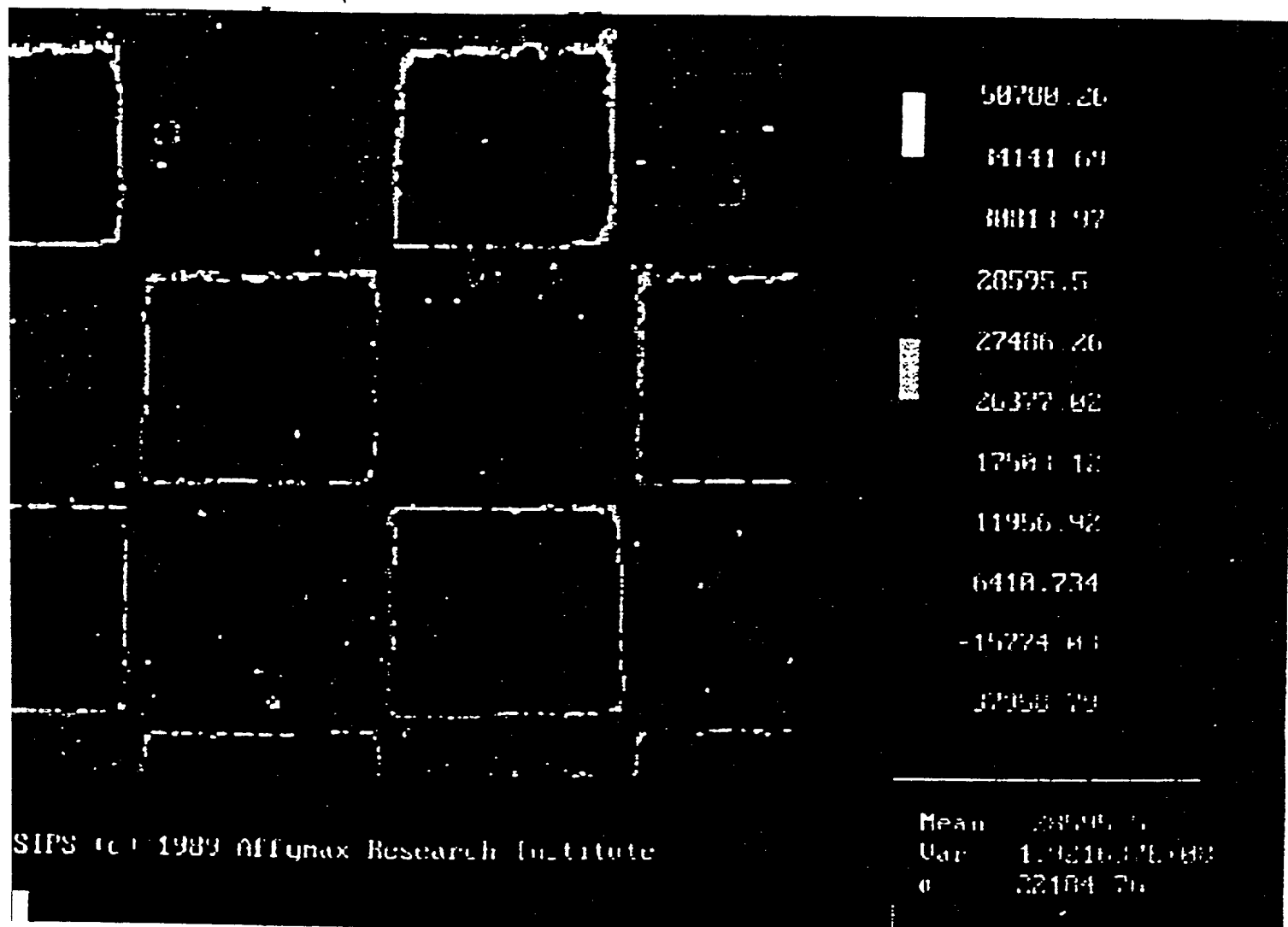


FIG. 19 D

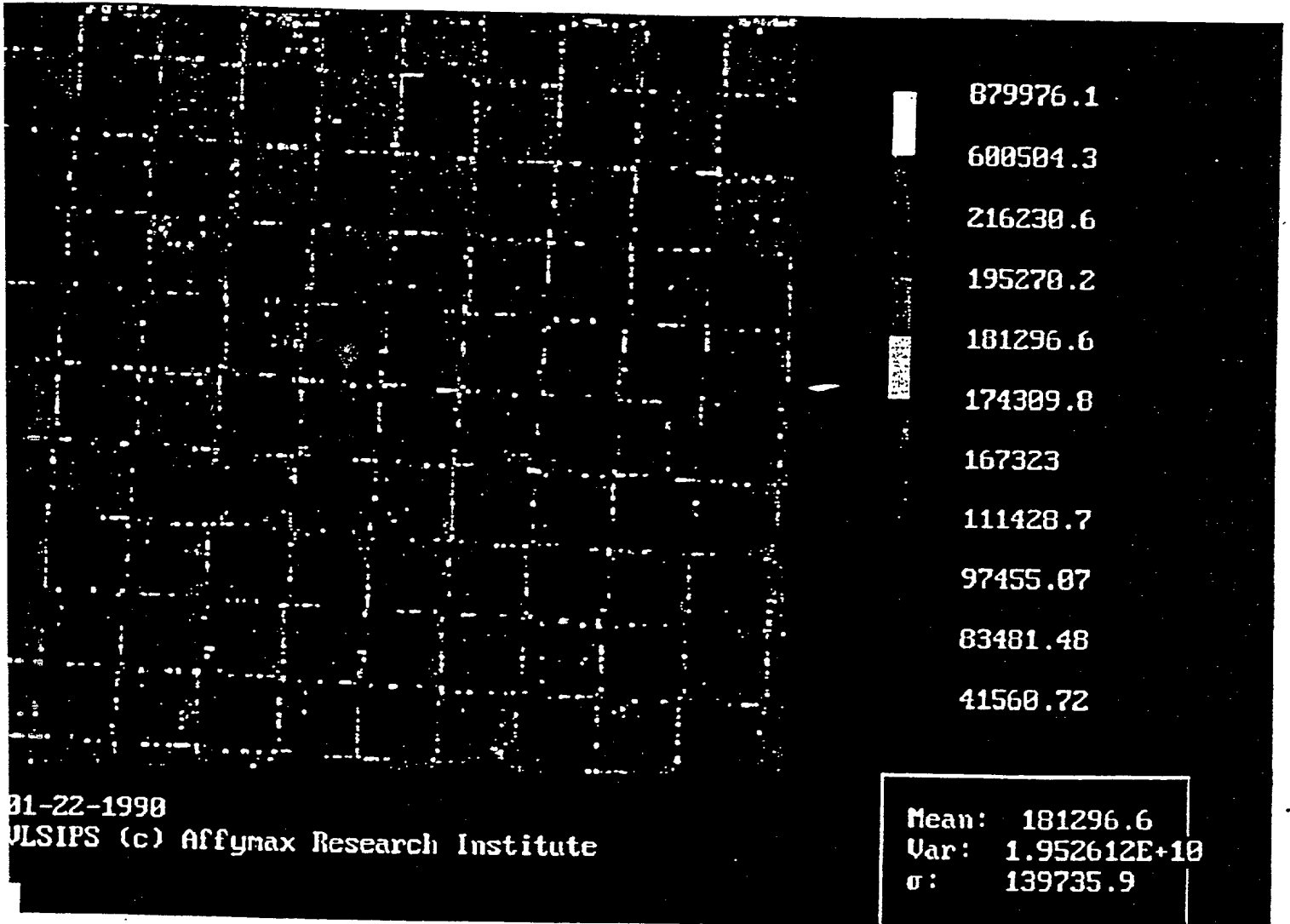


FIG. 20

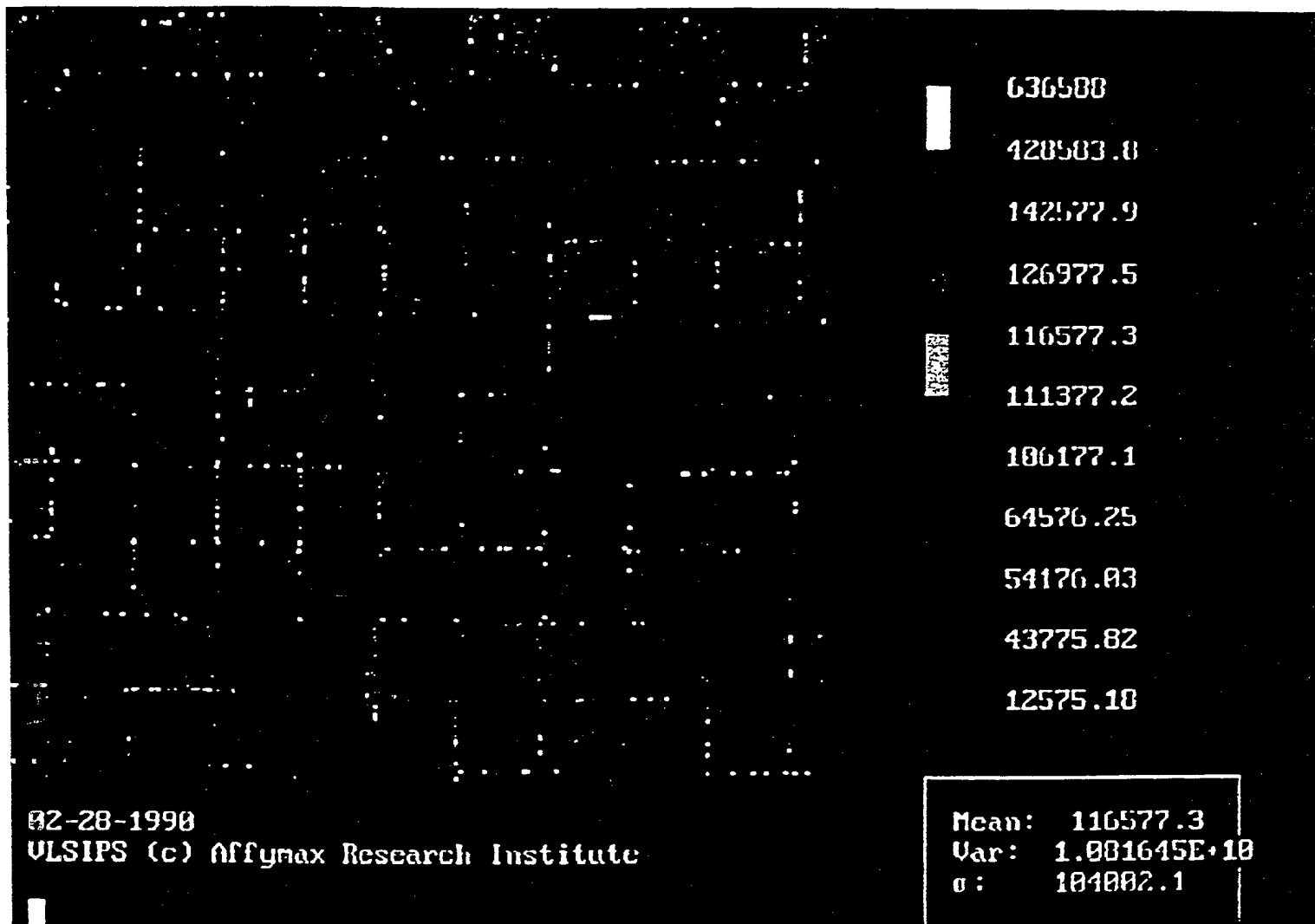
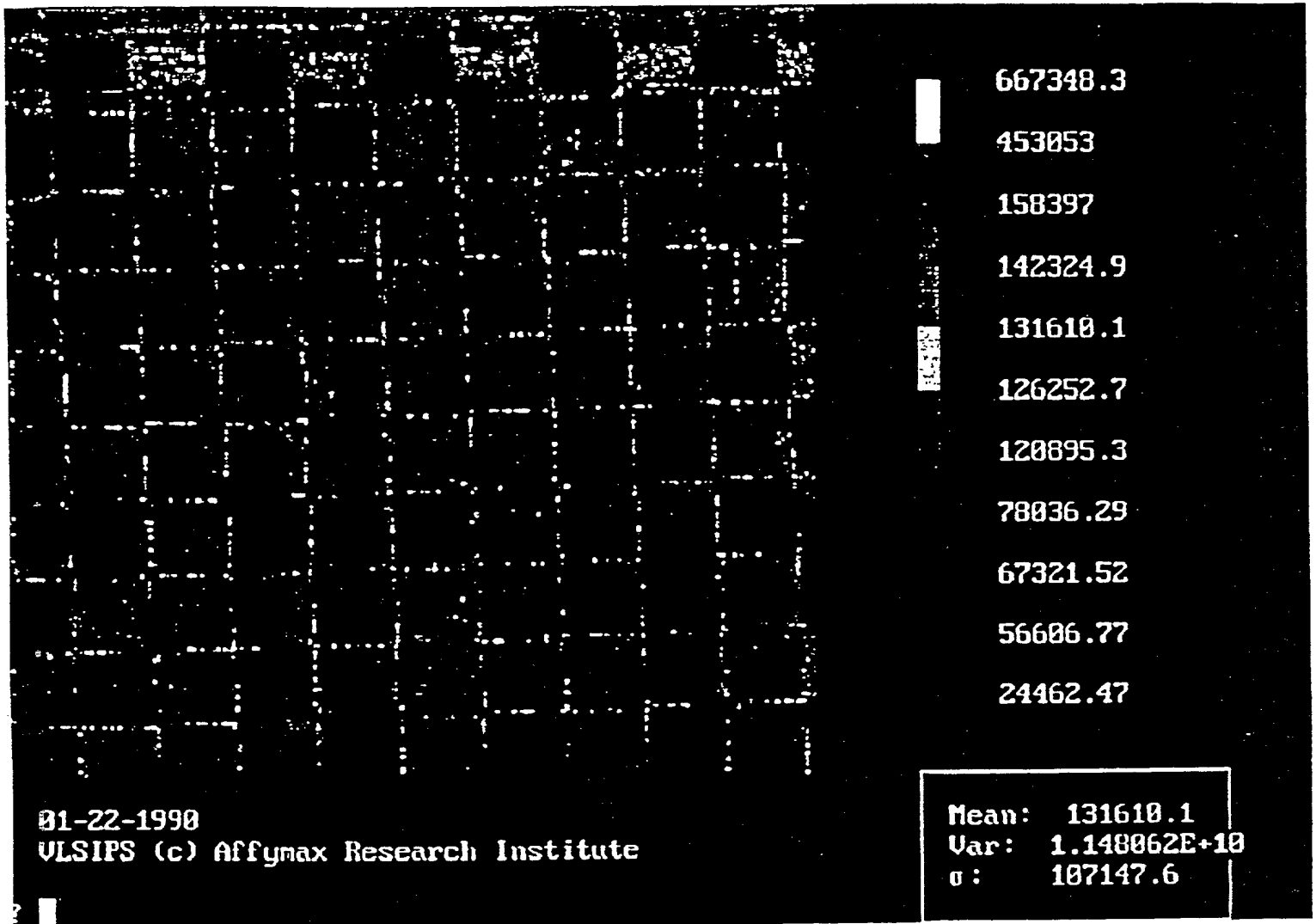


FIG. 21



P A S G

<u>L</u> P GFL	<u>L</u> A GFL	<u>L</u> S GFL	<u>L</u> G GFL
<u>F</u> P GFL	<u>F</u> A GFL	<u>F</u> S GFL	<u>F</u> G GFL
<u>W</u> P GFL	<u>W</u> A GFL	<u>W</u> S GFL	<u>W</u> G GFL
<u>Y</u> P GFL	<u>Y</u> A GFL	<u>Y</u> S GFL	<u>Y</u> G GFL

L

F

L set

W

Y

FIG. 22 A

P a A G

<u>Y</u> p GFL	<u>Y</u> a GFL	<u>Y</u> A GFL	<u>Y</u> G GFL
<u>f</u> p GFL	<u>f</u> a GFL	<u>f</u> A GFL	<u>f</u> G GFL
<u>w</u> p GFL	<u>w</u> a GFL	<u>w</u> A GFL	<u>w</u> G GFL
<u>y</u> p GFL	<u>y</u> a GFL	<u>y</u> A GFL	<u>y</u> G GFL

Y

F

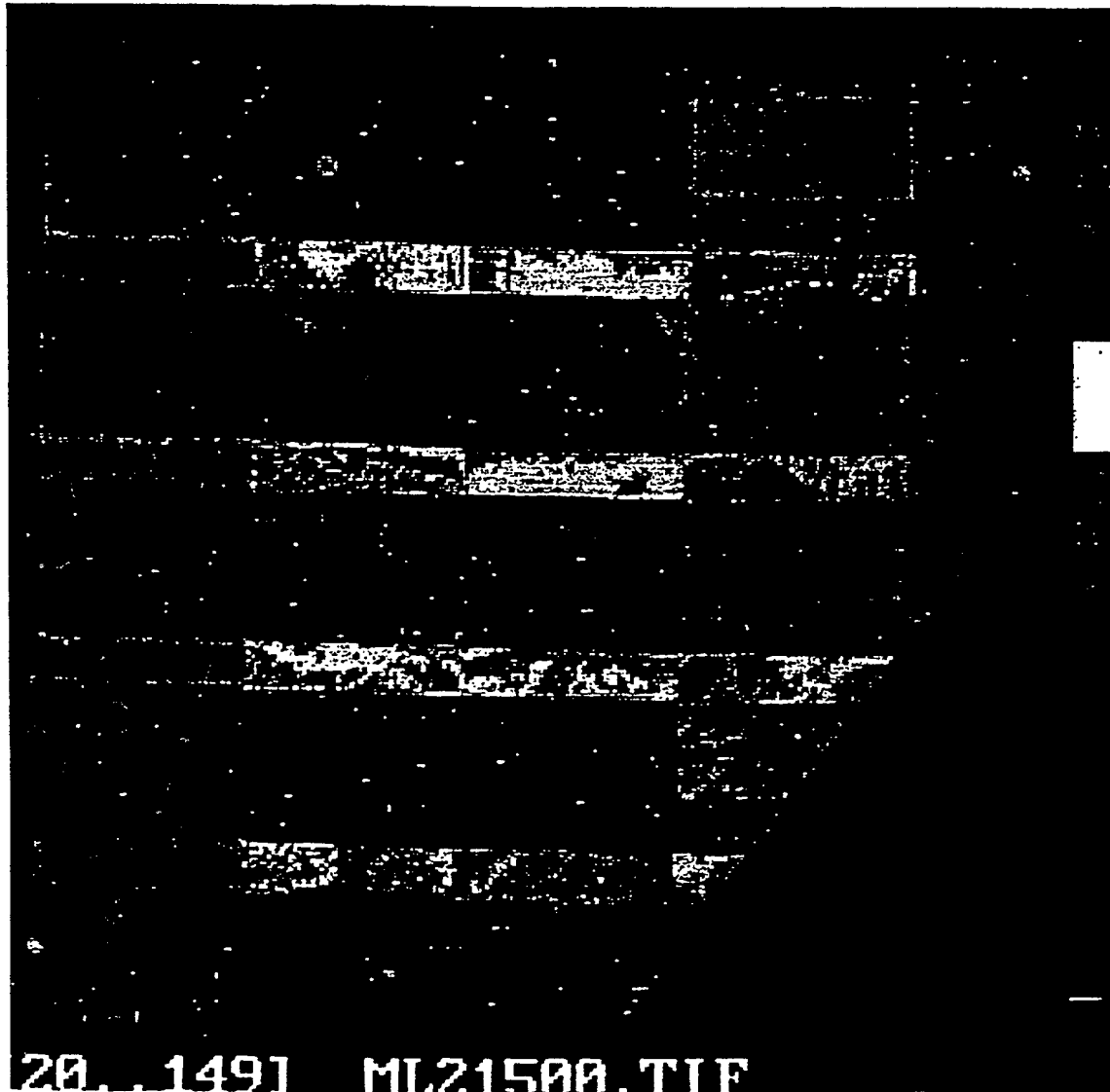
D set

W

Y

FIG. 22 B

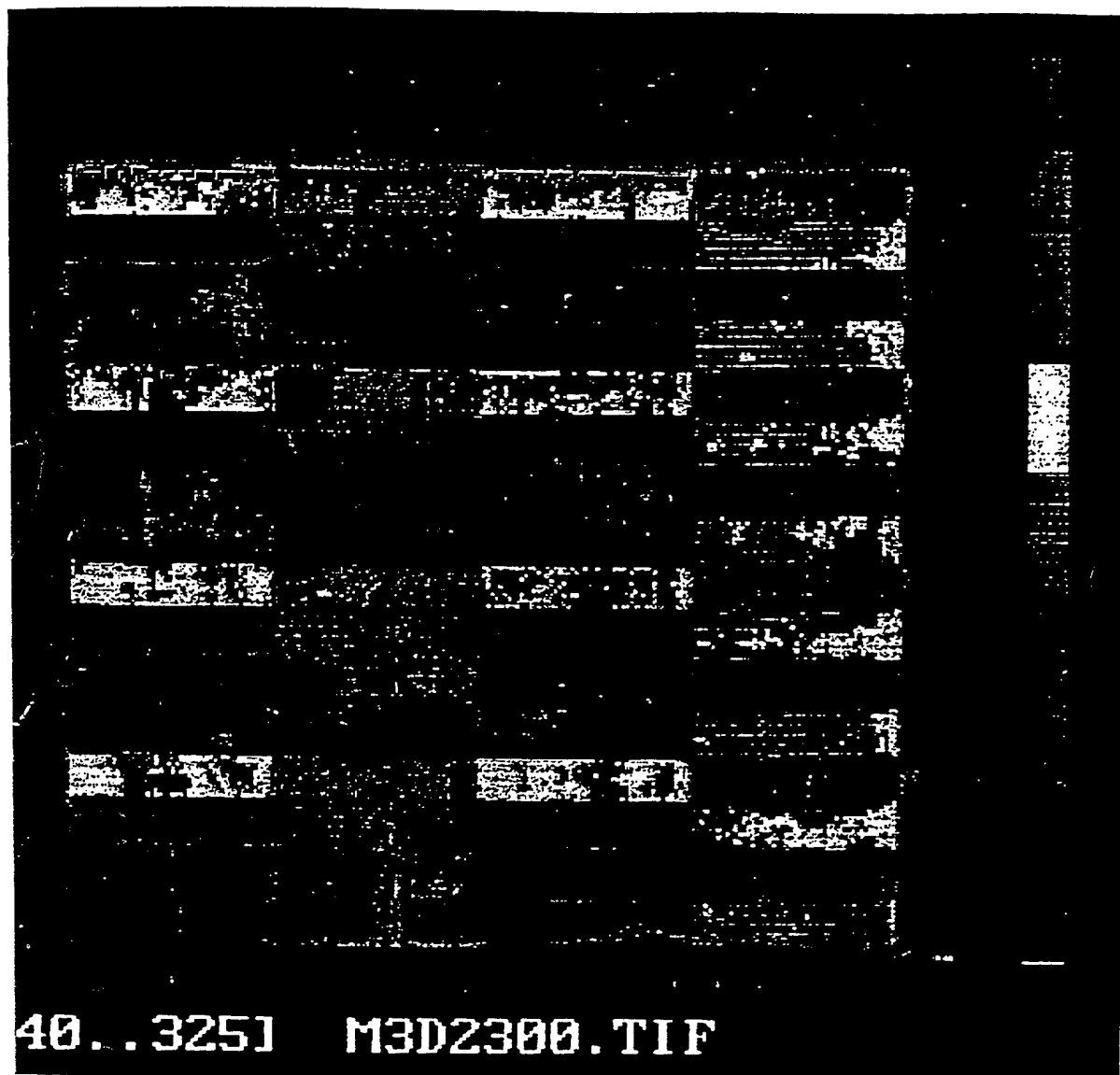
FIG. 23



149,000

20,000

FIG. 24



325,000

40,000

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: SUPPORT BOUND PROBES AND METHODS OF ANALYSIS USING THE SAME the specification of which was filed on April 24, 2000 as Application No. 09/557,875.

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56. I claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Country	Application No.	Date of Filing	Priority Claimed Under 35 USC 119

I hereby claim the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below:

Application No.	Filing Date

I claim the benefit under Title 35, United States Code, Section 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, Section 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, Section 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application No.	Date of Filing	Status
09/056,927	April 8, 1998	
08/670,118	June 25, 1996	
08/168,904	December 15, 1993	
07/624,114	December 6, 1990	
07/362,901	June 7, 1989	
08/348,471	November 30, 1994	
07/805,727	December 6, 1991	
07/492,462	March 7, 1990	

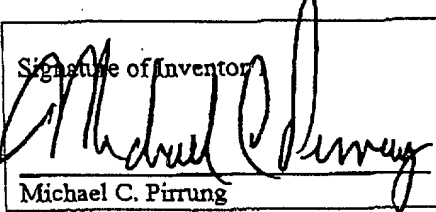
POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

Joe Liebeschuetz, Reg. No. 37,505
 William M. Smith, Reg. No. 30,223
 Rosemary Celli, Reg. No. 42,397
 Ted Apple, Reg. No. 36,429
 Philip L. McGarrigle, Reg. No. 31,395
 Wei Zhou, Reg. No. 44,419
 Ellen Gonzales, Reg. No. 44,128
 Vern Norviel, Reg. No. 32,483

Send Correspondence to: Joe Liebeschuetz TOWNSEND and TOWNSEND and CREW LLP Two Embarcadero Center, 8th Floor San Francisco, California 94111-3834	Direct Telephone Calls to: (Name, Reg. No., Telephone No.) Name: Joe Liebeschuetz Reg. No.: 37,505 Telephone: 650-326-2400
-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	-------------------------------------------------------------------------------------------------------------------------------------------------------------

Full Name of Inventor 1:	Last Name: PIRRUNG	First Name: MICHAEL	Middle Name or Initial: C.	
Residence & Citizenship:	City: Chapel Hill	State/Foreign Country: North Carolina	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 102 Whistlingtree Court	City: Chapel Hill	State/Country: NC	Postal Code: 27515
Full Name of Inventor 2:	Last Name: STRYER	First Name: LUBERT	Middle Name or Initial:	
Residence & Citizenship:	City: Stanford	State/Foreign Country: California	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 843 Sonoma Terrace	City: Stanford	State/Country: CA	Postal Code: 94305
Full Name of Inventor 3:	Last Name: FODOR	First Name: STEPHEN	Middle Name or Initial: P.A.	
Residence & Citizenship:	City: Palo Alto	State/Foreign Country: California	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 1120 Parkinson	City: Palo Alto	State/Country: CA	Postal Code: 94301
Full Name of Inventor 4:	Last Name: READ	First Name: J.	Middle Name or Initial: Leighton	
Residence & Citizenship:	City: Palo Alto	State/Foreign Country: California	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 1001 Ramona Avenue	City: Palo Alto	State/Country: CA	Postal Code: 94301

I further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of Inventor 1  Michael C. Pirrung	Signature of Inventor 2 Lubert Stryer	Signature of Inventor 3 Stephen P.A. Fodor
Date 6/13/2020	Date	Date
	Signature of Inventor 4 J. Leighton Read	
	Date	

PA 3063445 v1

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: SUPPORT BOUND PROBES AND METHODS OF ANALYSIS USING THE SAME the specification of which was filed on April 24, 2000 as Application No. 09/557,875.

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08/348,471	November 30, 1994	
07/805,727	December 6, 1991	
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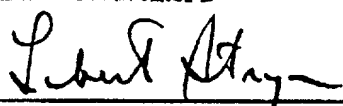
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 Wei Zhou, Reg. No. 44,419
 Ellen Gonzales, Reg. No. 44,128
 Vern Norviel, Reg. No. 32,483

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-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	-------------------------------------------------------------------------------------------------------------------------------------------------------------

Full Name of Inventor 1:	Last Name: PIRRUNG	First Name: MICHAEL	Middle Name or Initial: C.	
Residence & Citizenship:	City: Chapel Hill	State/Foreign Country: North Carolina	Country of Citizenship: USA	
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Residence & Citizenship:	City: Stanford	State/Foreign Country: California	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 843 Sonoma Terrace	City: Stanford	State/Country: CA	Postal Code: 94305
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Residence & Citizenship:	City: Palo Alto	State/Foreign Country: California	Country of Citizenship: USA	
Post Office Address:	Post Office Address: 1120 Parkinson	City: Palo Alto	State/Country: CA	Postal Code: 94301
Full Name of Inventor 4:	Last Name: READ	First Name: J.	Middle Name or Initial: Leighton	
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Signature of Inventor 1 _____ Michael C. Pirrung	Signature of Inventor 2  _____ Lubert Stryer	Signature of Inventor 3 _____ Stephen P.A. Fodor
Date	Date June 4, 2000	Date
	Signature of Inventor 4 _____ J. Leighton Read	
	Date	

PA 3063448 v1

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: SUPPORT BOUND PROBES AND METHODS OF ANALYSIS USING THE SAME the specification of which was filed on April 24, 2000 as Application No. 09/557,875.

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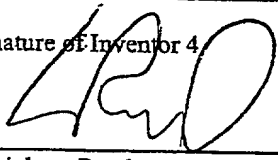
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 Philip L. McGarrigle, Reg. No. 31,395
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Signature of Inventor 1	Signature of Inventor 2	Signature of Inventor 3
Michael C. Pirrung	Lubert Stryer	Stephen P.A. Fodor
Date	Date	Date

Signature of Inventor 4

J. Leighton Read
Date June 20, 2000

PA 3063445 v1

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As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: SUPPORT BOUND PROBES AND METHODS OF ANALYSIS USING THE SAME the specification of which X is attached hereto or _____ was filed on _____ as Application No. _____ and was amended on _____ (if applicable).

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56. I claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Country	Application No.	Date of Filing	Priority Claimed Under 35 USC 119

I hereby claim the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below:

Application No.	Filing Date

I claim the benefit under Title 35, United States Code, Section 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, Section 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, Section 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application No.	Date of Filing	Status
09/056,927	April 8, 1998	
08/670,118	June 25, 1996	
08/168,904	December 15, 1993	
07/624,114	December 6, 1990	
07/362,901	June 7, 1989	
08/348,471	November 30, 1994	
07/805,727	December 6, 1991	
07/492,462	March 7, 1990	

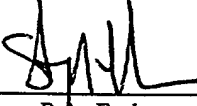
POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

Joe Liebeschuetz, Reg. No. 37,505
 William M. Smith, Reg. No. 30,223
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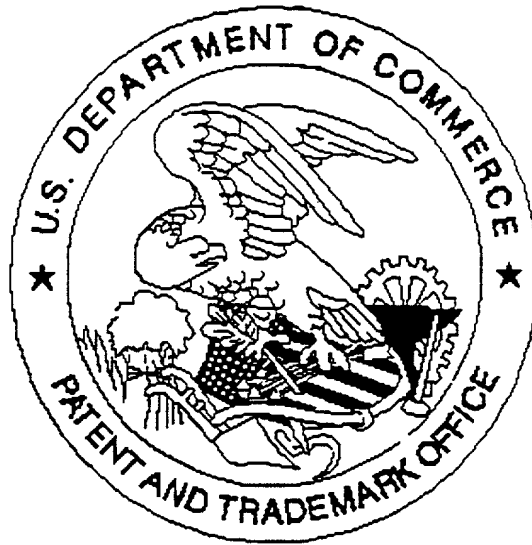
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I further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of Inventor 1  _____ Stephen P. A. Fodor Date AUG 14 2000	Signature of Inventor 2 _____ Lubert Stryer Date	Signature of Inventor 3 _____ Michael C. Pirrung Date
Signature of Inventor 4 _____ J. Leighton Read Date		

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Drawings